

ÚSTAV ZEMĚDĚLSKÝCH A POTRAVINÁŘSKÝCH INFORMACÍ

VETERINÁRNÍ MEDICÍNA

Veterinary Medicine – Czech

ČESKÁ AKADEMIE ZEMĚDĚLSKÝCH VĚD

7

VOLUME 45
PRAHA
JULY 2000
ISSN 0375-8427

Mezinárodní vědecký časopis vydávaný z pověření Ministerstva zemědělství České republiky a pod gesci České akademie zemědělských věd

An international journal published under the authorization by the Ministry of Agriculture and under the direction of the Czech Academy of Agricultural Sciences

Editorial Board – Redakční rada

Chairman – Předseda

Prof. MVDr. Karel Hruška, CSc., Veterinary Research Institute, Brno, Czech Republic

Members – Členové

Doc. MVDr. ing. Jiří Brož, CSc., Reinfelden, Switzerland

Arnost Cepica, DVM., PhD., Associate Professor (Virology/Immunology), Atlantic Veterinary College, U.P.E.I., Charlottetown, Canada

Dr. Milan Fránek, DrSc., Veterinary Research Institute, Brno, Czech Republic

Doc. MVDr. Ivan Herzig, CSc., Veterinary Research Institute, Brno, Czech Republic

Prof. MVDr. Bohumír Hofírek, DrSc., University of Veterinary and Pharmaceutical Sciences, Brno, Czech Republic

Prof. MUDr. Drahomír Horký, DrSc., Faculty of Medicine, Masaryk University, Brno, Czech Republic

Doc. MVDr. RNDr. Petr Hořín, CSc., University of Veterinary and Pharmaceutical Sciences, Brno, Czech Republic

Doc. MVDr. František Kovářů, DrSc., University of Veterinary and Pharmaceutical Sciences, Brno, Czech Republic

Doc. MVDr. Dr. Jozef Laurinčík, DrSc., Institute of Genetics and Experimental Biology, RIAP, Nitra, Slovak Republic

Prof. MUDr. M. V. Nermut, PhD., DSc. (h. c.), National Institute for Biological Standards and Control, United Kingdom

Prof. MUDr. MVDr. h. c. Leopold Pospíšil, DrSc., Veterinary Research Institute, Brno, Czech Republic

Prof. RNDr. Václav Suchý, DrSc., University of Veterinary and Pharmaceutical Sciences, Brno, Czech Republic

Prof. MVDr. Bohumil Ševčík, DrSc., BIOPHARM – Research Institute of Biopharmacy and Veterinary Drugs, a. s.,

Jilové u Prahy, Czech Republic

Prof. MVDr. Zdeněk Věžník, DrSc., Veterinary Research Institute, Brno, Czech Republic

Editor-in-Chief – Vedoucí redaktorka

Ing. Zdeňka Radošová

World Wide Web (URL): <http://www.vri.cz> or <http://www.uzpi.cz>

Cíl a odborná náplň: Časopis Veterinární medicína uveřejňuje původní vědecké práce a studie typu review ze všech oblastí veterinární medicíny v češtině, slovenštině a angličtině.

Časopis je citován v bibliografickém časopise Current Contents – Agriculture, Biology and Environmental Sciences, a abstrakty z časopisu jsou zahrnuty v těchto databázích: Agris, CAB Abstracts, Current Contents on Diskette – Agriculture, Biology and Environmental Sciences, Czech Agricultural Bibliography, Toxline Plus.

Periodicita: Časopis vychází měsíčně (12x ročně), ročník 45 vychází v roce 2000.

Přijímání rukopisů: Rukopisy ve třech vyhotoveních je třeba zaslat na adresu redakce: Ing. Zdeňka Radošová, vedoucí redaktorka, Ústav zemědělských a potravinářských informací, Slezská 7, 120 56 Praha 2, Česká republika. Tel.: +420 2 24 25 79 39, fax: +420 2 24 25 39 38, e-mail: edit@uzpi.cz. Podrobné pokyny pro autory jsou v redakci a na URL adrese <http://www.vri.cz> nebo <http://www.uzpi.cz>

Informace o předplatném: Objednávky na předplatné jsou přijímány pouze na celý rok (leden–prosinec) a zasílají se na adresu: Ústav zemědělských a potravinářských informací, vydavatelské oddělení, Slezská 7, 120 56 Praha 2. Cena předplatného pro rok 2000 je 696 Kč.

Aims and scope: The journal Veterinární medicína original publishes papers and reviews from all fields of veterinary medicine written in Czech, Slovak or English.

The journal is cited in the bibliographical journal Current Contents – Agriculture, Biology and Environmental Sciences, abstracts from the journal are comprised in the databases: Agris, CAB Abstracts, Current Contents on Diskette – Agriculture, Biology and Environmental Sciences, Czech Agricultural Bibliography, Toxline Plus.

Periodicity: The journal is published monthly (12 issues per year), Volume 45 appearing in 2000.

Acceptance of manuscripts: Three copies of manuscript should be addressed to: Ing. Zdeňka Radošová, editor-in-chief, Institute of Agricultural and Food Information, Slezská 7, 120 56 Praha 2, Czech Republic. Tel.: +420 2 24 25 79 39, fax: +420 2 24 25 39 38, e-mail: edit@uzpi.cz. Detailed instructions for authors are available in the editorial office and at URL address <http://www.vri.cz> or <http://www.uzpi.cz>

Subscription information: Subscription orders can be entered only by calendar year (January–December) and should be sent to: Institute of Agricultural and Food Information, Slezská 7, 120 56 Praha 2. Subscription price for 2000 is 159 USD (Europe), 167 USD (overseas).

THE EFFECT OF PLACEBO-INHALER ON THE TRACHEAL EPITHELIUM IN RABBITS*

ÚČINEK PREPARÁTU PLACEBO-INHALER NA EPITEL TRACHEY KRÁLÍKŮ

V. Konrádová¹, L. Vajner¹, J. Uhlík¹, J. Zocová²

¹ *Institute of Histology and Embryology, 2nd Medical Faculty,* ² *Department of Applied Mathematics and Computer Science, Faculty of Science, Charles University, Praha, Czech Republic*

ABSTRACT: In experiment, the effect of two puffs of aerosol of Placebo-inhaler on the ultrastructure of the tracheal epithelium and on the composition of glycoconjugates of the goblet cells was studied. Using standard methods of electron microscopy, the ciliary border and the functional state of the goblet cells were evaluated quantitatively. The character of glycoconjugates was studied by means of methods of conventional and lectin histochemistry. 30 minutes after inhalation of propellants contained in the metered-dose inhaler, the ciliated cells revealed only slight signs of alteration reflected in a mild impairment of the ciliary border. 16% of goblet cells were stimulated to discharge mucus, 4% of them degenerated. Neutral glycoconjugates disappeared from the epithelium and insignificant decrease in acid sialylated glycoconjugates was demonstrated. Morphological signs of impaired self-cleaning ability of the tracheal epithelium were not encountered. The propellants contained in the metered-dose inhalers do not cause the damage to the airway epithelium due to the inhalation of bronchospasmolytic drugs.

airways; epithelium; Placebo-inhaler, ultrastructure; lectin histochemistry; glycoconjugates

ABSTRAKT: V experimentu jsme studovali účinek dvou vdechů preparátu Placebo-inhaler na ultrastrukturu epitelu trachey a na složení glykokonjugátů obsažených v pohárkových buňkách. Kvantitativně jsme hodnotili stupeň narušení řasinkového lemu nad epitelem a funkční stav pohárkových buněk. Pro studium složení glykokonjugátů jsme užíli metody klasické a lektinové histochemie. 30 min po inhalaci propelentů obsažených v tlakových dávkovacích nádobkách jevíly řasinkové buňky jen lehké známky patologické alterace, které se odrazily v mírném narušení řasinkového lemu. 16 % pohárkových buněk bylo stimulováno k vydávání sekretu, 4% z nich byly zcela vyprázdňené a degenerovaly. V epitelu došlo k nevýznamnému snížení množství kyselých sialovaných glykokonjugátů a neutrální glykokonjugáty zcela vymizely. Morfologické známky narušení samočisticí schopnosti epitelu jsme nezaznamenali. Propelenty obsažené v tlakových dávkovacích nádobkách nezpůsobují poškození epitelu dýchacích cest, které jsme pozorovali po aplikaci aerosolu bronchospasmolytických preparátů.

dýchací cesty; epitel; Placebo-inhaler; ultrastruktura; lektinová histochemie; glykokonjugáty

INTRODUCTION

In our previous studies (Konrádová et al., 1997, 1998; Vajner, 1998), the ultrastructure of the tracheal epithelium and the composition of glycoconjugates contained in the goblet cells' secretion were studied after inhalation of single therapeutic doses of two β_2 adrenergic agonists – salbutamol (Ventolin) and fenoterol hydrobromide (Berotec) – and a cholinergic antagonist – ipratropium bromide (Atrovent). Due to this treatment, the injury

to the tracheal epithelium was moderate to severe. In the goblet cells' secretion, an absolute predominance of acid sulphated glycoconjugates was revealed. Using a light microscope, Spahr-Schopfer and Shorten with their fellow-workers described the adverse effect of salbutamol on the airway epithelium (Spahr-Schopfer et al., 1994; Shorten et al., 1995). As these authors suggested that the epithelial damage was most likely due to the Ventolin propellants, especially to the oleic acid contained in the metered-dose inhaler, we decided to study the effect of

* Supported by the Grant Agency of Charles University (Grant No. 185/98) and by the Ministry of Education, Youth and Sports of the Czech Republic (Research project No. 111300003).

a single dose of Placebo-inhaler on the airway epithelium. This aerosol contains all the substances of Ventolin with the exception of salbutamol.

MATERIAL AND METHODS

In our experiments, 6 specific pathogen free New Zealand White rabbits (body weight 1 500–3 000 g, Charles River Deutschland, Sulzfeld, Germany) were used. The untreated controls represented the same groups of animals used in our previous studies (Konrádová et al., 1997, 1998; Vajner, 1998). The animals were treated with two puffs of Placebo-inhaler (Glaxo, Greenford, Great Britain). The metered-dose inhaler containing only propellants was connected with a long thin catheter that was inserted into the mouth of a rabbit under general anaesthesia and two puffs of aerosol were administered into its airways during spontaneous inspiration. The material for the electron microscopic and his-

tochemical examination was collected 30 minutes post exposure. Tiny fragments of the tracheal mucous membrane were processed using standard methods for electron microscopy. The ciliary border and the functional state of the goblet cells were evaluated quantitatively as described in our previous paper (Konrádová et al., 1997, 1998). After administration of two puffs of Placebo-inhaler, 482 goblet cells and 2 150.5 μm^2 of ciliary border with 18 019 kinocilia were evaluated. Methods of conventional and lectin histochemistry described in our previous studies were used to study the character of glycoconjugates contained in the goblet cells' secretion (Vajner, 1998).

RESULTS

In control rabbits, the tracheae were lined with a ciliated pseudostratified columnar epithelium. The ciliated cells of standard ultrastructure were the most numerous

1. Glycoconjugates in the goblet cells (GC) in the tracheal epithelium of rabbits 30 minutes after inhalation of 2 puffs of propellants (Placebo-inhaler) and salbutamol (Ventolin)

| | | Control rabbits | Propellants (Placebo-inhaler) | Salbutamol (Ventolin) |
|--|---|-----------------|-------------------------------|-----------------------|
| GC containing neutral GCC (PAS positive) | % | 1.5 ± 2.4 | 0.0 ± 0.0 | 0.0 ± 0.0 |
| GC containing acid GCC (AB pH 2.5 positive) | % | 98.5 ± 2.4 | 100.0 ± 0.0 | 100.0 ± 0.0 |
| GC containing sulphated acid GCC (AB pH 1 positive) | % | 71.9 ± 6.4 | 74.2 ± 5.4 | *100.0 ± 0.0 |
| GC containing sialylated acid GCC (calculation) | % | 26.6 ± 6.4 | 28.5 ± 5.4 | *0.0 ± 0.0 |
| GC containing sialylated acid GCC (MAA positive) | % | 27.9 ± 9.6 | 24.4 ± 12.7 | *0.0 ± 0.0 |
| GC containing sialylated acid GCC (SNA positive) | % | 2.3 ± 2.9 | 0.3 ± 0.5 | 0.0 ± 0.0 |
| GC containing sialylated acid GCC (MAA+SNA positive) | % | 30.2 ± 9.6 | 24.7 ± 13.2 | *0.0 ± 0.0 |
| GC containing sialylated acid GCC (TML positive) | % | 26.6 ± 11.5 | 21.9 ± 7.4 | *0.3 ± 0.5 |

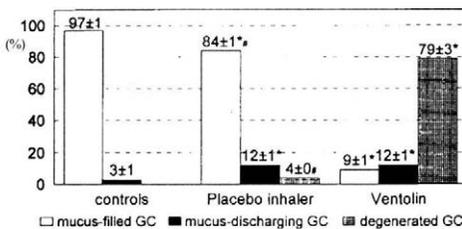
Explanation:

n = 4 in controls, 3 in others, mean ± SD

values marked * differ significantly ($\alpha = 0.01$) from controls, values connected by a line differ significantly ($\alpha = 0.01$) from each other

GCC = glycoconjugates, PAS = periodic acid - Schiff reaction, AB = alcian blue

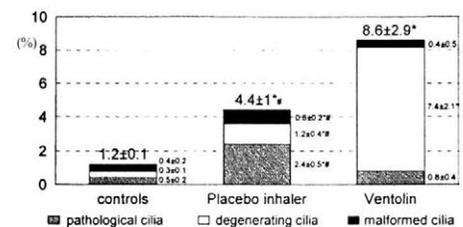
MAA = *Maackia amurensis* agglutinin, SNA = *Sambucus nigra* agglutinin, TML = *Trichomonas mobilensis* lectin



1. Functional state of goblet cells (GC) in the tracheal epithelium of rabbits 30 minutes after inhalation of 2 puffs of propellants (Placebo-inhaler) and salbutamol (Ventolin)

n = 3, mean ± SD

values marked * differ significantly ($\alpha = 0.01$) from controls values marked # differ significantly ($\alpha = 0.01$) from corresponding values in the group of rabbits after inhalation of Ventolin



2. Altered cilia in the tracheal ciliary border of rabbits 30 minutes after inhalation of 2 puffs of propellants (Placebo-inhaler) and salbutamol (Ventolin)

n = 3, mean ± SD

values marked * differ significantly ($\alpha = 0.01$) from controls values marked # differ significantly ($\alpha = 0.01$) from corresponding values in the group of rabbits after inhalation of Ventolin

in the epithelium. Goblet cells mostly filled with mucus were scattered among the ciliated ones. Only $3 \pm 1\%$ of them discharged their secretion by means of gradual evacuation of the individual apical mucous granules (Fig. 1). Both the conventional and lectin histochemistry revealed the dominance of goblet cells containing acid glycoconjugates. The proportion of goblet cells containing neutral and acid sulphated glycoconjugates, detected by conventional histochemistry, and the total percentage of goblet cells containing acid sialylated glycoconjugates visualised by the reactions with the lectins were given in Tab. I. In the regular ciliary border, 9.7 ± 0.3 cilia per $1 \mu\text{m}^2$ were found. $98.8 \pm 0.1\%$ of cilia were intact. The proportions of pathological, degenerating and malformed cilia were given in Fig. 2.

