

Therapeutic potential of cannabis for surgical wound healing in rats

GOKHAN DOGUKAN AKARSU^{1,2*}, RUKIYE HOBEEK AKARSU³

¹Laboratory for Advanced Genomics, Division of Molecular Medicine,
Institute of Ruder Boskovic, Zagreb, Croatia

²Pharmacy Services, Vocational School of Health Services, Yozgat Bozok University,
Yozgat, Turkiye

³Department of Nursing, Faculty of Health Services, Yozgat Bozok University,
Yozgat, Turkiye

*Corresponding author: gokhan_dogukan_akarsu@hotmail.com

Citation: Akarsu GD, Hobek Akarsu R (2024): Therapeutic potential of cannabis for surgical wound healing in rats. Vet Med-Czech 69, 297–306.

Abstract: This study was conducted to evaluate the wound-healing activities of a *Cannabis sativa* L. plant extract and cannabidiol on incision wounds. An incision was created and sutured in rats under anaesthesia. Routine wound care procedures were applied for 10 days, followed by histological wound examinations. The cellular bioactivities of the hemp extract and CBD were assessed for MCP-1, EGF, BFGF, IL-8, and COL-1 using ELISA on the rat skin wound healing activity. A one-way ANOVA was used for the data analysis. The EGF values in the plasma were similar in the povidone-iodine, hemp seed oil, and hemp essential oil groups ($P > 0.05$). However, the EGF levels were lower in the CBD group compared to the other groups ($P < 0.001$, $P < 0.005$). The MCP-1 values in the hemp seed oil, hemp essential oil, and CBD were similar ($P > 0.05$), whereas povidone iodine exhibited lower MCP-1 levels compared to the other groups ($P < 0.001$, $P < 0.005$). It was determined that the plasma BFGF, IL-8, and COL 1 values of the groups were similar ($P > 0.05$). To our knowledge, this study is the first to evaluate the effects of CBD, seed oil, and hemp leaf extract on incision wound healing. It demonstrates that hemp extract holds greater potential benefits for wound healing compared to CBD.

Keywords: cannabinoids; cannabis sativa; essential oil; hemp; treatment; wounds and injury

Wound healing is a complex biological process divided into four overlapping stages: haemostasis, inflammation, proliferation, and remodelling (Abbas et al. 2019). Depending on the depth and severity of the wound's thickness, wound healing takes several months (6–9 weeks). Due to intense inflammation and a higher rate of infection (bac-

terial proliferation), non-healing wounds pose a challenging approach for wound care specialists worldwide, especially in an ageing population, and in patients suffering from diabetes and obesity. Classical or traditional wound therapy usually uses gauze dressings, which do not have a therapeutic effect. They are inexpensive, but the outcome of the

Funded by Yozgat Bozok University Project Coordination Implementation and Research Institute (Grant No.: TKA-2022-901). The APC was funded by authors.

© The authors. This work is licensed under a Creative Commons Attribution-NonCommercial 4.0 International (CC BY-NC 4.0).

wound treatment with traditional dressings is often uncertain, and the process could be economically inefficient. Non-toxic and non-allergenic dressing materials are necessary, which allow gas exchange, reduce inflammation by creating a barrier against microbial agents, and support biological healing processes by preventing infection (Xu et al. 2015). Various wound dressing products have been developed using synthetic polymers to expedite wound healing (Hou et al. 2020). Synthetic polymers can potentially cause skin damage or lead to infection due to their structures. However, there is insufficient evidence to demonstrate the superiority of wound dressing products over synthetic polymers or traditional dressing methods (Mirasoglu 2015). As a result, research on naturally derived products with antibacterial and pro-collagen synthesis properties is continuing to advance to achieve optimal outcomes in wound healing. Numerous studies suggest that cannabinoids, due to their anti-inflammatory and antioxidant effects, might offer a novel approach to wound healing by alleviating inflammation and supporting the healing process (Thapa et al. 2020; Kibret et al. 2022; Kongkadee et al. 2022; Spinella et al. 2023).

Cannabinoids, biologically active compounds derived from *Cannabis sativa* (phytocannabinoids), have garnered attention for their anti-inflammatory and antioxidant effects, suggesting a novel approach to wound healing by alleviating inflammation and supporting the healing process (Passani et al. 2020). Despite the ancient recognition of the biological and pharmacological properties of *Cannabis sativa* in popular medicine, its pharmacological use has been restricted due to concerns about misuse related to the presence of tetrahydrocannabinol THC (Andre et al. 2016).

In modern times, the sativa subspecies is legally cultivated in many countries, not only for its seeds, an excellent source of lipids, proteins, carbohydrates, minerals, and vitamins, but also for the production of “Cannabidiol = CBD” (non-psychoactive cannabinoid), which is of significant importance (Ferrini et al. 2021). The existing literature primarily focuses on CBD, overlooking the biologically valuable molecules present in both the seeds and leaves of *Cannabis sativa*, which hold high nutritional value (Callaway 2004; Wang et al. 2008; Da Porto et al. 2012; Teh and Birch 2013; Pojic et al. 2014; Liang et al. 2015; Schluttenhofer and Yuan 2017; Mikulec et al. 2019; Rapa et al. 2019).