After inhalation of the aerosol of propellants contained in the metered-dose inhaler, a slightly altered pseudostratified columnar ciliated epithelium was encountered in the tracheae. The intercellular spaces were narrow and the apical junctional complexes remained intact (Fig. 3).

Only a few tiny cytoplasmic protrusions exceptionally containing isolated axonemes of degenerating kinocilia were found on the apical portions of the ciliated cells (Fig. 4). A slight increase in number of small vacuoles and lysosomes was observed in the deeper portions of their cytoplasm (Figs. 5, 6). In some cells, larger intracytoplasmic ciliated vacuoles were revealed. Only isolated differentiating ciliated or secretory cells were encountered in the epithelium.

The goblet cells were mostly scattered among the ciliated ones, only $8 \pm 1\%$ of them formed tiny groups. $16 \pm 1\%$ of the goblet cells were stimulated to discharge mucus (Figs 7, 8). The proportions of mucus-discharging and exhausted secretory elements were given in Fig. 1. Mucus was evacuated from apical mucous granules. Only exceptionally, whole packets of mucous granules were detached or signs of compound exocytosis were noticed in the stimulated goblet cells. In the goblet cells secretion, neutral glycoconjugates were not revealed. The percentage of goblet cells containing the acid sul-

phated and lectin-detected acid sialylated glycoconjugates was presented in the Tab. I.

A slightly impaired ciliary border with 8.4 ± 0.4 cilia per $1 \mu\text{m}^2$ was observed. The altered cilia represented only $4.4 \pm 1\%$. The proportions of the individual types of altered cilia were given in Fig. 2. In the area of the ciliary border, neither bacteria nor clumps of inspissated secretion were observed.

DISCUSSION

On the basis of our previous experiments, a classification of the degree of injury to the airway epithelium was proposed (Konrádová, 1991). Due to the administration of two puffs of Placebo-inhaler, mostly only mild damage of this epithelium was revealed. The mucus evacuation was induced in only one sixth of all goblet cells, the mechanism of their secretion was slightly accelerated.

In our previous studies, we demonstrated that degeneration of more than half of the goblet cells stimulated a massive differentiation of new secretory elements resulting in their hyperplasia and changes in their distribution with the occurrence of intraepithelial mucous glands (Konrádová et al., 1990; Konrádová, 1991). As only 4% of goblet cells degenerated, the differentiation of secretory elements was not stimulated.

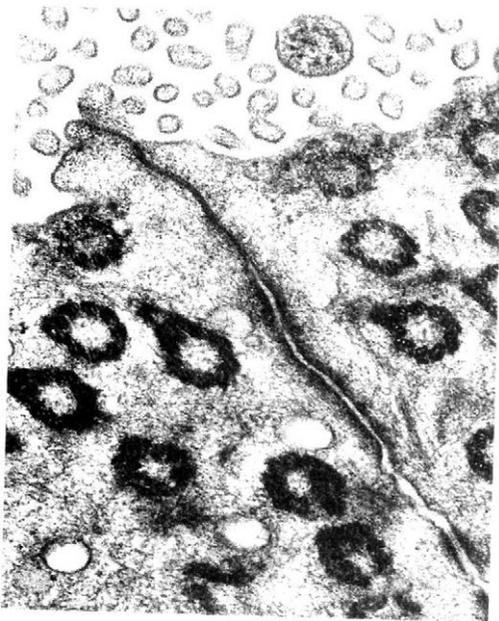
In accordance with other authors (Jacob and Poddar, 1982; Jeffery et al., 1982; Spicer et al., 1983; Mandal and Mandal, 1990; Castells et al., 1991, 1992), we ascertained the presence of neutral and both sulphated and sialylated glycoconjugates in the secretion of the goblet cells in healthy rabbits. The majority of acid glycoconjugates to neutral ones and the dominance of sulphated acid glycoconjugates were revealed by both histochemical and lectin-histochemistry methods. After administration of propellants, the neutral glycoconjugates disappeared from the epithelium. The composition of acid glycoconjugates produced by the goblet cells did not differ significantly from that found in healthy control rabbits. Only

II. Evaluation of the degree of damage to the tracheal epithelium of rabbits 30 minutes after inhalation of 2 puffs of propellants (Placebo-inhaler) and salbutamol (Ventolin)

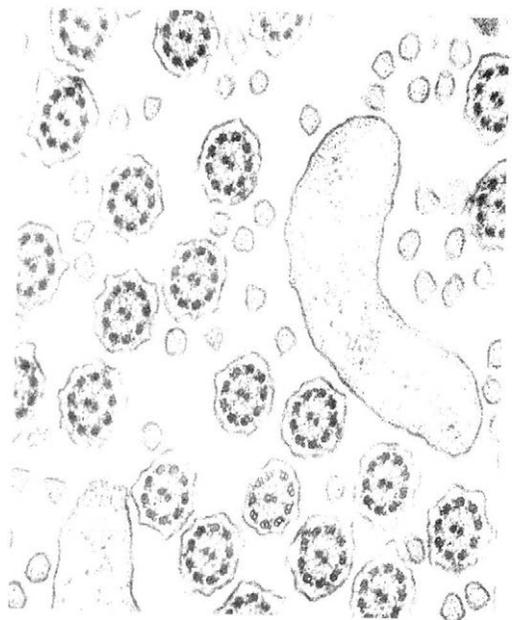
| | Degree of damage to the tracheal epithelium | | | Control rabbits | Propellants (Placebo-inhaler) | Salbutamol (Ventolin) |
|---|---|-----------|---------|--------------------------|-------------------------------|-----------------------|
| | mild | moderate | severe | | | |
| Stimulated GC | 3-50% | 50-90% | > 90% | 3% | 16% | 91% |
| Ratio $\frac{\text{discharging GC}}{\text{degenerated GC}}$ | > 1 | 0.1-1 | < 0.1 | degenerated GC not found | 3 | 0.15 |
| Number of cilia/ μm^2 | 7-9 | 3-7 | < 3 | 9.7 | 8.4 | 5.2 |
| Altered cilia | 1.2-2.0% | 2.0-10.0% | > 10.0% | 1.2% | 4.4% | 8.6% |
| Signs of impairment of the self cleaning ability | ± | + | ++ | 0 | 0 | + |

Explanation:

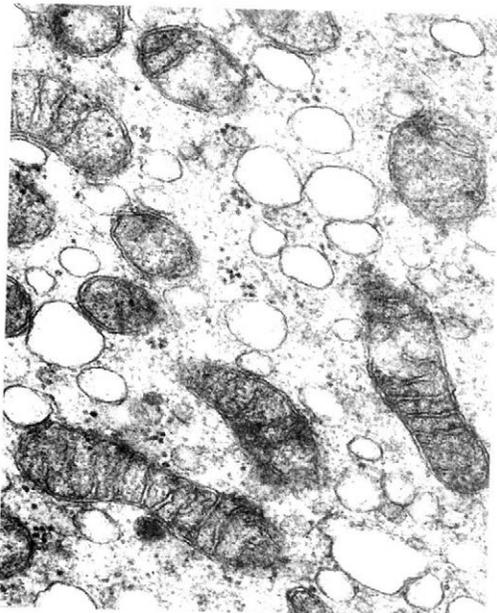
GC = goblet cells, 0 = no impairment of the self cleaning ability, ± = isolated bacteria in the area of the ciliary border, + = small clumps of condensed secretion and bacteria in the area of the ciliary border, ++ = voluminous clumps or layers of condensed secretion and numerous bacteria in the area of the ciliary border



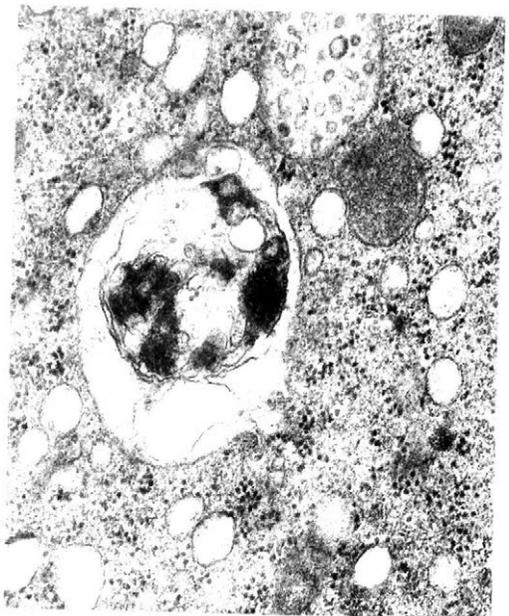
3. Apical junctional complexes between two ciliated cells. Tracheal epithelium (rabbit) - 30 min after inhalation of two puffs of Placebo-inhaler; 50 000x



4. Small apical cytoplasmic protrusions in the area of the ciliary border. Tracheal epithelium (rabbit) - 30 min after inhalation of two puffs of Placebo-inhaler; 50 000x



5. Numerous tiny vacuoles in the cytoplasm of a ciliated cell. Tracheal epithelium (rabbit) - 30 min after inhalation of two puffs of Placebo-inhaler; 50 000x



6. Larger secondary lysosome in the cytoplasm of a ciliated cell. Tracheal epithelium (rabbit) - 30 min after inhalation of two puffs of Placebo-inhaler; 50 000x



7. Apical portion of a stimulated goblet cell. Tracheal epithelium (rabbit) – 30 min after inhalation of two puffs of Placebo-inhaler; 37 500 \times



8. Apical portion of exhausted degenerated goblet protruding above the epithelium. Tracheal epithelium (rabbit) – 30 min after inhalation of two puffs of Placebo-inhaler; 37 000 \times

a slight, insignificant decrease in acid sialylated glycoconjugates, especially due to the absence of glycoconjugates sialylated at α (2–6) glycosidic linkage to galactose or N-acetylgalactosamine, was demonstrated.

As a result of Placebo-inhaler administration, the ciliated cells revealed slight signs of alteration that was reflected in the impairment of the ciliary border. In comparison with healthy control rabbits, only mild, but significant ($\alpha = 0.01$) decrease in average number of kinocilia together with an increase in number of altered kinocilia were noticed. Among the altered cilia, the slightly damaged pathological ones were the most numerous.

Morphological signs of impaired self-cleaning ability of the tracheal epithelium were not encountered (Konrádová, 1991; Stratmann et al., 1991; Konrádová et al., 1996a; Wanner et al., 1996; Geiser et al., 1997). In the area of the ciliary border, neither condensed mucus nor bacteria were observed.

To verify the theory that the damage produced in the airway epithelium by Ventolin aerosol is mostly due to the propellants contained in the metered-dose inhalers, we compared the effect of administration of Placebo-inhaler with that of Ventolin.

As we described in our previous studies (Konrádová et al., 1997, 1998), the treatment with two puffs of Ventolin aerosol, where 0.2 mg of salbutamol was contained, highly affected the secretory elements. More than 90%

of goblet cells were stimulated to discharge their mucus. Mucus release was significantly accelerated and the mechanism of secretion was influenced. Signs of an apocrine type of secretion and of a rapid compound exocytosis (Specian and Neutra, 1980; Kurosumi et al., 1981; Roumagnac and Laboisse, 1987; Specian and Oliver, 1991; Konrádová et al., 1996b; Newman et al., 1996) were noticed in numerous cells. An absolute predominance of acid sulphated glycoconjugates was revealed in their secretion. After rapid release of mucus, 79% of exhausted cells degenerated and were gradually sloughed off. Rather numerous differentiating secretory elements were noticed in the epithelium, but their distribution in the epithelium was not influenced. The ciliated cells were less damaged compared with the goblet ones. Blebbing of the apical portions of their cytoplasm was associated with destruction of rather numerous kinocilia. Marked signs of pathological alteration were also noticed in the deeper portions of the ciliated cells' cytoplasm. In the area of the ciliary border, a significant ($\alpha = 0.01$) decrease in the mean number of kinocilia to $5.2/\mu\text{m}^2$ accompanied with a significant ($\alpha = 0.01$) increase in percentage of altered cilia was recorded in comparison with healthy control rabbits. Layers of condensed secretion and bacteria representing morphological signs of impaired self-cleaning ability of the epithelium were discovered among free cilia.