Biochemical markers used to determine wound healing:

- Epidermal growth factor (EGF): It plays an important role in the regulation of cell growth, proliferation and differentiation (Gurtner and Wong 2006).
- Monocyte chemoattractant protein-1 (MCP-1/MCAF): It has been shown that the MCP-1 protein increases in the wound area. It has been reported that it regulates keratinocyte migration (Gurtner and Wong 2006).
- Basic fibroblast growth factors (BFGFs): By stimulating angiogenesis and the proliferation of fibroblasts, they form the granulation tissue that fills the wound cavity in the early phase of wound healing (Barrientos et al. 2008).
- Interleukin-8 (IL-8): The IL-8 level has been shown to increase in the dermis of acute surgical wounds (Metcalf and Ferguson 2007; Raja et al. 2007).
- Collagen-1 (COL-1): COL-1 has a central role as the main structural component of the dermis in the remodelling of cutaneous tissue defects (Metcalf and Ferguson 2007; Raja et al. 2007).

However, there is no study in the literature evaluating the superiority of both the seed oil and leaf extract over pure CBD or over routinely used dressing products. Therefore, the aim of this study is to evaluate the effectiveness of hemp essential oils, hemp seeds, and CBD against 10% povidone-iodine, routinely used in surgical operations for abdominal sutured incisions in rats.

MATERIAL AND METHODS

Ethical statement

The animal study protocol was approved by the Erciyes University Animal Experiments Ethics Committee (04/21/79).

Chemicals

“Dimethyl sulfoxide = DMSO” (purity > 99.5%) and ethanol (purity > 99.5%) from Sigma Aldrich (Massachusetts, USA), CBD (purity > 95%) from Yozgat Bozok University Hemp Research Institute (Yozgat, Türkiye), Primavilin from VILSAN

<https://doi.org/10.17221/21/2024-VETMED>

Veterinary Medicines Trade and Industry Company (Ankara, Türkiye), Ketolin from Pİ Farma İlaç Industry and Trade Company (Ankara, Türkiye), Xylazine (Bioveta PLC, Ivanovice na Hane, Czech Republic), Ketamine (Richter Pharma AG, Wels, Austria) were obtained and 10% povidone iodine was purchased from Alfa Medical Health Products Company (Istanbul, Türkiye).

Study design

CANNABIS SATIVA PROCUREMENT

The hemp was obtained from Yozgat Bozok University, which has a “Hemp Research Institute” and was authorised with special permission. All phases of the research were carried out within legal responsibilities and limitations.

PREPARATION OF THE CBD

The CBD was obtained from the Hemp Research Institute of Yozgat Bozok University. The CBD prepared by the institute is 98% pure. CBD was stored at 4 °C until the CBD solution was prepared. To prepare the solution, it was brought to room temperature and 96% ethanol was added. The resulting mixture was vortexed to obtain a solution containing 15% CBD by volume. The prepared solution was transferred to a spray can and wrapped with aluminium foil to prevent exposure to light.

PREPARATION OF THE SEED OIL

Hemp seed oil was obtained from hemp seeds by cold pressing. The obtained oil was stored at 4 °C until required during the experiment. It was wrapped in aluminium foil to avoid exposure to light.

PREPARATION OF THE LEAF EXTRACT

The process steps for obtaining the essential oil of hemp are as follows: The green parts of the hemp were harvested in season. Since drying in the oven may damage the essential oils, it was allowed to dry at room temperature and in the shade. The dried hemp leaves were ground at 28 000 g for 40 s and

turned into a micron-sized powder. Four hundred grams (400 g) of powdered hemp leaves were transferred to a 5-litre balloon jug. Three (3) litres of distilled water was added to it. The heater was placed in the mantle and allowed to boil for 210 min after it initially boiled. The essential oils were concentrated by water cooling and the essential oil was obtained with a separating funnel. The obtained essential oils were placed in Eppendorf tubes, covered with paraffin and stored at –20 °C. Eppendorf tubes were brought to room temperature just before the surgical operation. The oil was placed in a capped tube and a 15% suspension was created with DMSO. The oil was applied to the suture area after the surgical operation. The stock tubes containing the hemp essential oil were wrapped in aluminium foil to prevent exposure to light. DMSO was used as a solvent for the essential oils. DMSO was preferred because it can pass through biological membranes and has no toxicity at low concentrations (Majno 1975; Scharffetter et al. 1989; Liska et al. 1994).

SUPPLY AND PREPARATION OF EXPERIMENTAL ANIMALS

The experimental animals were obtained from Erciyes University Experimental Research Application and Research Center. The rats received for experimentation were checked by the veterinarian for their health status, sex and weight. Female rats were not included in the experiment due to their menstrual cycle and hormonal balance. The weight of each rat used in the experiment ranged from 190 to 253 grams. Wistar Albino male rats were used in the study. The received rats were kept in the study room for 7 days without any treatment so that they got accustomed to it. The rats were randomly divided into 4 groups of 12 animals. During the experiment, a 12/12-hour night/day period was created and the rats were given standard rat pellet food and tap water *ad libitum*. The room temperature was maintained between 20.0 and 21.4 °C.

CREATION OF THE INCISION MODEL

During the night, before the working day, the rats were not given food, but they continued to drink water. All the rats were anaesthetised with

8.00 mg/kg of xylazine (Bioveta PLC, Ivanovice na Hane, Czech Republic) and 60.00 mg/kg of ketamine (Richter Pharma AG, Wels, Austria). The abdominal region was sterilised. After sterilisation, the abdominal region was shaved with a disposable razor blade. A sterile surgical drape was covered, leaving the abdominal area open. The abdomen was sterilised again and the surgical operation was started. A 4 cm long cut of the skin was performed with a scalpel. The peritoneum was opened. The peritoneum was not damaged. This area was then closed with sutures. Each layer was sutured with a simple separate suture technique. The incision was closed with 4 sutures for each rat. The suture zone was 3–3.5 cm long and 1.6 cm wide.