After administration of propellants contained in Placebo-inhaler, significantly less pronounced ultrastructural alteration and only slight changes in the composition of glycoconjugates in the goblet cells were encountered. We are thus not able to confirm Spahr-Schopfer's and Shorten's hypothesis (Spahr-Schopfer et al., 1994; Shorten et al., 1995) that damage to the airway epithelium due to the inhalation of bronchospasmodic drug Ventolin is mostly caused by the propellants contained in the metered-dose inhalers.

Acknowledgements

We thank Dr. Pavel Babál (Department of Pathology, University of South Alabama, Mobile, U.S.A.) for the generous supply of the *Tritrichomonas mobilensis* lectin and the mouse primary monoclonal antibody.

REFERENCES

- Castells M. T., Ballesta J., Madrid J. F., Avilés M., Martínez-Menárguez J. A. (1991): Characterization of glycoconjugates in developing rat respiratory system by means of conventional and lectin histochemistry. *Histochemistry*, *95*, 419–426.
- Castells M. T., Ballesta J., Madrid J. F., Martínez-Menárguez J. A., Avilés M. (1992): Ultrastructural localization of glycoconjugates in human bronchial glands: The subcellular organization of N- and O-linked oligosaccharide chains. *J. Histochem. Cytochem.*, *40*, 265–274.
- Geiser M., Imhof V., Siegenthaler W., Grunder R., Gehr P. (1997): Ultrastructure of the aqueous lining layer in hamster airways. *Microsc. Res. Tech.*, *36*, 428–437.
- Jacob S., Poddar S. (1982): Mucous cells of the tracheobronchial tree in the ferret. *Histochemistry*, *73*, 599–605.
- Jeffery P. K., Ayers M. M., Rogers D. F. (1982): The mechanism and control of bronchial mucous cell hyperplasia. *Chest*, *81*, 27–29.
- Konrádová V. (1991): Quantitative evaluation of the degree of damage to tracheal epithelium. *Funct. Dev. Morphol.*, *1*, 47–50.
- Konrádová V., Kanta J., Šulová J. (1990): Effect of bronchoalveolar lavage on the ultrastructure of the tracheal epithelium in rabbits. *Respiration*, *57*, 14–20.
- Konrádová V., Uhlík J., Zocová J., Zajícová A. (1996a): Comparison of the effect of a high and low dose of atropine on the ultrastructure of the tracheal epithelium. *Respiration*, *63*, 150–154.
- Konrádová V., Uhlík J., Vajner L., Zocová J. (1996b): Reaction of the goblet cells to the cholinergic stimulation. *Acta Vet. Brno*, *65*, 175–180.
- Konrádová V., Uhlík J., Vajner L., Zocová J. (1997): Effect of an adrenergic agonist and a cholinergic antagonist on the airway epithelium. *Vet. Med. – Czech*, *42*, 289–293.
- Konrádová V., Uhlík J., Vajner L., Zocová J. (1998): The effect of two β_2 adrenergic agonists on the ultrastructure of the airway epithelium in rabbits. *Vet. Med. – Czech*, *43*, 187–192.
- Kurosumi K., Shibuichi I., Tosaka, K. (1981): Ultrastructural studies on the secretory mechanism of goblet cells in the jejunal epithelium. *Arch. Histol. Jpn.*, *44*, 263–284.
- Mandal C., Mandal C. (1990): Sialic acid binding lectins. *Experientia*, *46*, 433–441.
- Newman T. M., Robichaud A., Rogers D. F. (1996): Microanatomy of secretory granule release from guinea pig tracheal goblet cells. *Am. J. Respir. Cell. Mol. Biol.*, *15*, 529–539.
- Roumagnac I., Laboisie C. (1987): A mucus-secreting human colonic epithelial cell line responsive to cholinergic stimulation. *Biol. Cell.*, *61*, 65–68.
- Shorten G. D., Dolovich M., Eng P., Lerman J., Cutz E. (1995): Metered-dose inhaler salbutamol-induced tracheal epithelial lesions in intubated rabbits. *Chest*, *108*, 1668–1672.
- Spahr-Schopfer I. A., Lerman J., Cutz E., Newhouse M. T., Dolovich M. (1994): Proximate delivery of a large experimental dose from salbutamol MDI induces epithelial airway lesions in intubated rabbits. *Am. J. Respir. Crit. Care Med.*, *150*, 790–794.
- Specian R. D., Neutra M. R. (1980): Mechanism of rapid mucus secretion in goblet cells stimulated by acetylcholine. *J. Cell. Biol.*, *85*, 626–640.
- Specian R. D., Oliver M. G. (1991): Functional biology of intestinal goblet cells. *Am. J. Physiol.*, *260*, C183–C193.
- Spicer S. S., Schulte B. A., Thomopoulos G. N. (1983): Histochemical properties of the respiratory tract epithelium in different species. *Am. Rev. Respir. Dis.*, *128*, S20–S26.
- Stratmann U., Lehmann R. R., Steinbach T., Wessling G. (1991): Effect of sulphur dioxide inhalation on the respiratory tract of the rat. *Zbl. Hyg.*, *192*, 324–335.
- Vajner L. (1998): Účinek dvou β_2 sympatomimetických bronchospasmodik na podíl sialovaných a sulfonovaných glykokonjugátů v sekretu pohárkových buněk tracheálního epitelu. *Čs. patol.*, *34*, 13–16.
- Wanner A., Salathé M., O'Riordan T. G. (1996): Mucociliary clearance in the airways. *Am. J. Respir. Crit. Care Med.*, *154*, 1868–1902.

Received: 00–04–28

Accepted after corrections: 00–06–07

Contact Address:

Prof. MUDr. Václava Konrádová, DrSc., Ústav histologie a embryologie, 2. Lékařská fakulta, Univerzita Karlova, V Úvalu 84, 150 06 Praha 5, Česká republika
Tel. + 420 2 24 43 59 80, fax + 420 2 24 43 58 20, e-mail: vaclava.konradova@lfmotol.cuni.cz

BIOCHEMICAL MARKERS OF DIOXIN-LIKE TOXICITY AND OXIDATIVE STRESS IN HEPATIC MICROSOMES OF BREAM (*ABRAMIS BRAMA*) AND PERCH (*PERCA FLUVIATILIS*) IN THE ELBE RIVER*

BIOCHEMICKÉ MARKERY TOXICITY DIOXINOVÉHO TYPU A OXIDATIVNÍHO STRESU V CEJNU VELKÉM (*ABRAMIS BRAMA*) A OKOUNU ŘÍČNÍM (*PERCA FLUVIATILIS*) Z ŘEKY LABE

M. Machala¹, K. Hilscherová², R. Kubínová³, R. Ulrich¹, B. Vykusová⁴, J. Kolářová⁴, J. Máchová⁴, Z. Svobodová^{3,4}

¹ *Veterinary Research Institute, Brno, Czech Republic*

² *Faculty of Science, Masaryk University, Brno, Czech Republic*

³ *Veterinary and Pharmaceutical University, Brno, Czech Republic*

⁴ *Research Institute of Fisheries and Hydrobiology, Vodňany, Czech Republic*

ABSTRACT: Biochemical markers of dioxin-like toxicity and oxidative stress were investigated in hepatic microsomal fractions of bream (*Abramis brama*) and perch (*Perca fluviatilis*) collected at six sites of the Elbe river (Czech Republic). Concentrations of polychlorinated biphenyls (PCBs) and organochlorine pesticides were determined in pooled muscle samples to screen contamination rates of the sampling sites. Induction of the cytochrome P4501A-dependent 7-ethoxyresorufin *O*-deethylase (EROD) activity was used as the biomarker of dioxin-like toxicity, and the activation of microsomal glutathione *S*-transferase (mGST) and *in vitro* susceptibility to lipid peroxidation were chosen as the biochemical markers of oxidative stress in fish. The concentrations of PCBs and DDT metabolites were higher in bream than perch tissues; the EROD activity was higher in perch which were also found to be more susceptible to oxidative damage to cellular membranes measured as Fe/NADPH-enhanced *in vitro* lipid peroxidation. Exposure to dioxin-like contaminants was detected at several sampling sites at which higher concentrations of PCBs and hexachlorobenzene were determined analytically. Both the oxidative stress parameters were increased significantly at two sampling sites. At one of them, however, the results of chemical analyses indicate that the increases in lipid peroxidation and mGST activity were caused by chemicals other than the contaminants monitored within this study. The parameters of oxidative stress can be regarded as important biomarkers of toxicity suited to complete routine chemical analyses and screen the total chemical impact.

biomarkers; PCBs; organochlorines; CYP1A; dioxin toxicity; glutathione; lipid peroxidation; oxidative stress

ABSTRAKT: Biochemické markery dioxinové toxicity a oxidativního stresu byly sledovány v jaterních mikrosomech cejna velkého (*Abramis brama*) a okouna říčního (*Perca fluviatilis*) odchytených v šesti lokalitách řeky Labe. Ve svalovině ryb byly analyticky stanoveny koncentrace polychlorovaných bifenyly (PCB) a organochlorových pesticidů, aby byla zjištěna úroveň kontaminace na odběrových místech. Indukce cytochrom P4501A-dependenční 7-ethoxyresorufin-*O*-deetylázové aktivity (EROD) byla užita jako biomarker toxicity dioxinového typu; aktivace mikrosomální glutathion-*S*-transferázy a lipidní peroxidace *in vitro* byly vybrány jako biochemické markery oxidativního stresu v rybách. V tkáni cejna byly nalezeny vyšší koncentrace PCB a reziduí DDT. EROD aktivita byla vyšší u okouna, který se také ukázal citlivější k oxidativnímu poškození buněčných membrán (stanovenému jako Fe/NADPH-indukovaná lipidní peroxidace *in vitro*). Expozice kontaminanty dioxinového typu byla detekována na odběrových místech, na nichž zároveň byly stanoveny vyšší koncentrace PCB a hexachlorbenzenu. Na dvou odběrových místech byly také zvýšeny oba parametry oxidativního stresu; na jednom z nich však pravděpodobně byla modulace těchto biomarkerů způsobena kontaminanty, které nebyly analyticky sledovány. Stanovení parametrů oxidativního stresu a biomarkeru dioxinové toxicity ukázalo, že data z biochemického monitorování vhodně doplňují rutinní chemické analýzy a sledují celkový impakt chemických látek na organismus.

biomarkery; PCB; organochlorové kontaminanty; CYP1A; dioxinová toxicita; lipidní peroxidace; oxidativní stres

* Supported by the National Agency for Agricultural Research in Prague (Grant No. RE5561).

INTRODUCTION

The objective of biochemical monitoring is to assess actual toxic effects of contamination by measuring responses of specific biochemical parameters (biomarkers) to the most important modes of toxic action. Along with chemical analyses of some major groups of contaminants (Peakall and Walker, 1994; van Gestel and van Brummelen, 1996), this approach to the monitoring of aquatic pollution is becoming a part of ecotoxicological risk assessment. Only some toxicity mechanisms, such as dioxin-like toxicity and oxidative stress (Stegeman and Hahn, 1994; Kelly et al., 1998), have so far been measured *in vivo* to demonstrate chemical stress.

The dioxin-like toxicity includes effects, whose activation is connected with the interaction of the toxicant with a certain cellular receptor (Ah-receptor). This interaction results in increased metabolic activation of promutagens and procarcinogens, increased cellular proliferation, neurotoxicity, endocrine dysbalance, etc. (Safe, 1990; van den Berg et al., 1998). A typical compound with such effects is 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD); the respective biochemical marker is the induction of cytochrome P4501A and an increase in the specific activity of this enzyme, which is often determined as 7-ethoxyresorufin O-deethylase (EROD).

Oxidative stress is defined as a dysbalance between the natural production of prooxidants and the levels of antioxidant defence systems. An overproduction of prooxidants arises from an exposure to noxious xenobiotics. Various modes of action can be involved in this process (Klaunig et al., 1998). Oxidative stress is thus one of the important manifestations of general stress caused by pollutants. The activation of the liver microsomal antioxidative enzyme glutathione *S*-transferase (mGST) is apparently an effective biomarker of an intensification of prooxidative processes pursuant to an exposure to xenobiotics in fish. This activation is an adaptation mechanism; mGST is rather activated by reactive oxygen species than synthesised *de novo* (Moorhouse and Cassida, 1992). Our previous laboratory and field studies (Petřivalský et al., 1997; Machala et al., 1998) have shown that the increase in the activity of hepatic mGST in some fish species correlates well with an intensification of prooxidative processes after an exposure to aquatic contaminants, such as organochlorine pesticides.

Exposures to xenobiotics are followed by changes in the levels of certain enzymes involved in prooxidative processes. Such changes can result in increased susceptibility of membrane lipids to oxidative damage as indicated by the level of Fe(II)/NADPH-induced lipid peroxidation *in vitro* in the hepatic microsomal fraction (Machala et al., submitted for publication).

The objective of this study was to determine the impact of dioxin-like contaminants and to compare levels and modulations of oxidative stress biomarkers in hepat-

ic tissue of bream and perch captured at different sites of the Czech part of the Elbe river.

MATERIAL AND METHODS

Chemicals

7-ethoxyresorufin was obtained from Molecular Probes (Eugene, MI, USA), resorufin, NADPH, bicinchoninic acid, reduced glutathione, 1-chloro-2,4-dinitrobenzene, thiobarbituric acid, and 1,1,3,3-tetraethoxypropane were purchased from Sigma-Aldrich (Prague, Czech Republic). Organochlorine and PCB congeners and isomers were obtained from Dr. Ehrenstorfer (Darmstadt, Germany). All other chemicals were of the highest commercially available purity.