Subsequently, the experimental animals were divided into four groups.

- Group 1: Povidone-iodine group.
- Group 2: Hemp seed oil group.
- Group 3: Hemp essential oil group.
- Group 4: CBD group.

After the surgical operation, the suture was closed with sterile gauze pads and the rats were placed in separate cages due to the possibility of damaging each other.

Wound care process

The 1st group, the povidone-iodine group: After the surgical operation, 10% povidone iodine was applied on the suture and on a circular area, 2 cm in diameter, around the suture area.

The 2nd group, the hemp seed oil group: After the surgical operation, hemp seed oil was applied on the suture and on a circular area with a diameter of 2 cm around the suture area.

The 3rd group, the hemp essential oil group: After the surgical operation, hemp essential oil was applied on the suture and on a circular area with a diameter of 2 cm around the suture area.

The 4th group, the CBD group: After the surgical operation, CBD was applied on the suture and on a circular area with a diameter of 2 cm around the suture area.

After each group's own solution was applied, the sutures of the rats were closed with sterile gauze. The rats were given one dose of antibiotics and one dose of painkillers according to their weight. Each rat was placed in a separate cage in case the rats

harmed each other. The wound care product applied after the surgery was applied in the morning every day for 10 days. The cages, feed and water were changed daily due to the possibility of infection.

Evaluation of the wound tissue: The day when the incision was made was considered Day 0, and the suture area of all the rats was photographed every day. The wound was evaluated histologically on the 1st, 2nd, 3rd, 10th and 21st days. The photographs were evaluated by a histologist, independent from the study.

Preparation of plasma

Plasma was directly collected from the heart and carefully transferred to centrifuge tubes to conduct analysis while avoiding haemolysis. It was then centrifuged at 3 000 *g* for 30 minutes. The supernatants were placed into Eppendorf tubes with transfer pipettes and were stored at –20 °C until analysis.

Biochemical analysis

Enzyme-linked immunosorbent assay (ELISA) analyses were performed in the Erciyes University Medical Biochemistry laboratories (Kayseri, Türkiye). On the day of the analysis, the samples were removed from the –20 °C storage. In the samples taken, MCP-1, IL-8, BFGF, EGF and COL-1 were analysed by the ELISA method. The ELISA analysis was performed according to the kit's instructions.

Statistical analysis

The data were evaluated using the SPSS (v26; IBM, Chicago, USA) program. The descriptive data were given as numbers, percentages, the mean, standard deviation, minimum and maximum. The conformity of the data to the normal distribution was evaluated with the Shapiro-Wilk test. It was found that the data fit the normal distribution. A one way analysis of variance (ANOVA) was used in the analysis of the data. Post hoc, Bonferroni and Games-Howell tests were applied to determine which group caused the difference. The statistical significance level was accepted as $P < 0.05$.

<https://doi.org/10.17221/21/2024-VETMED>

RESULTS

Histological results

When the wound tissue was observed on the 1st day, no signs of infection, oedema and redness were observed on the wound. For the 1st day, stability was achieved in the wound tissue of the 2nd and 3rd group rats. Although the 1st and 4th groups were evaluated as good, it can be said that the wound tissue of the 2nd and 3rd groups was healthier.

When the wound tissue was evaluated on the 2nd day, it can be stated that the wound tissue of the 3rd group rats was better than the other groups, the wound tissue began to become evident, and no signs of active infection were observed in the groups.

When the wound tissue was evaluated on the 3rd day, it can be stated that the wound development process of the 2nd group is in a very good condition, the other groups are in similar conditions to the 2nd day and there was no sign of infection.

In the evaluation of the wound tissue on the 10th day, it was observed that the skin hairs were getting longer. It was not considered a distinctive feature because there were rats with or without skin hair in each group. It can be stated that the wound tissue healed completely in the Group 2 rats, there were no sutures (scars), the incision site was not visible to the naked eye, and similar findings occurred

in Group 3. On day 10, the incisions in Groups 1 and 4 had not healed completely. It was observed that a 0.75 cm non-healing incision line remained from the 4 cm incision. It can be stated that there was localised tissue stiffness and stability was not provided.

In the evaluation of the wound tissue on the 21st day, when the maturation phase was passed, it was observed that the incision line of the 1st, 2nd, and 3rd group rats became unclear, the skin hairs increased, the wound tissue disappeared and the wound was in the maturation phase. On the other hand, although the wound tissue of the 4th group rats was healed, it was thought that there was localised damage, and partial inflammation due to the wound tissue, it was thought that the wound tissue was damaged due to itching and redness as the rats were in a free state, and it can be stated that complete healing was not achieved.

The blood tissue analyses

The plasma EGF, MCP-1, BFGF, IL-8, COL-1 data of the rats obtained after the stages specified in the method section are given in Table 1 and Figures 1–5.

The plasma EGF values were similar in Group 1, Group 2 and Group 3 ($P > 0.05$), the EGF levels

Table 1. Comparison of the plasma parameters between the groups

Biochemical parameters	$\bar{x} \pm \text{SD (min.–max.)}$				Test value	P-value
	Group 1 (a)	Group 2 (b)	Group 3 (c)	Group 4 (d)		
EGF (ng/l)	43.95 $\pm$ 51.46 (1.55–105.97)	77.37 $\pm$ 71.95 (142–198.75)	53.15 $\pm$ 54.37 (0.85–134.85)	4.29 $\pm$ 2.34 (1.88–9.14)	3.489	0.025
b > d, c > d, a = b = c, a = d*						
MCP-1 (pg/ml)	16.38 $\pm$ 12.71 (1.05–40.16)	208.31 $\pm$ 75.96 (10.32–324.87)	229.12 $\pm$ 44.14 (165.66–328.58)	203.53 $\pm$ 74.89 (5.41–271.16)	20.939	0.000
b = c = d > a*						
BFGF (pg/ml)	56.82 $\pm$ 14.95 (39.41–83.55)	53.13 $\pm$ 14.84 (31.67–87.87)	43.89 $\pm$ 18.35 (0.22–74.90)	56.50 $\pm$ 11.96 (38.34–71.02)	1.647	0.195
IL-8 (ng/l)	47.66 $\pm$ 20.46 (2.45–63.18)	43.08 $\pm$ 6.29 (32.00–52.91)	41.53 $\pm$ 5.37 (32.33–48.71)	40.89 $\pm$ 6.83 (30.40–48.56)	0.743	0.533
COL-1 (ng/l)	336.58 $\pm$ 43.34 (307.67–400.31)	362.83 $\pm$ 44.72 (323.45–488.92)	341.06 $\pm$ 48.24 (276.42–410.14)	353.08 $\pm$ 26.17 (312.88–389.18)	0.673	0.575

*Statistical differences

BFGF = basic fibroblast growth factor; COL-1 = collagen-1; EGF = epidermal growth factor; IL-8 = interleukin-8; MCP-1 = monocyte chemoattractant protein-1