Sampling, chemical and biochemical analyses

Bream (*Abramis brama*) and perch (*Perca fluviatilis*) were captured at six sites of the Elbe river in August 1996. After cervical dislocation, hepatic and muscular tissues were sampled for the measurement of biochemical toxicity markers and chemical analyses, respectively. The samples were frozen on dry ice, transported to lab in less than 24 hr, and stored at -80°C . The age of fish was estimated by scale analysis.

Dorsal muscle samples (approximately 5 g/fish) collected at each site were pooled and homogenised before the analysis. The pooled samples were analysed for the contents of the major classes of persistent chlorinated contaminants using conventional analytical methods including Soxhlet extraction with petroleum ether-acetone, clean-up on a gel permeation column, and fractionation in a HYPERCARB column (Shandon, Runcorn, U.K.). Indicator and mono-*ortho*-chlorinated PCBs (IUPAC Nos. 28, 52, 101, 105, 118, 138, 153, 156, and 180), DDT and their residues, hexachlorobenzene and hexachlorocyclohexane were determined by HRGC/ECD (Machala et al., 2000).

Individual hepatic tissue samples were homogenised and microsomal fraction was separated by differential centrifugation as described previously (Petřivalský et al., 1997). Thereafter, it was stored at -80°C until the biochemical analysis. The EROD activity was determined fluorimetrically (Prough et al., 1978); the final concentration of the substrate 7-ethoxyresorufin was 2 μM . The conjugation activity of microsomal GST was determined spectrofluorometrically with 1-chloro-2,4-dinitrobenzene as the substrate (Moorhouse and Cassida, 1992). All the enzyme activities were determined at 30°C .

Lipid peroxidation was induced *in vitro* by Fe(II)/NADPH (final concentrations of FeSO_4 and NADPH were 0.33 mM and 0.36 mM, respectively) in hepatic microsomes for 30 min and determined spectrophotometrically

with thiobarbituric acid (Uchiyama and Mihara, 1978). Protein concentration was measured using the bicinchoninic acid assay (Smith et al., 1985).

RESULTS AND DISCUSSION

The concentrations of polychlorinated biphenyls (PCBs) and the organochlorine pesticides hexachlorobenzene (HCB), hexachlorohexane (HCH) and DDT and their metabolites were determined in pooled bream and perch muscle samples collected at each of the six sampling sites in the Czech part of the Elbe river. Sampling data and concentrations of the contaminants are summarised in Tab. I.

The results of chemical analyses indicate that fish were contaminated by PCBs already at site 1 situated upstream from the Pardubice industrial area. Significantly higher PCB contamination rates were found at sites 5 and 6, but even here the concentrations in muscle tissue were below maximum value tolerated in the Czech Republic. The concentrations of DDT, incl. its metabolites, and hexachlorobenzene at sites 5 and 6 were also rather high. On the other hand, lindane and other HCH isomers were present at very low concentrations only (maximum of 0.002 µg/kg fresh weight at site 4, data not shown). The bioaccumulation of chlorinated aquatic contaminants was apparently higher in bream than in perch muscle tissues.

Candidate biochemical *in vivo* markers should indicate exposure to or specific noxious effects of aquatic contaminants. Only such biochemical parameters are considered suitable whose modulation due to chemical contamination is sensitive enough. In view of a variety of mechanisms of adverse effects of xenobiotics, one biomarker is insufficient and it will be necessary in the near future to make up a battery of sensitive biomarkers responding to major modes of action of contaminants including dioxin-like toxicity and oxidative stress. In our study, biochemical markers in individual hepatic microsomal fractions of bream and perch from the same sampling sites were induced in a very similar way (Figs. 1–3). Sampling site 1 was taken as a reference site with a low con-

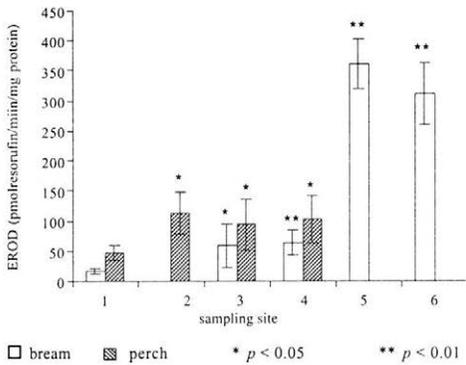
tamination rate. The EROD activity, as the specific biomarker of exposure to planar organic contaminants with dioxin-type toxic potencies, was significantly increased at sites 5 and 6 which were heavily contaminated by persistent chlorinated chemicals. EROD induction was found also at sites 2–4 where only moderate contamination by these compounds was demonstrated by chemical analyses (Fig. 1). This finding agreed well with the results of previous studies where this biomarker was shown to be sensitive to coplanar PCB congeners in a number of fish species (Stegeman and Hahn, 1994). The induction of hepatic microsomal EROD activity in bream was studied as a biomarker of dioxin-like toxicity along with the determination of PCB and Hg concentration in liver tissue of fish collected in the lower course of the Elbe river in Germany (Jedamski-Grymlas et al., 1995). Like in our study, the induction of the EROD activity was associated with higher concentrations of PCBs. However, the EROD induction threshold for PCBs and other dioxin-like contaminants and the induction variability of this biomarker are still to be estimated in fish species coming into consideration for biochemical monitoring. Remarkably, in our study, the EROD activities were significantly higher in perch than in bream (Fig. 1).

Besides the contribution from PCBs, when considering which organic contaminants might induce the EROD activity in bream and perch, HCB has also a potency to dioxin-like activity in the concentration of 20 ng HCB per kg wet weight. Compared with the reference toxicant TCDD, its toxic equivalency factor is 0.0001 (van Birgelen, 1998), which is a value similar to that of mono-*ortho*-chlorinated PCBs (van den Berg et al., 1998). Moreover the presence of other dioxin-like toxicants, halogenated dibenzo-*p*-dioxins or dibenzofurans, which were not determined analytically, must also be considered. In any case, EROD was definitely found to be a suitable parameter complementary to chemical analysis and its induction indicated a risk of contamination by planar dioxin-like contaminants, especially at sites 5 and 6.

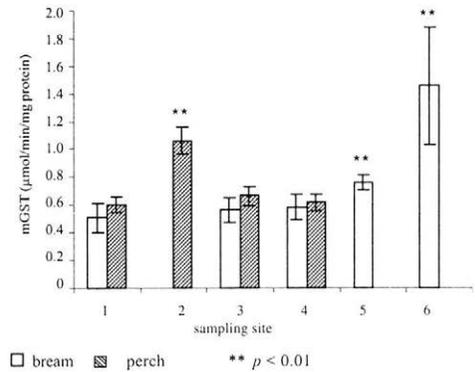
The two oxidative stress parameters under study, namely the microsomal GST activity and the Fe(II)/NADPH-enhanced *in vitro* lipid peroxidation, were significantly increased at sampling sites 2 and 6 (Figs. 2, 3).

I. Sampling sites, number of samples and concentration of contaminants in pooled fish muscle (mg/kg fresh weight). Distances of the sampling sites are expressed in km (measured upstream)

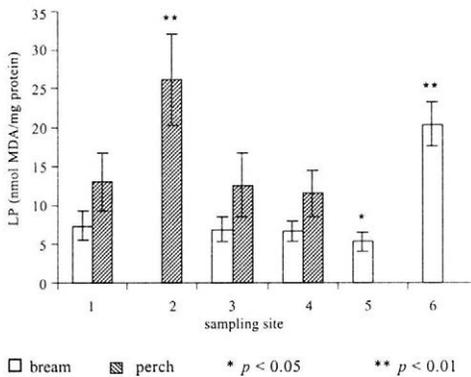
| | Sampling site (km) | Species (n) | Age | Σ PCB | Σ DDT + DDE | HCB |
|---|--------------------|-------------|-----|-------|-------------|-------|
| 1 | Opatovice (262.6) | bream (5) | 5–6 | 0.111 | 0.030 | 0.001 |
| | | perch (8) | 2–5 | 0.038 | 0.010 | 0.001 |
| 2 | Valy (224.6) | perch (7) | 3–4 | 0.096 | 0.008 | 0.002 |
| 3 | Lysá n. L. (150.7) | bream (4) | 3–7 | 0.174 | 0.018 | 0.003 |
| | | perch (3) | 4–7 | 0.069 | 0.008 | 0.002 |
| 4 | Obříství (116.2) | bream (5) | 4–8 | 0.187 | 0.026 | 0.002 |
| | | perch (10) | 5–8 | 0.050 | 0.007 | 0.001 |
| 5 | Děčín (12.0) | bream (4) | 6–9 | 0.307 | 0.093 | 0.023 |
| 6 | Hřensko (0) | bream (4) | 5–7 | 0.251 | 0.087 | 0.028 |



1. Hepatic microsomal CYP1A-dependent 7-ethoxyresorufin *O*-deethylase (EROD) activities in bream and perch collected at six sampling sites in the Elbe river (mean ± SD); for the description of sampling sites and sample sizes see Tab. 1



2. Microsomal glutathione *S*-transferase activities toward 1-chloro-2,4-dinitrobenzene (mGST) in liver of bream and perch in the Elbe river (mean ± SD); for sampling sites see Tab. 1



3. Fe(II)/NADPH-dependent *in vitro* lipid peroxidation in hepatic microsomal fraction of bream and perch determined as thiobarbituric acid-reactive species (mean ± SD); for sampling sites see Tab. 1)

A comparison of analytical data with increased values of the biomarkers at the two sampling sites let us assume that oxidative stress found at site 2 was probably caused by other than the monitored contaminants. Antioxidant enzymes have been proposed as biochemical markers of noxious prooxidative processes for environmental risk assessment (Winston and DiGiulio, 1991; Kelly et al., 1998). The microsomal GST is a membrane-bound antioxidant enzyme. An increased mGST activity was described in marine fish from contaminated littoral sites (Rodríguez-Ariza et al., 1993), carp collected from contaminated lakes (Machala et al., 1997), and rainbow trout exposed to model oxidative stressors (Petřivalský et al., 1997; Machala et al., 1998).

The activation of mGST directly correlates with the increase of prooxidative processes (Petřivalský et al., 1997;

Machala et al., 1998). On the other hand, *in vitro* lipid peroxidation in the hepatic microsomal membrane is not an unambiguous parameter as shown in a recent field study in chub (Machala et al., submitted for publication). Increased or decreased lipid peroxidation *in vitro* can be found in hepatic microsomal samples of fish collected at more contaminated sites. Compared with reference site 1 (Fig. 3), the *in vitro* lipid peroxidation in the present study was lower in the samples collected at site 5 and higher in those collected at site 6.

This difference in the modulation between the two oxidative stress parameters can be explained as a result of increased capacity of antioxidant systems accompanied by a less severe damage to membrane phospholipids (Rodríguez-Ariza et al., 1993). In this study, the lipid peroxidation *in vitro* in the hepatic microsomal fraction was higher in perch than in bream at all the sampling sites; this difference may have been associated with a higher susceptibility of cell membrane to oxidative damage in the former species.

In conclusion, the hepatic microsomal EROD activity was shown to be a suitable biomarker responding to the dioxin-like toxicity in both bream and perch. The activation of mGST was found to be a sensitive biochemical marker of oxidative stress in these fish species. Practical application of Fe(II)/NADPH-enhanced *in vitro* lipid peroxidation as another candidate oxidative stress indicator will require a more detailed explanation of its modulation mechanisms.

Acknowledgements

The authors thank Mrs. Anna Kocová (Research Institute of Fisheries and Hydrobiology, Vodňany) for the help in sampling and Mrs. Eva Gröbnerová and Mrs. Marie Gájová (Veterinary Research Institute, Brno) for the lab work.