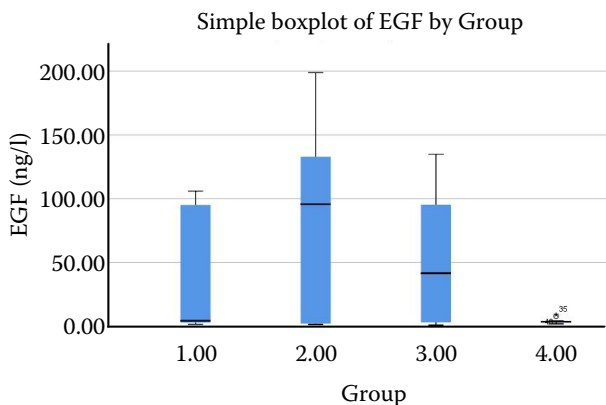


Figure 1. Graph of the plasma epidermal growth factor (EGF) levels

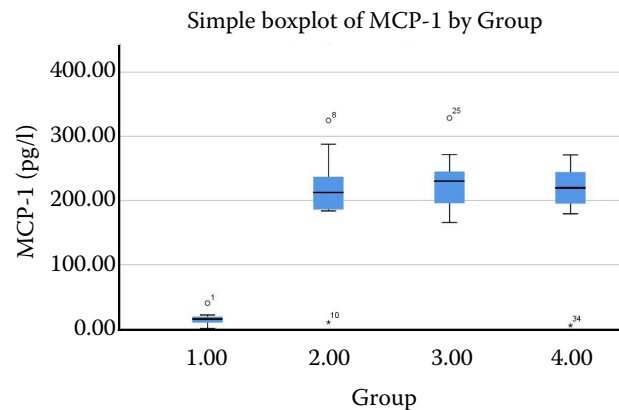


Figure 2. Graph of the plasma monocyte chemoattractant protein-1 (MCP-1) levels

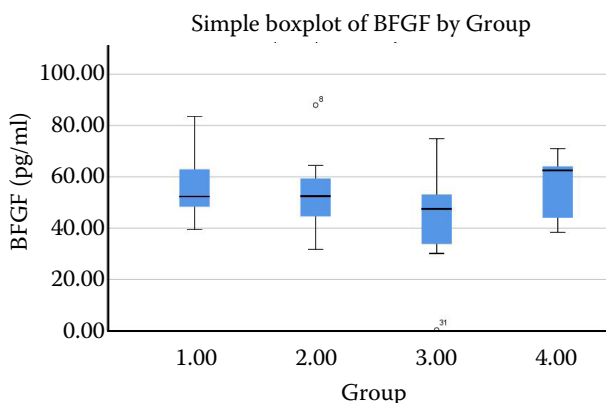


Figure 3. Graph of the plasma basic fibroblast growth factor (BFGF) levels

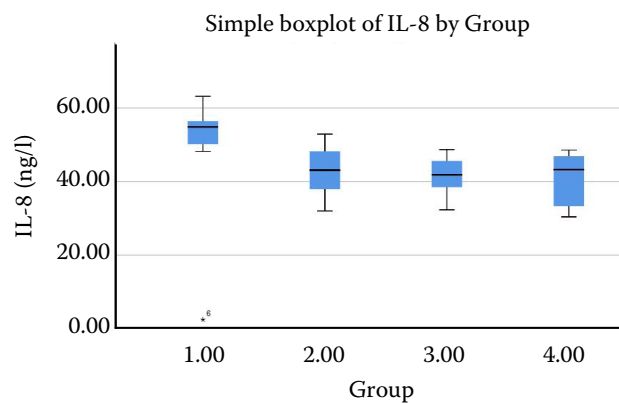


Figure 4. Graph of the plasma interleukin-8 (IL-8) levels

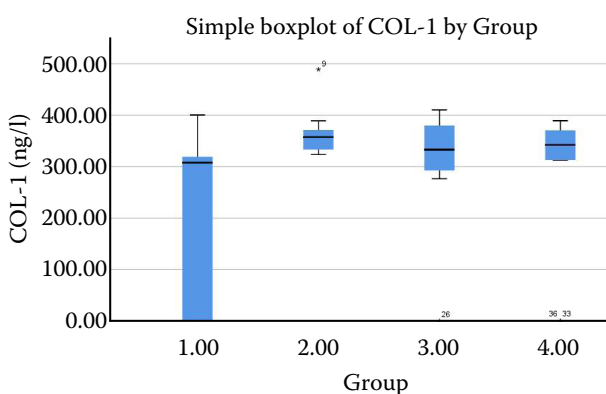


Figure 5. Graph of the plasma collagen-1 (COL-1) levels

were lower in Group 4 compared to the other groups ($P < 0.001$, $P < 0.005$), the MCP-1 values in Group 2, Group 3 and Group 4 were similar ($P > 0.05$), Group 1 had lower MCP-1 levels compared to the other groups ($P < 0.001$, $P < 0.005$), it was determined that the plasma BFGF, IL-8 and COL-1 values of the groups were similar ($P > 0.05$) (Table 1, Figures 1–5).

DISCUSSION

In contemporary medical practices, tissue in the post-operative period remains pivotal for patients to resume their normal lives promptly (Majno 1975; Scharffetter et al. 1989; Liska et al. 1994; Konca and Akbay 2000). This study aimed to assess the efficacy of hemp essential oils, hemp seeds, and CBD in abdominal sutured incisions in rats, concurrently evaluating their effectiveness against the routine wound care agent, povidone-iodine.

Wound healing involves a myriad of biochemical parameters (Nunan et al. 2014). Firstly, we examined the epidermal growth factor (EGF).

Previous studies highlighted the positive contribution of increased EGF levels to wound healing (Busanello-Costa et al. 2023; Kim et al. 2023; Yasti et al. 2023). In our study, the plasma EGF level in Group 2 was significantly higher. Although Group 3 exhibited a higher EGF level than Group 1, the difference was not statistically significant. This suggests that the hemp seed oil may enhance the

<https://doi.org/10.17221/21/2024-VETMED>

EGF levels. Notably, there is a gap in the literature regarding the impact of hemp extract on the EGF. However, a study investigating a standardised *Cannabis sativa* L. ethanolic extract containing cannabidiol demonstrated the potential effects on the vascular endothelial growth factor release (Kurtoglu et al. 2014).

Monocyte chemoattractant protein-1 (MCP-1) is another crucial parameter in wound tissue healing. Studies have reported that a decrease in the MCP-1 expression indicates that the wound tissue progresses toward healing (Aguayo-Morales et al. 2023; Chen et al. 2023; Zhang et al. 2023). In our study, it was determined that the 21st day plasma MCP-1 level of the group treated with the 10% povidone iodine solution was lower than the other groups and this was statistically significant. It can be stated that povidone-iodine solution provides a lower amount of MCP-1 expression than the products obtained from hemp. In another conducted study, it was determined that THC reduces the MCP-1 secretion (Sangiovanni et al. 2019). Another study found that hemp extracts suppress MCP-1 (Busanello-Costa et al. 2023). There is also a study demonstrating that a high-CBD extract reduces the MCP-1 levels in a lung inflammation mouse model (Aguayo-Morales et al. 2023). In our study, contrary to the literature, it was found that povidone-iodine suppresses MCP-1 to a greater extent. Moreover, CBD without THC and hemp essential oils containing trace amounts of THC exhibited the same effect as the hemp seed. Therefore, it can be stated that the effect on MCP-1 is not solely attributed to THC, but also to the presence of both THC and CBD together, or solely to CBD (Aguayo-Morales et al. 2023).

Yang et al. (2014) and Ebrahimpour-Malekshah et al. (2023) showed, in their studies, that increased BFGF levels indicate the rapid healing of the wound tissue. In our study, it was determined that there was no statistically significant difference between the BFGF levels between the groups and that all the groups were similar to each other. No study evaluating the direct effects of hemp extracts on BFGF was found in the literature. However, in our study, materials containing *Cannabis sativa* showed a significant increase in cell proliferation compared to collagen hydrogels containing silver nanoparticles. Our study findings are supportive of the literature. It can be stated that CBD and hemp extracts increase BFGF at least as much as povidone-iodine (Chen et al. 2023).

Rizzo et al. (2023), Alyami et al. (2023) and Jayabal et al. (2023), in their studies, reported that the wound tissue increased the IL-8 expression. In our study, on the other hand, it was determined that there was a similar IL-8 expression without any statistical difference between the groups. None of the hemp products that we used for the experimental groups were different from the 10% povidone-iodine solution for the IL-8 levels. In a study utilising a standardised *Cannabis sativa* ethanolic extract (CSE) containing 5% CBD and a low THC concentration, it was observed that both CSE and CBD inhibited the nuclear transcription factor κ B (NF- κ B). However, only CSE exhibited a decrease in the interleukin-8 (IL-8) secretion. Another study reported that an extract fraction derived from the *C. sativa* Arbel strain (F CBD) significantly reduced the IL-6 and IL-8 levels in an alveolar epithelial (A549) cell line, dose-dependently (Konca and Akbay 2000; Fumagalli et al. 2022).

In our study, hemp essential oils containing CBD and low levels of THC, as well as hemp seeds, were found to suppress the IL-8 factor. These substances exhibited similar effects to each other and were comparable to the routine wound care material povidone-iodine. In the literature, no study was encountered evaluating the superiority of CBD and THC over each other or over any commonly used material (wound care product) in terms of the IL-8 factor. Therefore, it can be stated that CBD and hemp extracts suppress IL-8 at least as effectively as povidone-iodine.

Collagen contributes to the mechanical strength and flexibility of tissues and serves as a natural substrate for cellular adhesion, proliferation, and differentiation. Narisepalli et al. (2023), Hu et al. (2022) and Gong et al. (2022) reported, in their studies, that an increased COL-1 expression showed that wound healing was in a normal state.

In our study, none of the hemp products used for the experimental groups was different from the 10% povidone-iodine solution. It provided a similar COL-1 expression.

No study evaluating the direct effects of hemp products on COL-1 was found in the literature. However, in a study assessing the superiority of pure CBD over a routine care product in a wound opened on mice tongues, it was noted that the groups treated with CBD exhibited well-organised accumulated collagen fibres, newly formed blood vessels, and fibroplasia. The marginal epithe-

lium was also reported to be thicker (Bellocchio et al. 2023).

In our study, histologically, groups treated with CBD and hemp extracts for wound care exhibited well-organised accumulated collagen fibres, newly formed blood vessels, and fibroplasia on the 10th day. On the 21st day, the CBD, hemp seed oil, and hemp essential oil groups showed complete wound tissue formation, while the povidone-iodine group still displayed local damage in mice. Moreover, no signs of infection were observed in the wounds. In another study, hemp extracts were indicated as a valuable source of biologically active substances that can reduce oxidative stress and positively influence the vitality of skin cells (Zagorska-Dizok et al. 2021).

Our study findings are consistent with the literature; however, it can be said that hemp seed oil and hemp essential oil are superior in terms of collagen production and wound tissue development. The reason for this could potentially be attributed to the trace amounts of THC present in these compounds.

As a result, the phytocannabinoids, terpenes and other molecules in the hemp seed oil, from the first days of the experiment, showed a good healing process that could be differentiated from the other groups. Generally, there was general well-being in the 1st group, 2nd group, 3rd group and 4th group of rats. No signs of infection were found. The recovery rate in the Group 1 and 2 rats was different from the other groups. It was determined that this may be due to the effect of THC, albeit in trace amounts. It is considered that the decrease in the stress levels facilitated the activation of the immune system and may accelerate the molecular mechanism. However, there was no drowsiness or confusion. The hemp seed oil and hemp essential oil increased the EGF at a higher rate than other groups, improving the wound healing process.

It can be said that the CBD, hemp seed oil and hemp essential oil are at least as safe as the 10% povidone-iodine in the wound healing process, and they even have superior qualities over povidone-iodine. Since CBD has no superiority over cannabis extracts in the wound healing process, the use of cannabis extracts, which are easier and more effective than pure CBD, can be recommended. It can be used in routine wound care because the solution prepared from cannabis is cheap, natural, does not cause infection, and does not put the thyroid glands at risk because it does not contain iodine.

Acknowledgement

The researchers would like to express their gratitude to GY, the staff of the Hemp Research Institute, the staff of the Yozgat Bozok University Project Coordination Application and Research Institute, the staff of the Erciyes University Experimental Research, Application and Research Center, and the faculty member of the Erciyes University Medical Biochemistry Department, A.Ç. and her assistants for their support in the conduct of this research.

Conflict of interest

The authors declare no conflict of interest.

REFERENCES

- Abbas M, Hussain T, Arshad M, Ansari AR, Irshad A, Nisar J, Hussain F, Masood N, Nazir A, Iqbal M. Wound healing potential of curcumin cross-linked chitosan/polyvinyl alcohol. *Int J Biol Macromol*. 2019 Nov 1; 140:871-6.
- Aguayo-Morales H, Sierra-Rivera CA, Claudio-Rizo JA, Cobos-Puc LE. Horsetail (*Equisetum hyemale*) extract accelerates wound healing in diabetic rats by modulating IL-10 and MCP-1 release and collagen synthesis. *Pharmaceuticals (Basel)*. 2023 Mar 30;16(4):514.
- Alyami R, Al Jasser R, Alshehri FA, Alshibani N, Bin Hamdan S, Alyami RA, Niazy AA. Vitamin C influences antioxidant, anti-inflammatory and wound healing markers in smokers' gingival fibroblasts in vitro. *Saudi Dent J*. 2023 May;35(4):337-44.
- Andre CM, Hausman JF, Guerriero G. Cannabis sativa: The plant of the thousand and one molecules. *Front Plant Sci*. 2016 Feb 4;7:1-17.
- Barrientos S, Stojadinovic O, Golinko MS, Brem H, Tomic-Canic M. Growth factors and cytokines in wound healing. *Wound Repair Regen*. 2008 Sep-Oct;16(5):585-601.
- Bellocchio L, Patano A, Inchingolo AD, Inchingolo F, Dipalma G, Isacco CG, de Ruvo E, Rapone B, Mancini A, Lorusso F, Scarano A, Malcangi G, Inchingolo AM. Cannabidiol for oral health: A new promising therapeutical tool in dentistry. *Int J Mol Sci*. 2023 Jun 2;24(11):9693.
- Busanello-Costa M, Renno ACM, de Goes Santos CP, Quintana HT, Martignago CCS, Tim CR, Assis L. Red LED light therapy associated with epidermal growth factor on wound repair process in rats. *Lasers Med Sci*. 2023 Jan 10;38(1):36.