REFERENCES

- Jedamski-Grymlas J., Kammann U., Tempelmann A., Karbe L., Siebers D. (1995): Biochemical responses and environmental contaminants in breams (*Abramis brama* L.) caught in the river Elbe. *Ecotoxicol. Environ. Saf.*, *31*, 49–56.
- Kelly S. A., Havrilla C. M., Brady T. C., Abramo K. H., Levin E. D. (1998): Oxidative stress in toxicology: established mammalian and emerging piscine model systems. *Environ. Hlth Persp.*, *106*, 375–384.
- Klaunig J. E., Xu Y., Isenberg J. S., Bachowski S., Kolaja K. L., Jiang A., Stevenson D. E., Walborg E. F. (1998): The role of oxidative stress in chemical carcinogenesis. *Environ. Hlth Persp.*, *106*, 289–295.
- Machala M., Petřivalský M., Nezveda K., Ulrich R., Dušek L., Piačka V., Svobodová Z. (1997): Responses of carp hepatopancreatic 7-ethoxyresorufin *O*-deethylase and glutathione-dependent enzymes to organic pollutants – a field study. *Environ. Toxicol. Chem.*, *16*, 1410–1416.
- Machala M., Drábek P., Neča J., Kolářová J., Svobodová Z. (1998): Biochemical markers for differentiation of exposures to nonplanar polychlorinated biphenyls, organochlorine pesticides, or 2,3,7,8-tetrachlorodibenzo-*p*-dioxin in trout liver. *Ecotoxicol. Environ. Saf.*, *41*, 107–111.
- Machala M., Ulrich R., Neča J., Vykusová B., Kolářová J., Máchová J., Svobodová Z. (2000): Biochemical monitoring of aquatic pollution: indicators of dioxin-like toxicity and oxidative stress in the roach (*Rutilus rutilus*) and chub (*Leuciscus cephalus*) in the Skalnice river. *Vet. Med. – Czech*, *45*, 55–60.
- Moorhouse K. G., Cassida J. E. (1992): Pesticides as activators of mouse liver microsomal glutathione *S*-transferase. *Pest. Biochem. Physiol.*, *44*, 83–90.
- Peakall D. W., Walker C. H. (1994): The role of biomarkers in environmental assessment. *Ecotoxicology*, *3*, 173–179.
- Petřivalský M., Machala M., Nezveda K., Piačka V., Svobodová Z., Drábek P. (1997): Glutathione-dependent detoxifying enzymes in rainbow trout liver: search for specific biochemical markers of chemical stress. *Environ. Toxicol. Chem.*, *16*, 1417–1421.
- Prough R. A., Burke M. D., Mayer R. T. (1978): Direct fluorimetric methods for measuring mixed-function oxidase activity. *Method. Enzymol.*, *52*, 372–377.
- Rodríguez-Ariza A., Peinado J., Pueyo C., López-Barea J. (1993): Biochemical indicators of oxidative stress in fish from polluted littoral areas. *Can. J. Fish. Aquat. Sci.*, *50*, 2568–2573.
- Safe S. (1990): Polychlorinated biphenyls (PCBs), dibenzo-*p*-dioxins (PCDDs), dibenzofurans (PCDFs), and related compounds: environmental and mechanistic considerations which support the development of toxic equivalency factors (TEFs). *Crit. Rev. Toxicol.*, *21*, 51–88.
- Smith P. K., Krohn R. I., Hermanson G. T., Mallia A. K., Gartner F. H., Provenzano M. D., Fujimoto E. K., Goeke N. M., Olson B. J., Klenk D. C. (1985): Measurement of protein using bicinchoninic acid. *Anal. Biochem.*, *150*, 76–85.
- Stegeman J. J., Hahn M. E. (1994). *Biochemistry and molecular biology of monooxygenases: current perspectives on forms, functions, and regulation of cytochrome P450 in aquatic species.* In: Malins D. C., Ostrander G. K. (eds.): *Aquatic Toxicology. Molecular, Biochemical, and Cellular Perspectives.* Boca Raton, FL, USA, Lewis Publishers, CRC Press Inc. 87–206.
- Uchiyama M., Mihara M. (1978): Determination of malondialdehyde precursor in tissues by thiobarbituric acid test. *Anal. Biochem.*, *86*, 271–278.
- Van Birgelen A. P. J. M. (1998): Hexachlorobenzene as a possible major contributor to the dioxin activity of human milk. *Environ. Hlth Perspect.*, *106*, 683–688.
- Van den Berg M., Birnbaum L., Bosveld A. T. C., Brunström B., Cook P., Feeley M., Giesy J. P. et al. (1998): Toxic equivalency factors (TEFs) for PCBs, PCDDs, PCDFs for humans and wildlife. *Environ. Hlth Perspect.*, *106*, 775–792.
- Van Gestel C. A. M., Van Brummelen T. C. (1996): Incorporation of the biomarker concept in ecotoxicology calls for a redefinition of terms. *Ecotoxicology*, *5*, 217–225.
- Winston G. W., Di Giulio R. T. (1991): Prooxidant and antioxidant mechanisms in aquatic organisms. *Aquat. Toxicol.*, *19*, 137–161.

Received: 00–04–17

Accepted after corrections: 00–05–31

Contact Address:

RNDr. Miroslav Machala, CSc., Výzkumný ústav veterinárního lékařství, Hudcova 70, 621 32 Brno, Česká republika
Tel. +420 5 41 32 12 41, fax +420 5 41 21 12 29, e-mail: machala@vri.cz

Změna publikačního jazyka ve vědeckých časopisech ČAZV

Předsednictvo České akademie zemědělských věd přijalo na zasedání dne 6. 4. 2000 usnesení, kde mj. doporučuje změnu publikačního jazyka ve vědeckých časopisech vydávaných pod gescí ČAZV. Předsednictvo navrhuje Vydavatelské radě ČAZV zavést angličtinu jako jediný jazyk ve všech vědeckých časopisech od 1. 1. 2001.

Od 1. 7. 2000 redakce časopisu Veterinární medicína přijímá příspěvky psané pouze v angličtině. Příspěvky přijaté do tohoto termínu budou ještě uveřejněny v češtině a slovenštině.



A change of publication language in Scientific Journals of the Czech Academy of Agricultural Sciences

At its session on the 6th April 2000, the Presidium of the Czech Academy of Agricultural Sciences adopted a resolution recommending, among other things, to change the publication language in scientific journals published under the Academy patronage. The Presidium proposes to the Publishing Board of the Academy to introduce English as the only language in all scientific journals from the 1st January 2001.

The papers written exclusively in English are accepted by the editor's office of the journal Veterinary Medicine - Czech from the 1st July 2000. The papers that were accepted before this date will be published in Czech and Slovak.

EFFECTS OF HEAT-PROCESSING OF MILK ON VALIDATION CHARACTERISTICS OF THE CHARM AIM-96 AND DELVOTEST SP SCREENING TESTS

VLIV TEPELNÉHO OŠETŘENÍ MLÉKA NA CHARAKTERISTIKY VALIDACE SCREENINGOVÝCH TESTŮ CHARM AIM-96 A DELVOTEST SP

J. Schlegelová, D. Ryšánek

Veterinary Research Institute, Brno, Czech Republic

ABSTRACT: Two commercial screening tests (Charm AIM-96 and Delvotest SP) for the detection of antimicrobial residues in milk were evaluated using samples of heat-processed milk. Heating to 85 °C for 5 min had no effect on detection limits for selected antimicrobial agents diluted in milk including penicillin, streptomycin, neomycin, chlortetracycline, cloxacillin, chloramphenicol and sulphadiazine. Heat degradation of the interfering natural inhibitors (lysozyme, IgG) resulted in considerable changes of almost all characteristics of validation of raw and heat-processed milk in both the tests. The value of true specificity increased from 96.55% for Charm AIM-96 and 90.62% for Delvotest SP to 100.0%. The most marked change occurred in the cutoff specificity which is decisive for correct scoring of milk with the concentrations of antimicrobial residues <MRL (maximum residue limit) as negative. Heat processing reduced the number of false positive samples detected by Delvotest SP by up to 18.21% ($P < 0.01$). The characteristics of "cutoff sensitivity" were unaffected. Both raw and heat-processed milk samples containing residues of antimicrobial agents at levels corresponding of MRL were scored correctly by both the tests. Heat inactivation of milk samples before screening tests, used usually for bulk milk samples, is necessary considering the intended checks of individual milk samples for residues of antimicrobial agents.

milk; residues; antimicrobial drug; lysozyme; degradation

ABSTRAKT: Dva komerčně dostupné screeningové testy Charm AIM-96 a Delvotest SP pro stanovení reziduí antimikrobiálních látek v mléce byly evaluovány s přihlédnutím k vlivu tepelného ošetření mléka před testací. Detekční limity testů pro vybrané antimikrobiální látky, a to penicilin, streptomycin, neomycin, chlortetracyklin, cloxacilin, chloramfenikol a sulfadiazin, byly shodné při použití mléka tepelně ošetřeného (85 °C, 5 min) před přípravou vzorků (HB) a po tepelném ošetření vzorků s antimikrobiálními látkami (HA). Tepelná denaturace přirozených inhibitorů v mléce (lysozymu, IgG) způsobila v důsledku diference hodnot charakteristik validace při použití syrového mléka oproti tepelně ošetřenému mléku (HA a HB) u obou testů. Hodnota charakteristiky "true specificity" testů vzrostla pro test Charm AIM-96 z 96,55 % a pro test Delvotest SP z 90,62 % ($P < 0,01$) na 100 % u obou testů. Identifikace vzorků bez reziduí a s reziduí antimikrobiálních látek pod povolenými maximálními limity reziduí (MRL) jako negativní ("cutoff specifity"), byla ovlivněna významně u testu Delvotest SP ($P < 0,05$ a $P < 0,01$) i u testu Charm AIM-96 ($P < 0,05$). Schopnost testů detekovat vzorky s reziduí antimikrobiálních látek nejméně na úrovních MRL ("cutoff sensitivity") zůstala nezměněna. Schopnost testu Delvotest SP odlišit negativní vzorky od vzorků pozitivních se významně zvýšila ($P < 0,05$). Tepelná inaktivace vzorků před provedením screeningových testů, obvykle používaných pro směsné vzorky, je obzvláště nutná pro detekci reziduí antimikrobiálních látek v individuálních vzorcích mléka.

mléko; rezidua; antimikrobiální látka; lysozym; degradace

INTRODUCTION

Treatment of food animals with antimicrobial agents has given rise to problems resulting from the presence of drug residues in edible tissues. Recent surveys carried out in the Czech Republic repeatedly demonstrated the

presence in raw milk of β -lactam, aminoglycoside, and tetracycline antibiotics in up to 0.5% of the samples tested in the central laboratories (SCL, 1999). Research has concentrated not only on adverse effects of such drugs on human health (Dewdney and Edwards, 1984; Kidd, 1995) and attempts to remove from foods (Charm, 1980),

but particularly on the development of effective residue detection methods. In most laboratories (Havlová et al., 1993; Suhren, 1995), routine tests are carried out using microtitre plate methods based on growth inhibition of the test strain C-953 (ATCC 10149-IDF, 1991) of *Bacillus stearothermophilus* var. *calidolactis*, such as Intest – Czech Republic, BR tests – Germany, Delvotest P – The Netherlands, and Charm AIM-96 test – USA. The presence of inhibitory agents is revealed by absence of a colour reaction of an indicator on metabolites of the growing test strain.

Various integrated systems for the monitoring of residues of antimicrobial agents have been suggested in recent years. They include both rapid screening methods without specifying the agent, and tests confirming the identity and determining the concentration of the chemical agent. "US Safe/Tolerance Levels" and "Maximum Residue Levels" (MRL) for drug residues have been laid down in the USA and the European Communities, respectively. Screening methods detecting a broad spectrum of antimicrobial agents at such levels are preferred as the first step of any integrated system of residue monitoring (Heeschen and Suhren, 1995).

The milk processing stage at which the test is to be applied must be considered in the selection of the method. The results of validation and/or operation of the methods can be significantly affected by several factors. Thus, for instance, the results of screening tests for β -lactam antibiotics can be influenced by factors used as parameters for milk quality assessment (total microorganism, coliform bacteria, and milk somatic cell counts), and by characteristics of milk production, such as parity, day of lactation and milking performance (Andrew et al., 1997). Such effects were demonstrated, among other authors, by Van Eenennaam et al. (1993) who used the screening methods CITE* probe (β -lactam), Delvotest-P*, Charm Farm*, Lac Tek*, and the diffusion method with *Bacillus stearothermophilus* var. *calidolactis* to test individual milk samples collected from cows suffering from clinical mastitis. Before the sampling, the cows were treated with drugs containing amoxicillin, cefapirin, or oxytocine. False positive tests were obtained in four tests in samples containing increased milk somatic cell counts.

Another factor influencing the results of screening tests is the presence in milk of natural inhibitors of microbial growth. The occurrence of false positive results of the Delvotest P screening method correlates well with increased concentrations of lactoferrin and lysozyme in milk (Carlsson and Björck, 1989). Therefore, heat inactivation of natural inhibitors has been recommended (IDF, 1991). Such recommendation has not been included in instructions for use provided by the manufacturers of Charm AIM-96 and Delvotest SP, however.

The aim of this study was to verify the detection limits of Charm AIM-96 and Delvotest SP and to assess elementary characteristics of the two tests after heat inactivation of the natural inhibitors in milk samples.

MATERIAL AND METHODS

Milk

Milk intended for the dilution of antimicrobial agents was collected aseptically from a healthy, untreated cow after lactation day 182 of the eighth lactation period. Analyses done during the experimental period (10 days) showed the following values: fat 2.33%; proteins 3.37%; lactose 4.32%; SCC 50 000 to 191 000; pH 6.70 to 6.86; IgG by radial diffusion technique (Mancini et al., 1965) 1.0 to 1.9 mg/ml; lysozyme by radial diffusion technique with *Micrococcus lysodeicticus* (Richter, 1986) 3.8 to > 4.0 μ g/ml.

Samples

Three series of samples were prepared for each of the seven antimicrobial agents and for each day of the experiment as follows: antimicrobial agents diluted in raw milk (RM); antimicrobial agents diluted in raw milk which was subsequently heat-treated (HA); antimicrobial agents diluted in heat-treated milk (HB). The treatment consisted in heating to 85 °C for 5 min and immediate cooling to 20 °C.

Antimicrobial drugs

The antimicrobial agents, their dilution series and approved MRL are summarised in Tab. I. The selection of the agents was done considering the frequency of their use in drugs intended for therapy of mastitis in the Czech Republic. Chloramphenicol is present in one of them, but its use for the treatment of food animals has been limited by the World Health Organisation since 1969 (WHO, 1969). Checks for illegal use are necessary.

Screening tests

Two screening tests for the detection of residues of antimicrobial agents in milk were validated – Charm AIM-96 test (Charm Sciences, Inc., USA) and Delvotest SP (Gist-brocades Food Ingredients Inc., The Netherlands). The manufacturers' instructions for the preparation of microplates and execution of the tests were strictly adhered to.

For the Charm AIM-96 test, the freeze-dried nutrient medium with the pH indicator, and a tablet containing the bacterial culture were reconstituted in the attached solution. This mixture was then pipetted into wells of microtitre plates containing the milk samples to be tested. The plates were then incubated in an appliance supplied by the manufacturer of the test kit.

For the Delvotest SP, tablets of the bacterial culture were put into individual microtitre plate wells containing

nutrient agar with pH indicator. The tablets dissolved after the addition of milk samples into the wells. The plates were incubated in a water bath at 64 °C for 2.5 to 3 h. The length of the incubation period depended on the development of yellow colour in the negative control well.

Raw and heat-treated milk free of microbial agents were used as negative controls which were run in each microtitre plate along with the samples containing the antimicrobial agents. The numbers of the negative controls for Charm AIM-96 were 29 raw milk and 34 heat-treated milk samples. The corresponding numbers for Delvotest SP were 32 and 42, respectively. The original positive control supplied by Charm, containing 4 µg penicillin and 50 µg sulphamethazine per litre, as well as a negative control (heat-treated milk without antibiotics) were run in each plate.

The Charm AIM-96 test and Delvotest SP were repeated 7 and 8 times, respectively.

Evaluation

The evaluation of the reactions was done according to the manufacturers' recommendations with the exception that all samples showing at least a weak reaction were scored as positive. Moreover, the detection limits and detectability of MRL (the potential of the method to detect antimicrobial agents at least at MRL) were assessed.

The following characteristics were calculated for each test and each variant of milk treatment:

$$\text{"true" specificity [\%]} = \left[\frac{TN}{TN+FP} \right] \cdot 100 \quad (1)$$

where:

TN = "true" negative samples (free of antimicrobial agents),

FP = "false" positive samples (free of antimicrobial agents, but yielding positive reactions);

$$\text{"cutoff" specificity [\%]} = \left[\frac{CN}{CN+CFP} \right] \cdot 100 \quad (2)$$

where:

CN = "cutoff" negative samples (free of antimicrobial agents or containing antimicrobial agents at concentrations lower than the "cutoff" value, where "cutoff" = MRL for the given agent);

CFP = "cutoff" false positive samples (free of antimicrobial agents or containing antimicrobial agents at concentrations lower than the "cutoff" value, but showing positive reactions).

The parameters define the potential of the method to identify correctly samples free of antimicrobial agents and samples containing residues of antimicrobial agents at concentrations lower than MRL as negative.

$$\text{"cutoff" sensitivity [\%]} = \left[\frac{CP}{CP+CFN} \right] \cdot 100 \quad (3)$$

where:

CP = "cutoff" positive samples (containing antimicrobial agents at MRL),

CFN = "cutoff" false negative samples (containing antimicrobial agents at the level of MRL, but yielding negative reactions).

The parameter defines the potential of the method to identify samples containing residues of antimicrobial agents at least at MRL as positive.

$$\text{"predictive value" of positive tests [\%]} = \left[\frac{CP}{CP+CFP} \right] \cdot 100 \quad (4)$$

$$\text{"predictive value" of negative tests [\%]} = \left[\frac{CN}{CN+CFN} \right] \cdot 100 \quad (5)$$

$$\text{"cutoff" efficacy [\%]} = \left[\frac{CP+CN}{CP+CFP+CN+CFN} \right] \cdot 100 \quad (6)$$

The parameters define the potential of the method to distinguish samples free of residues of antimicrobial agents and samples containing residues of antimicrobial agents below MRL from samples containing residues at or above MRL.

An ideal test should reach 100% in all the above parameters (Deshpande, 1996).

Differences in the characteristics of the individual tests between the raw milk and heat-treated milk samples were analysed by the χ^2 test (Armitage and Barry, 1987).

RESULTS

A decrease of lysozyme and IgG levels to ≤ 0.2 mg/ml and ≤ 1.0 mg/ml, respectively, was observed after heating the milk to 85 °C for 5 min and immediate cooling to 20 °C.

The ranges of detection limits for the individual antimicrobial agents and the three variants of milk as the diluting medium are summarised in Tab. II. The detection limits corresponding to the lowest value in the dilution series include also limits below this value which were not

I. Selection of antimicrobial agents, dilutions used in the tests and MRL values for milk

| Antimicrobial agent* | Dilution (µg/l) | | | MRL (µg/l) |
|---------------------------------|-----------------|------|-----|------------|
| Benzylpenicillin potassium salt | 6 | 4 | 2 | 4 |
| Streptomycin sulphate | 500 | 200 | 100 | 200 |
| Neomycin sulphate | 500 | 200 | 100 | 500 |
| Chlortetracycline hydrochloride | 500 | 100 | 50 | 100 |
| Cloxacillin sodium salt | 50 | 30 | 10 | 30 |
| Chloramphenicol | 6000 | 1000 | 500 | 0 |
| Sulphadiazine | 500 | 100 | 50 | 100 |

*antimicrobial agents (Sigma-Aldrich Ltd)

II. Detection limits of Charm AIM-96 and Delvotest SP for selected antimicrobial agents

| Antimicrobial agent | Detection limits ($\mu\text{g/l}$) | | | | | |
|---------------------|--------------------------------------|--------------|--------------|-----------------|------------|-------------|
| | Charm AIM-96 | | | Delvotest SP | | |
| | RM | HA | HB | RM | HA | HB |
| Penicillin | 2-4 | 2-4 | 2-4 | 2 | 2 | 2 |
| Streptomycin | 200-500 | ≥ 500 | ≥ 500 | 200- ≥ 500 | ≥ 500 | > 500 |
| Neomycin | 100 | 100 | 100 | 200-500 | ≥ 500 | ≥ 500 |
| Chlortetracycline | 50-500 | 100-500 | 100-500 | 50-500 | 100-500 | 100-500 |
| Cloxacillin | 10-30 | 30 | 30 | 10-50 | 30 | 10-30 |
| Chloramphenicol | 1 000-6 000 | $\geq 6 000$ | $\geq 6 000$ | 500-6 000 | 500-6 000 | 1 000-6 000 |
| Sulphadiazine | 50 | 50 | 50 | 50-100 | 50-100 | 50-100 |

RM – raw milk; HA – milk heat-processed after the addition of the antimicrobial agent; HB – milk heat-processed before the addition of the antimicrobial agent

examined. It can be seen that the detection limits for penicillin were equal to, or lower (Delvotest SP) than MRL (Tab. I). Residues of chloramphenicol were detected by any of the tests only at concentrations $\geq 500 \mu\text{g/l}$. The sensitivity of the two tests for cloxacillin, sulphadiazine, chlortetracycline, and insensitivity for streptomycin ($\geq 500 \mu\text{g/l}$) were identical or almost identical when HA or HB samples were tested. The sensitivity for neomycin was lower in Delvotest SP than in Charm AIM-96 test (200 to ≥ 500 vs. 100 $\mu\text{g/l}$).

Heat treatment of milk did not affect the activity of the antimicrobial agents, as demonstrated by equal detection limits in milk samples treated before or after the addition of the agents (HA vs. HB).

The number of samples containing residues of antimicrobial agents at MRL and scored as positive are given in Tab. III. Both tests reliably identified milk samples containing penicillin, cloxacillin, and sulphadiazine at MRL in both heat-treated milk (100%) and raw milk (75 and 100% for Delvotest SP and Charm AIM-96, respectively), and were insensitive for chloramphenicol (0%). Residues of streptomycin at MRL were detected in only one sample of raw milk. Residues of chlortetracycline were detected by Charm AIM-96 more effectively (43% for all milk treatments) than by Delvotest SP (25, 12, and 25% for RM, HA, and HB, respectively). A considerable dif-

ference between the tests was found in the demonstration of neomycin residues. While Charm AIM-96 identified all samples at the level of 100 $\mu\text{g/l}$ (below MRL) irrespective of the milk treatment, the efficacy of Delvotest SP was only 75, 50 and 50% for RM, HA, and HB, respectively. The detection limit was $\geq 500 \mu\text{g}$ neomycin/l, however.

Tab. IV summarises basic validation characteristics of the screening tests for the three milk treatments, as well as results of statistical analysis of between-treatment differences. All differences were tested, but only those significant at $P < 0.05$ or $P < 0.01$ are presented.

It is evident that Charm AIM-96 identified actually negative RM samples more effectively than Delvotest SP (96% vs. 90%). Both tests identified correctly all heat-treated samples that were free of antibiotics (100% both). Heat treatment contributed to correct identification of actually negative samples, particularly when Delvotest SP ($P < 0.01$) was used, and also to correct identification of "cutoff" negative samples by Charm AIM-96 (76.32 and 77.11%, $P < 0.05$) and particularly by Delvotest SP (83.33 and 85.71%, $P < 0.01$) Charm AIM-96 proved to be more reliable for the detection of antimicrobial agents at MRL than Delvotest SP (63.26 to 65.30% vs. 51.79 to 57.14%) in RM, HA, and HB milk samples. Negative samples were identified more reliably than positive samples

III. Detectability of MRLs by Charm AIM-96 (in 7 repeated assays) and by Delvotest SP (in 8 repeated assays)

| Antimicrobial agent | Percentage of recorded MRL values | | | | | |
|---------------------|-----------------------------------|-----|-----|--------------|-----|-----|
| | Charm AIM-96 | | | Delvotest SP | | |
| | RM* | HA* | HB* | RM | HA | HB |
| Penicillin | 100 | 100 | 100 | 100 | 100 | 100 |
| Streptomycin | 14 | 0 | 0 | 12 | 0 | 0 |
| Neomycin | 100 | 100 | 100 | 75 | 50 | 50 |
| Chlortetracycline | 43 | 43 | 43 | 25 | 12 | 25 |
| Cloxacillin | 100 | 100 | 100 | 75 | 100 | 100 |
| Chloramphenicol | 0 | 0 | 0 | 0 | 0 | 0 |
| Sulphadiazine | 100 | 100 | 100 | 100 | 100 | 100 |

*For abbreviations see Tab. II

IV. Effects of heat-processing on characteristics of validation of the screening tests Charm AIM-96 and Delvotest SP and significance (*P*) of the differences between raw milk (RM) and heat-processed milks (HA, HB)

| Test characteristics | | Tests | | | | | |
|-----------------------------------|-----|--------------|-----------------|-------|--------------|-----------------|-----------------|
| | | Charm AIM-96 | | | Delvotest SP | | |
| | | RM* | HA* | HB* | RM | HA | HB |
| True specificity | (%) | 96.55 | 100 | 100 | 90.62 | 100 | 100 |
| significance | | | | | | | <i>P</i> < 0.01 |
| Cutoff specificity | (%) | 61.97 | 77.11 | 76.32 | 67.50 | 85.71 | 83.33 |
| significance | | | <i>P</i> < 0.05 | | | <i>P</i> < 0.01 | <i>P</i> < 0.05 |
| Cutoff sensitivity | (%) | 65.30 | 63.29 | 63.26 | 57.14 | 51.79 | 53.57 |
| Predictive value of positive test | (%) | 54.24 | 62.00 | 63.27 | 55.17 | 67.44 | 66.67 |
| Predictive value of negative test | (%) | 72.13 | 78.05 | 76.32 | 69.23 | 75.68 | 74.26 |
| Cutoff efficacy | (%) | 63.33 | 71.97 | 71.20 | 63.23 | 73.38 | 71.19 |
| significance | | | | | | <i>P</i> < 0.05 | <i>P</i> < 0.05 |

* For abbreviations see Tab. II

irrespective of the test used (69.23 to 78.05% vs. 54.24 to 67.44%). The efficacy of both tests was almost identical for all the variants of milk treatment, but heat treatment apparently contributed to correct identification particularly when Delvotest SP was used (*P* < 0.05 and *P* < 0.05).

DISCUSSION

The effects of heat treatment of milk samples on validation characteristics of the screening tests Charm AIM-96 and Delvotest SP for the determination of antimicrobial drug residues in milk were investigated. Conventional characteristics of screening tests, including detection limit, "true" specificity, and "predictive value" of positive and negative results, were completed with "cutoff" specificity, "cutoff" sensitivity and "cutoff" efficacy of the tests.

Investigations have demonstrated a high sensitivity of both tests for β -lactam antibiotics. The detection limit for penicillin 2 to 4 $\mu\text{g/l}$ corresponded to data supplied by the manufacturers and results of interlaboratory trials of routine tests (Suhren et al., 1995). Although most of the results were false positive considering MRL laid down with regard to the protection of human health, the high sensitivity of the tests is beneficial for the dairy industry using very sensitive technological cultures. A high efficacy in the detection of β -lactam antibiotic residues at the tolerance and safety levels was demonstrated by comparison with results of liquid chromatography of milk samples collected from cows with clinical mastitis which were collected before, during, and after the treatment with amoxicillin or penicillin. Anderson et al. (1998) demonstrated agreement of results of the screening test and liquid chromatography in up to 94% of the samples.

Charm AIM-96 detected residues of neomycin already at the concentration of 100 $\mu\text{g/l}$, i.e. five times lower than the temporarily accepted MRL (500 $\mu\text{g/l}$). However, microbial cultures used in the dairy industry are more sen-

sitive to aminoglycoside antibiotics being inhibited already at concentrations of 40 to 200 $\mu\text{g/l}$ (Kraack and Tolle, 1967). Neomycin is an active component of several drugs (Vademecum, 1999) intended for the treatment of bovine mastitis. A more frequent occurrence of neomycin residues in milk can therefore be expected. Considering the data given above, Charm AIM-96 should be preferred to Delvotest SP in milk testing, although the result may be false positive in terms of MRL.

The two tests are almost equivalent as far as sensitivity for chlortetracycline and sulphadiazine is concerned. The detection limits for the latter were lower than MRL; this finding applied particularly to Charm AIM-96. Our results agree with data published for similar antimicrobial agents (Suhren et al., 1995; Charm and Zomer, 1995). In our investigations, Charm AIM-96 proved to be less sensitive for the detection of sulphonamide residues than claimed by the manufacturer, but residues below MRL were still detectable.

Chloramphenicol and streptomycin at MRL were undetectable by any of the tests. The same insensitivity was described also in other conventional screening tests (Charm and Zomer, 1995), which means that samples containing residues of these antibiotics at concentrations exceeding MRL are not identified as positive by the tests. A new systemic approach will be necessary to cope with this problem.