<https://doi.org/10.17221/21/2024-VETMED>

- Callaway J. Hempseed as a nutritional resource: An overview. *Euphytica*. 2004 Jan;140:65-72.
- Chen J, Ma H, Meng Y, Liu Q, Wang Y, Lin Y, Yang D, Yao W, Wang Y, He X, Li P. Analysis of the mechanism underlying diabetic wound healing acceleration by Calycosin-7-glycoside using network pharmacology and molecular docking. *Phytomedicine*. 2023 Jun;114:154773.
- Da Porto C, Decorti D, Tubaro F. Fatty acid composition and oxidation stability of hemp (*Cannabis sativa* L.) seed oil extracted by supercritical carbon dioxide. *Ind Crops Prod*. 2012 Mar;36(1):401-4.
- Ebrahimpour-Malekshah R, Amini A, Mostafavinia A, Ahmadi H, Zare F, Safaju S, Shahbazi A, Chien S, Rezaei F, Hasan A, Bayat M. The stereological, immunohistological, and gene expression studies in an infected ischemic wound in diabetic rats treated by human adipose-derived stem cells and photobiomodulation. *Arch Dermatol Res*. 2023 Aug;315(6):1717-34.
- Ferrini F, Fraternali D, Donati Zeppa S, Verardo G, Gorassini A, Carrabs V, Albertini MC, Sestili P. Yield, characterization, and possible exploitation of *Cannabis sativa* L. roots grown under aeroponics cultivation. *Molecules*. 2021 Aug 12;26(16):4889.
- Fumagalli M, Paladino G, Piazza S, Rossini N, Ciriello U, Magnavacca A, Martinelli G, Pozzoli C, Dell'Agli M, Sangiovanni E. Cannabidiol has a central role in the NF- κ B inhibition of a *Cannabis sativa* L. extract in human keratinocytes. *Z Phytother*. 2022 Jun;43(S_01):45-6.
- Gong C, Lu Y, Jia C, Xu N. Low-level green laser promotes wound healing after carbon dioxide fractional laser therapy. *J Cosmet Dermatol*. 2022 Nov;21(11):5696-703.
- Gurtner GC, Wong VW. Wound healing: Normal and abnormal. In: Thorne CH, Bartlett SP, Beasley RW, Aston SJ, Gurtner GC, Spear SL. *Grabb and Smith's plastic surgery*. 6th ed. Philadelphia: Lippincott Williams and Wilkins; 2006. p. 15-22.
- Hou Y, Jiang N, Sun D, Wang Y, Chen X, Zhu S, Zhang L. A fast UV-curable PU-PAAm hydrogel with mechanical flexibility and self-adhesion for wound healing. *RSC Adv*. 2020 Jan 29;10(9):4907-15.
- Hu X, Gu Y, Wu Q, Jiang F, Song Z. Celastrol enhances bone wound healing in rats. *Acta Biochim Pol*. 2022 Dec 2; 69(4):839-45.
- Jayabal H, Abiakam NS, Filingeri D, Bader DL, Worsley PR. Inflammatory biomarkers in sebum for identifying skin damage in patients with a Stage I pressure ulcer in the pelvic region: A single centre observational, longitudinal cohort study with elderly patients. *Int Wound J*. 2023 Sep; 20(7):2594-607.
- Kibret BG, Patel S, Niezgoda J, Guns W, Niezgoda J, Gopalakrishnan S, Baban B, Cubillos P, Villeneuve D, Kumar P. Evidence-based potential therapeutic applications of cannabinoids in wound management. *Adv Skin Wound Care*. 2022 Aug 1;35(8):447-53.
- Kim SE, Kim YJ, Cheon YW. Role of enkephalin derivative in promoting wound healing and scar remodeling via increased epidermal growth factor in a mouse model. *Arch Aesthetic Plast Surg*. 2023 Jan;29(1):1-7.
- Konca P, Akbay Y. Bronsial astma ve interlokin 8 [Bronchial asthma and interleukin-8]. *CMJ*. 2000;22(2):105-9. Turkish.
- Kongkadee K, Wisuitiprot W, Ingkaninan K, Waranuch N. Anti-inflammation and gingival wound healing activities of *Cannabis sativa* L. subsp. *sativa* (hemp) extract and cannabidiol: An in vitro study. *Arch Oral Biol*. 2022 Aug; 140:105464.
- Kurtoglu S, Bastug O, Daar G, Halis H, Korkmaz L, Memur S, Korkut S, Gunes T, Ozturk MA. Effect of iodine loading on the thyroid hormone level of newborns living in Kayseri province. *Am J Perinatol*. 2014 Dec;31(12):1087-92.
- Liang J, Appukuttan Aachary A, Thiyam-Hollander U. Hemp seed oil: Minor components and oil quality. *Lipid Technol*. 2015 Sep;27(10):231-3.
- Liska DJ, Reed MJ, Sage EH, Bornstein P. Cell-specific expression of alpha 1(I) collagen-hGH minigenes in transgenic mice. *J Cell Biol*. 1994 May;125(3):695-704.
- Majno G. *The healing hand: Man and wound in the ancient world*. Harvard University Press; 1975. 571 p.
- Metcalfe AD, Ferguson MW. Tissue engineering of replacement skin: The crossroads of biomaterials, wound healing, embryonic development, stem cells and regeneration. *J R Soc Interface*. 2007 Jun 22;4(14):413-37.
- Mikulec A, Kowalski S, Sabat R, Skoczylas L, Tabaszewska M, Wywrocka-Gurgul A. Hemp flour as a valuable component for enriching physicochemical and antioxidant properties of wheat bread. *LWT*. 2019 Mar;102:164-72.
- Mirasoglu B. Yara bakim urunleri [Wound care products]. *TOTBİD Dergisi*. 2015 Jul;14:456-61. Turkish.
- Narisepalli S, Salunkhe SA, Chitkara D, Mittal A. Asiaticoside polymeric nanoparticles for effective diabetic wound healing through increased collagen biosynthesis: In-vitro and in-vivo evaluation. *Int J Pharm*. 2023 Jan 25;631: 122508.
- Nunan R, Harding KG, Martin P. Clinical challenges of chronic wounds: Searching for an optimal animal model to recapitulate their complexity. *Dis Model Mech*. 2014 Nov;7(11):1205-13.
- Passani A, Posarelli C, Sframeli AT, Perciballi L, Pellegrini M, Guidi G, Figus M. Cannabinoids in glaucoma patients: The never-ending story. *J Clin Med*. 2020 Dec 8;9(12):3978.
- Pojic M, Misan A, Sakac M, Dapcevic Hadnadev T, Saric B, Milovanovic I, Hadnadev M. Characterization of byprod-