Heat treatment had no apparent effects on detection limits of the two tests. Lower values obtained in tests of raw milk can be attributed to the combined action of the antimicrobial agents and natural inhibitors present in milk.

The test characteristics were validated using a system common in the interpretation of results of enzymeimmunoanalytical tests in clinical diagnostics (Deshpande, 1996). The "cutoff value" ("decision value", "reference value") was identical with MRL.

The requirement to detect antimicrobial agents at MRL was met better by the Charm AIM-96 test that identified

reliably all samples containing penicillin, neomycin, cloxacillin, or sulphadiazine. The results obtained by any of the tests in RM samples containing streptomycin can be scored as false positive.

It is evident from the results that heat treatment of samples resulted in changes of all the validation characteristics excepting "cutoff" sensitivity. It is probable that inactivation of lysozyme was responsible for these changes, particularly in Delvotest SP.

The decrease in lysozyme concentration observed in heat-treated samples agrees with the results of Weaver and Kroger (1978) who demonstrated a degradation of this enzyme at 80 °C and higher temperatures. Carlsson and Björck (1987) observed an inhibition of pH shift due to the growth of *Bacillus stearothermophilus* var. *calidolactis* already at concentrations $\geq 0.1 \mu\text{g}$ lysozyme per 1 ml broth. However, the concentration of 1000 $\mu\text{g}/\text{ml}$ was necessary to influence the results of Delvotest P Multi. The authors attribute this difference to different physical and chemical conditions in the agar medium, but point out that synergetic inhibitory activity of lysozyme and lactoferrin at their physiological concentrations in milk can affect the results of Delvotest P.

A decrease of milk lysozyme level due to heat treatment, observed in our experiment, was also described in the experimental study published by Weaver and Kroger (1978). Heat treatment of milk before testing for residues does not cause any significant change in the concentration of antimicrobial agents.

Heat inactivation of milk samples prior to screening, which is usual in bulk milk testing, is necessary particularly for the detection of residues of antimicrobial agents in individual milk samples. Testing of heat-treated samples protects the farmer against losses due to an exclusion from supply of milk yielding a false positive reaction.

A very detailed study of validation of methods for the detection of residues of antimicrobial agents in milk was published by Gardner et al. (1996). Its authors proposed a validation procedure for methods of assessment of individual milk samples which consists of the following four phases: 1. preliminary assessment of sensitivity and specificity; 2. determination of the detection limit; 3. assessment of factors that may cause false positive results; 4. testing under field conditions. For a system of monitoring of antimicrobial agents in milk, they proposed to use in the first phase a highly sensitive method for the initial screening and subsequently an inexpensive and highly specific confirmation method. Our paper is a contribution to the validation of methods used in the third phase of the above procedure.

The validation of screening tests for residues of antimicrobial agents described in this paper is beneficial particularly when testing of individual milk samples at the farm level is considered. In our opinion, such screening at the level of individual samples is an unavoidable step within each integrated monitoring system for residues of antimicrobial drugs. Information on tests suitable for the detection of residues should be provided by drug manufacturers.

Acknowledgement

The authors wish to thank Tru-Test Distributor Ltd for providing the Charm AIM-96 test materials and lending the AIM-96 incubator, Mrs. H. Vlková for technical assistance, and Mrs. P. Hořavová and Mrs. H. Kudláčková for the determination of lysozyme and IgG.

REFERENCES

- Anderson K. L., Moats W. A., Rushing J. E., O'Carroll J. M. (1998): Detection of milk antibiotic residues by use of screening tests and liquid chromatography after administration of amoxicillin or penicillin G in cows with clinical mastitis. *Am. J. Vet. Res.*, 59, 1096–1100.
- Andrew S. M., Frobish R. A., Paape M. J., Maturin L. J. (1997): Evaluation of selected antibiotic residue screening tests for milk from individual cows and examination of factors that affect the probability of false-positive outcomes. *J. Dairy Sci.*, 80, 3050–3057.
- Armitage P., Barry G. (1987): *Statistical methods in medical research*. 2th ed. Oxford, Blackwell Scientific Publications, 559 pp.
- Carlsson N. A., Björck L. (1987): The effect of some indigenous antibacterial factors in milk on the growth of *Bacillus stearothermophilus* var. *calidolactis*. *Milchwissenschaft*, 42, 282–285.
- Carlsson A., Björck L. (1989): Lactoferrin and lysozyme in milk during acute mastitis and their inhibitory effect in Delvotest P. *J. Dairy Sci.*, 72, 3166–3175.
- Charm S. E. (1980): Process for the removal of antibiotic from milk. U. S. Patent 4,238,521. 6 pp.
- Charm S. E., Zomer E. (1995): The evolution and direction of rapid detection/identification of antimicrobial drug residues. In: *Proc. Symp. Residues of Antimicrobial Drugs and Other Inhibitors in Milk*. Kiel, Germany, 224–233.
- Deshpande S. S. (1996): *Enzyme Immunoassay: From Concept to Product Development*. New York, Chapman & Hall, 464 pp.
- Dewdney J. M., Edwards R. G. (1984): Penicillin hypersensitivity – is milk a significant hazard: a review. *J. Royal Soc. Med.*, 77, 866–877.
- Gardner I. A., Cullor J. S., Galey F. D., Sisco W., Salman M., Slenning B., Erb H. N., Tyler J. F. (1996): Alternatives for validation of diagnostic assays used to detect antibiotic residues in milk. *JAVMA*, 209, 46–52.
- Havlová J., Jiěinská E., Hrabová H. (1993): *Mikrobiologické metody v kontrole jakosti mléka a mlékárenských výrobků*. Praha, ÚZPI, 243 pp.
- Heeschen V. H., Suhren G. (1995): IDF integrated detection system for antimicrobials: Introductory statement and practical experiences in Germany. In: *Proc. Symp. Residues of Antimicrobial Drugs and Other Inhibitors in Milk*. Kiel, Germany, 310–318.
- IDF (1991): Detection and confirmation of inhibitors in milk and milk products. *Doc. IDF*, 258, 99 pp.

- Kidd A. R. M. (1995): Potential risk of effects of antimicrobial residues on human gastrointestinal microflora. *Vet. Rec.*, 137, 496.
- Kraack J., Tolle A. (1967): Brillantschwartz-reduktionstest mit *Bacillus stearothermophilus* var. *calidolactis* zum Nachweis von Hemmstoffen in der Milch. *Milchwissenschaft*, 22, 669–673.
- Mancini G., Carbonara A. O., Heremans J. F. (1965): Immunochemical quantitation of antigens by single radial immunodiffusion. *Immunochem.*, 2, 235–238.
- Richter J. (1986): Lysozym, difuzní metody. In: Procházková J., John C. a kol.: Vybrané diagnostické metody lékařské imunologie. Praha, Avicenum. 107–109.
- SCL (1999): Hodnocení jakosti syrového mléka v centrálních laboratořích České republiky v roce 1998. Praha, Sdružení centrálních laboratoří pro hodnocení jakosti mléka. 24 s.
- Suhren G. (1995): Possibilities and limitations of microbiological inhibitor tests. In: Proc. Symp. Residues of Antimicrobial Drugs and Other Inhibitors in Milk. Kiel, Germany. 159–171.
- Suhren G., Heeschen W., Reichmuth J. (1995): Antibiotics testing: Results of IDF – intercomparisons 1989 and 1992. *Bull. IDF*, 305, 18–28.
- Vademecum (1999): Vademecum registrovaných veterinárních přípravků. Praha, Strategie. 1156 s.
- Van Eenennaam A. L., Cullor J. S., Perani L., Gardner I. A., Smith V. L., Dellinger J., Guterbock W. M. (1993): Evaluation of milk antibiotic screening tests in cattle with naturally occurring clinical mastitis. *J. Dairy Sci.*, 76, 3041–3053.
- Weaver G. L., Kroger M. (1978): Lysozyme activity of high-leucocyte-count milk and the effect of heat and potassium dichromate on lysozyme activity. *J. Dairy Sci.*, 61, 1093–1102.
- WHO (1969): Specifications for the identity and purity of food additives and their toxicological evaluation: Some antibiotics. W. H. O. Tech. Rep. Ser. No. 430, Geneva, Switzerland. 49 pp.

Received: 99–02–04

Accepted after correction: 00–05–30

Contact Address:

RNDr. Jarmila Schlegelová, Výzkumný ústav veterinárního lékařství, Hudcova 70, 621 32 Brno, Česká republika
Tel. +420 5 41 32 12 41, fax +420 5 41 21 12 29, e-mail: schlegelova@vri.cz

INSTITUTE OF AGRICULTURAL AND FOOD INFORMATION
Slezská 7, 120 56 Praha 2, Czech Republic
Fax: (00422) 24 25 39 38

In this institute scientific journals dealing with the problems of agriculture and related sciences are published on behalf of the Czech Academy of Agricultural Sciences. The periodicals are published in the Czech or Slovak languages with long summaries in English or in English language with summaries in Czech or Slovak.

Subscription to these journals should be sent to the above-mentioned address.

| Periodical | Number of issues per year |
|--|---------------------------|
| Rostlinná výroba (Plant Production) | 12 |
| Czech Journal of Animal Science (Živočišná výroba) | 12 |
| Veterinární medicína (Veterinary Medicine – Czech) | 12 |
| Zemědělská ekonomika (Agricultural Economics) | 12 |
| Journal of Forest Science | 12 |
| Research in Agricultural Engineering | 4 |
| Plant Protection Science (Ochrana rostlin) | 4 |
| Czech Journal of Genetics and Plant Breeding (Genetika a šlechtění) | 4 |
| Záhradnictví (Horticultural Science) | 4 |
| Czech Journal of Food Sciences (Potravinařské vědy) | 6 |

OSTEOCALCIN: A BIOCHEMICAL MARKER OF BONE FORMATION IN EQUINE MEDICINE

OSTEOKALCIN: BIOCHEMICKÝ MARKER TVORBY KOSTÍ V MEDICÍNĚ KONÍ

P. Grafenau¹, B. Carstanjen², O. M. Lepage²

¹ Janskeho 2, 94 901 Nitra, Slovak Republic

² Département Hippique, École Nationale, Vétérinaire de Lyon, Marcy L'Étoile, France

ABSTRACT: Osteocalcin is the most abundant non-collagenous bone protein. It is produced by osteoblasts and is accepted as a biochemical marker of bone formation. The authors summarize the current knowledge of physicochemical properties, biosynthesis and functional aspects of osteocalcin in various species. In addition, determinants of plasma/serum osteocalcin concentration in the equine species are described. Factors affecting blood osteocalcin concentration recognized in horses are: age, type of horse, circadian rhythm, season, administration of exogenous corticosteroids and general anesthesia. In combination with osteocalcin measurements the kidney function should always be evaluated because it is the main way for osteocalcin clearance. Serum/plasma osteocalcin measurements are used in horses for training evaluation, for diagnostic purpose and to monitor the effect of a medical therapy on equine bone.

horse; osteocalcin; bone metabolism; bone formation

ABSTRAKT: Osteokalcin je nejhogněji se vyskytující nekolagenní kostní bílkovina. Tvoří se v osteoblastech a je chápán jako biochemický marker tvorby kostí. Je předložen přehled současných poznatků o fyzikálně chemických vlastnostech osteokalcinu, jeho biosyntéze a funkčních aspektech v různých druhů zvířat. Kromě toho jsou popsány určující faktory koncentrace osteokalcinu v krevní plazmě nebo séru. Tyto faktory ovlivňují koncentraci osteokalcinu v krvi koní: věk, typ koně, denní rytmus, roční období, podávání exogenních kortikosteroidů a celková anestezie. Spolu se zjišťováním koncentrace osteokalcinu by se vždy měla hodnotit funkce ledvin, protože se jedná o hlavní cestu vylučování osteokalcinu. Zjišťování koncentrace osteokalcinu v krevním séru nebo plazmě se používá k hodnocení fyzické kondice, k diagnostickým účelům a k monitorování vlivu léčebné terapie na kosti koní.

kůň; osteokalcin; kostní metabolismus; tvorba kostí

CONTENTS

1. Introduction
2. General considerations
3. Osteocalcin in equine medicine
 - 3.1. Sample collection and handling of osteocalcin
 - 3.2. Physiological determinants
 - 3.2.1. Age
 - 3.2.2. Gender
 - 3.2.3. Type and breed of horses
 - 3.2.4. Circadian rhythm
 - 3.2.5. Season
 - 3.3. Effect of endogenous or exogenous corticosteroids
 - 3.4. Influence of anesthesia and surgery
 - 3.5. Influence of physical training
 - 3.6. Influence of feeding
 - 3.7. Influence of bone diseases
 - 3.8. Future for bone markers in horses
4. References

OBSAH

1. Úvod
2. Všeobecné úvahy
3. Osteokalcin v medicíně koní
 - 3.1. Odběr vzorků a manipulace s osteokalcinem
 - 3.2. Fyziologicky určující faktory
 - 3.2.1. Věk
 - 3.2.2. Pohlaví
 - 3.2.3. Typ a plemeno koní
 - 3.2.4. Denní rytmus
 - 3.2.5. Roční období
 - 3.3. Vliv endogenních nebo exogenních kortikosteroidů
 - 3.4. Vliv anestezie a chirurgického zákroku
 - 3.5. Vliv fyzické trénovanosti
 - 3.6. Vliv výživy
 - 3.7. Vliv chorob kostí
 - 3.8. Budoucnost markerů kostí u koní
4. Literatura

1. INTRODUCTION

Fatigue failure of bone is the most common cause of musculoskeletal injuries in horses and is an important economic and welfare problem to the equine industry (Estberg et al., 1996). The assessment of bone metabolism and the early detection of bone diseases remain still a difficult problem in horses (Lepage, 1998).