<https://doi.org/10.17221/21/2024-VETMED>

- ucts originating from hemp oil processing. *J Agric Food Chem.* 2014 Dec 24;62(51):12436-42.
- Raja SK, Garcia MS, Isseroff RR. Wound re-epithelialization: Modulating keratinocyte migration in wound healing. *Front Biosci.* 2007 May 1;12:2849-68.
- Rapa M, Ciano S, Rocchi A, D'Ascenzo F, Ruggieri R, Vinci G. Hempseed oil quality parameters: Optimization of sustainable methods by miniaturization. *Sustainability.* 2019 Jun 1;11:1-13.
- Rizzo G, Pineda Chavez SE, Vandenkoornhuysen E, Cardenas Rincon CL, Cento V, Garlatti V, Wozny M, Sammarco G, Di Claudio A, Meanti L, Elangovan S, Romano A, Roda G, Loy L, Dal Buono A, Gabbiadini R, Lovisa S, Rusconi R, Repici A, Armuzzi A, Vetrano S. Pomegranate extract affects gut biofilm forming bacteria and promotes intestinal mucosal healing regulating the crosstalk between epithelial cells and intestinal fibroblasts. *Nutrients.* 2023 Apr 5;15(7):1771.
- Sangiovanni E, Fumagalli M, Pacchetti B, Piazza S, Magnavacca A, Khalilpour S, Melzi G, Martinelli G, Dell'Agli M. Cannabis sativa L. extract and cannabidiol inhibit in vitro mediators of skin inflammation and wound injury. *Phytother Res.* 2019 Aug;33(8):2083-93.
- Scharffetter K, Kulozik M, Stolz W, Lankat-Buttgereit B, Hatamochi A, Sohnchen R, Krieg T. Localization of collagen $\alpha 1$ (I) gene expression during wound healing by in situ hybridization. *J Invest Dermatol.* 1989 Sep;93(3):405-12.
- Schluttenhofer C, Yuan L. Challenges towards revitalizing hemp: A multifaceted crop. *Trends Plant Sci.* 2017 Nov;22(11):917-29.
- Spinella A, de Pinto M, Baraldi C, Galluzzo C, Testoni S, Lumetti F, Parenti L, Guerzoni S, Salvarani C, Giuggioli D. Topical cannabidiol in the treatment of digital ulcers in patients with scleroderma: Comparative analysis and literature review. *Adv Skin Wound Care.* 2023 Jan 1;36(1):18-23.
- Teh SS, Birch J. Physicochemical and quality characteristics of cold-pressed hemp, flax and canola seed oils. *J Food Compost Anal.* 2013 May;30(1):26-31.
- Thapa D, Cairns EA, Szczesniak AM, Kulkarni PM, Straiker AJ, Thakur GA, Kelly MEM. Allosteric cannabinoid receptor 1 (CB1) ligands reduce ocular pain and inflammation. *Molecules.* 2020 Jan 20;25(2):417.
- Wang XS, Tang CH, Yang XQ, Gao WR. Characterization, amino acid composition and in vitro digestibility of hemp (*Cannabis sativa* L.) proteins. *Food Chem.* 2008 Mar;107(1):11-8.
- Xu R, Luo G, Xia H, He W, Zhao J, Liu B, Tan J, Zhou J, Liu D, Wang Y, Yao Z, Zhan R, Yang S, Wu J. Novel bilayer wound dressing composed of silicone rubber with particular micropores enhanced wound re-epithelialization and contraction. *Biomaterials.* 2015 Feb;40:1-11.
- Yang SL, Han R, Liu Y, Hu LY, Li XL, Zhu LY. Negative pressure wound therapy is associated with up-regulation of bFGF and ERK1/2 in human diabetic foot wounds. *Wound Repair Regen.* 2014 Jul-Aug;22(4):548-54.
- Yasti AC, Karaca T, Kendirci M, Akgun AE, Sahiner IT, Akin M. Comparison of the efficiency of epidermal growth factor and negative pressure wound therapy in diabetic foot patients. *Int J Low Extrem Wounds.* 2023 Mar;22(1):93-102.
- Zagorska-Dziok M, Bujak T, Ziemlewska A, Nizioł-Lukaszewska Z. positive effect of Cannabis sativa L. herb extracts on skin cells and assessment of cannabinoid-based hydrogels properties. *Molecules.* 2021 Feb 4;26(4):802.
- Zhang H, Zhou M, Wang Y, Zhang D, Qi B, Yu A. Role of autologous fat transplantation combined with negative-pressure wound therapy in treating rat diabetic wounds. *Plast Reconstr Surg.* 2023 Sep 1;152(3):561-70.

Received: March 5, 2024

Accepted: May 28, 2024

Published online: August 16, 2024