Bone metabolism is characterized by two opposite but complementary activities: bone resorption and bone formation. The rate of bone turnover can be assessed by histomorphometry (Lepage et al., 1996) or by measuring the amount of specific biochemical bone markers, which are released into the bloodstream or into urine (Arnaud, 1996; Christenson, 1997). Bone resorption markers include urinary hydroxyproline, pyridinoline crosslinks, plasma telopeptide of type I collagen and tartrate-resistant acid phosphatase (TRAP). Bone formation markers include serum bone specific alkaline phosphatase (b-AP), propeptides of procollagen type I and osteocalcin (OC; Christenson, 1997).

Until a few years ago, blood measurements of total alkaline phosphatase (total AP) – an ectoenzyme located on the cell surface of osteoblasts – was the most widely used biochemical bone marker (Prockop et al., 1979). But total AP is not specific for bone formation because liver, gastrointestinal tract, placenta, certain tumors and perhaps other sources contribute to its concentration in the bloodstream. Bone specific alkaline phosphatase (b-AP) is much more bone specific than total AP. Nevertheless, b-AP can only be detected in horses in large amounts during the first year of life (Thor n-Tolling, 1988). Research was performed to find a new bone formation marker in horses and this led to OC. Osteocalcin is a small, non-collagenous protein which is specific for bone tissue and dentin. It is accepted as a biochemical marker of bone formation (Ducy et al., 1996).

In this paper, general considerations for the use of OC and the current knowledge of OC in the equine species are described.

2. GENERAL CONSIDERATIONS

The name osteocalcin (*osteo* (Greek): for bone; *calc* (Latin): for lime salts; *in*: for protein) derives from Ca^{2+} affinity of this protein (Hauschka and Carr, 1982) and from the abundance of this protein in bone tissue (Hauschka et al., 1975; Lian et al., 1982; Conn and Termine, 1985). This protein was discovered in chicken (Hauschka et al., 1975) and bovine bone (Price et al., 1976). Osteocalcin is predominantly synthesized by osteoblasts and is then incorporated into the extracellular matrix of bone. A fraction of newly synthesized OC is released into the bloodstream, where it can be measured by radioimmunoassay (RIA; Delmas, 1990) or enzyme linked immuno-specific assay (ELISA; Hyldstrup et al., 1989). Osteocalcin has also been called the vitamin K-depend

ent protein of bone and bone gamma-carboxyglutamic acid protein or BGP (Price et al., 1976; Nishimoto and Price, 1979). The protein contains, depending on the species considered, three Glu residues at positions 17, 21 and 24 in the primary structure of 47–51 amino acid residues (Poser et al., 1980; Linde et al., 1980; Carr et al., 1981; Hauschka et al., 1982). Osteocalcin shows avid binding to solid calcium phosphate mineral surfaces, particularly in the case of hydroxyapatite crystals (Hauschka et al., 1975; Price et al., 1976; Poser and Price, 1979) and has a modest affinity for ionic calcium. The conformation of OC was described by a model of two antiparallel α -helical domains, connected by a peptide segment containing a β -turn (residues 26–29) and stabilized by Cys-23 – Cys-29 disulfide bond (Hauschka and Carr, 1982).

The biosynthesis of OC involves three vitamins: vitamin K for Glu residue formation (Hauschka and Reid, 1978), vitamin C for hydroxylation of prohydroxyproline into hydroxyproline (Hauschka et al., 1989) and vitamin D for stimulation of OC production (Price and Baukol, 1980). It was shown that the mouse genome contains an OC cluster composed of three genes (Rahman et al., 1993; Desbois et al., 1994). The first two, named OC gene 1 (OC1) and OC gene 2 (OC2), are expressed only in bone, while the third one, the OC related gene (ORG), is transcribed in the kidney, where a protein similar to OC has been described and called nefrocalcin (Desbois et al., 1994). Osteocalcin is a product of osteoblasts and odontoblasts (Camarda et al., 1987). The plasma/serum OC concentration correlates with new protein synthesis (Hauschka et al., 1989). Histomorphometric studies have shown that the absence of OC led to an increase in bone formation without impairing bone resorption (Ducy et al., 1996). A variety of other OC properties are resumed in Tab. I.

3. OSTEOCALCIN IN EQUINE MEDICINE

3.1. Sample collection and handling of osteocalcin

For accurate measurement of equine OC, samples are centrifuged within 90 minutes after venous blood collection. Serum/plasma are then stored at $-20^{\circ}C$ in plastic tubes for up to 26 weeks (Hope et al., 1993). Serum may be thawed and refrozen up to 5 times without significant changes (Hope et al., 1993). The suggested optimum time for sampling would be daytime (light period) before exercise, since daytime fluctuations in serum OC concentrations occur within a narrow range (Lepage et al., 1991; Black et al., 1999). In addition, the effect of exercise is still not completely known in horses. Osteocalcin can be measured in equine serum or plasma, using a commercially available bovine RIA-Kit (Incestar Co, Stillwater Minn. USA; DSL, Webster TX, USA; Patterson-Allen et al., 1982; Lepage et al., 1990; Black et al., 1999). The human immunoradiometric assay (IRMA) does not recognize equine OC (Lepage et al., 1997). When interpreting the

| Properties | References |
|--|---|
| Inhibitor of hydroxyapatite precipitation | Menanteau et al. (1982), Price (1985) |
| Informative molecule for $1.25(\text{OH})_2\text{D}$ | Pan and Price (1984), Skjodt et al. (1985) |
| Inhibitor of leukocyte elastase | Hauschka (1985) |
| Informative molecule for Ca^{2+} and Mg^{2+} | Hauschka and Carr (1982) |
| Activator/chemoattractant for cells with bone-resorbing properties | Malone et al. (1982), Mundy and Poser (1983), Chenu et al. (1994) |
| Chemoattractant for other cell types | Lucas et al. (1988), Lucas and Caplan (1988) |

plasma/serum OC concentration in horses, creatinine and/or urea measurements have to be done for renal function evaluation (Lepage et al., 1990). This is based on studies in human medicine, where renal glomerular function impairment induces a variation in the circulating OC concentration (Cole et al., 1985).

3.2. Physiological determinants

3.2.1. Age

An inverse correlation between age and serum OC concentration was observed in horses with an important decrease of the OC level during the first 30 months of life (Lepage et al., 1990). Individuals less than one year of age have mean concentrations of serum OC of 47.3 ng/ml, individuals aged 1.5 to 2.5 years have 35.7 ng/ml and individuals over 3.5 years have a mean OC concentration of 6.7 ng/ml. These data indicate a significant slowdown in the rate of bone formation in adults, compared to foals (Lepage et al., 1990; Price et al., 1995; Black et al., 1999) and a predictive model for serum OC concentration based on 99 standardbred horses aged 0 to 5 years was established: $y = 52.19 - 0.026 \cdot x$ age (x = age expressed in months; $r^2 = 0.76$; Lepage et al., 1992).

3.2.2. Gender

No significant influence of sex on serum OC concentration was observed (Lepage et al., 1992). The study was based on 52 male and 47 female standardbreds aged 36 to 60 months. However, two to three years old female thoroughbreds showed a significantly higher serum OC concentration than the same aged males (Chiappa et al., 1999). The authors explained this with the beginning of puberty in two to three years old thoroughbreds, which can cause a significant difference in OC levels. In another study, four months old quarter horse colts showed higher OC levels than the same aged fillies (Fletcher et al., 1998). This finding was explained by a faster growth rate in males when compared to females (Goyal et al., 1981).

Another study, treating the influence of general anesthesia on plasma OC concentration showed a difference between stallions and geldings/females (Grafenau et al., 1999). The decrease in plasma OC concentration after general anesthesia was significantly lower in stallions.

This discrepancy was not explained and might be due to the small number of stallions in the above mentioned study ($n = 5$).

3.2.3. Type and breed of horses

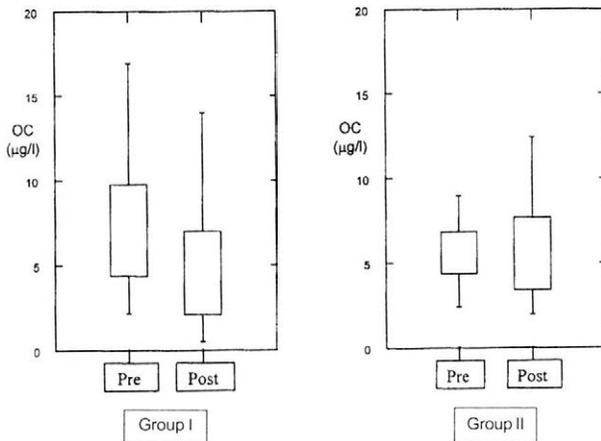
No significant difference was observed between four different breeds of horses (Lepage et al., 1998a). Nevertheless, when taking into account the horses' types, lower serum OC concentrations were observed in draught horses (adjusted least square mean (LSM) = 6.612 $\mu\text{g/l}$) compared to warmblood horses (LSM = 8.596 $\mu\text{g/l}$). Values for draught horses showed the smallest variation, resulting in a higher determination coefficient. A hereditary morphologic and physiologic characteristic of these horses might reflect a lower rate in bone remodeling in draught horses compared to warmblood horses.

3.2.4. Circadian rhythm

The circadian serum OC concentration showed a biphasic pattern in adult horses (Lepage et al., 1991). Levels were constant during daytime (light period) and showed significant variations during the night (dark period), going through a nadir at 8.00 p.m. and through a maximum peak at five o'clock in the morning. Another study indicated similar serum OC concentrations in adult geldings (Black et al., 1999), but not in weanlings. For the understanding of diurnal serum OC variations it is therefore inappropriate to include skeletally mature and immature animals in the same study group (Black et al., 1999). In contrast, a continuous lightening program made the circadian serum OC variations disappear in horses (Hope et al., 1993).

3.2.5. Season

Seasonal variations of serum OC concentration are observed in horses (Maenpää et al., 1988). Serum OC concentration decreased markedly in November and increased in December and January. These data were interpreted as an indication of substantial slowdown of bone formation after transfer from pasture to stables. One explanation given by the author of the study is a decrease in physical activity and a decrease in serum free thyroxin concentration. Other studies showed an increase in serum OC concentration in yearlings and two year old thoroughbred horses in spring-time (Jackson et al., 1998; Price et al., 1997).



1. Plasma osteocalcin concentration shortly before sedation (Pre) and 24 hours later (Post). A significant decrease is observed for the horses who had an anaesthesia (Group I) compared to the control group (Group II)

3.3. Effect of endogenous or exogenous corticosteroids

Endogenous cortisol has been measured in different studies (Hope et al., 1993; Geor et al., 1995; Grafenau et al., 1999). In none of these studies a correlation between OC and cortisol has been described. In addition, neither mean nor individual data demonstrated a consistent relationship between 24-h rhythms in serum cortisol and those of serum OC (Black et al., 1999).

In humans, a dose-dependent effect of exogenous corticosteroids on OC synthesis was documented *in vitro* (Beresford et al., 1984) and *in vivo* (Reid et al., 1986; Patter-son-Buckendahl et al., 1988; Nielsen et al., 1988). In horses, serum OC concentration decreased within 24 to 48 hours after intramuscular administration of triamcinolone ace-tonide at a dosage of 0.09 mg/kg of body weight (Lepage et al., 1993). In seven out of ten horses of that study a return to pretreatment values was observed after 28 days. Pretreatment values for the other three horses were reached between 62 and 150 days later. Serum OC concentration also decreased markedly within 12 hours of intravenous, intramuscular or oral dexamethasone ad-ministration and returned to the pretreatment concentra-tion within three days (Geor et al., 1995). The decrease of serum OC concentration in glucocorticoid-treated horses reflects reduction in the rate of bone formation. There-fore, serum OC concentration may be an early sensitive index of the deleterious effect of long-term corticoste-roid therapy on bone metabolism in the equine species.

3.4. Influence of anesthesia and surgery

General anesthesia and surgery were associated with some metabolic and hormonal changes in horses (Rob-ertson et al., 1987; 1990). Plasma OC concentration de-creased significantly within 24 hours after general

anesthesia (Fig. 1) performed with or without a surgi-cal procedure (Grafenau et al., 1999). The duration of anes-thesia had a significant influence on the degree of plas-ma OC concentration decrease. Plasma OC concentration increased slowly within the first 24 hours after anes-thesia and reached normal values four days after. The de-scribed decrease of serum OC after an orthopedic pro-cedure in a foal (Lepage and Marcoux, 1991) should be attributed to the effect of general anesthesia and probably not to the surgical therapy.

3.5. Influence of physical training

One of the fundamental elements to get horses with strong bones is a correct exercise. Indeed the bone tries to change its shape when getting strains. This adapta-tion to strains is performed by modeling and remodeling activities. The main problem in horses is to find a sensi-tive way to individually monitor exercise based on ob-jective parameters. Therefore studies were performed to detect the influence of exercise on biochemical bone markers.

The effect of treadmill exercise on the cortex of the third metacarpal bone of young horses has been described (Buckingham and Jeffcott, 1991; McCarthy and Jeffcott, 1992). In that study histomorphometric evaluation of the dorsal cortical bone showed extensive modeling in an exercised group of horses and it was shown that the se-rum OC concentration was lower after 14 weeks of exer-cise compared to horses of the unexercised group. These results are in accordance with another study, where two years old thoroughbred mares were divided in a tread-mill-exercised group and a control group. A significantly lower serum OC concentration was observed in the exer-cised group within the first month compared to a control group (Jackson et al., 1998).

In another study stall housed Arabian yearlings showed a lower serum OC concentration at day 14 compared to a pasture held control group. Following day 14 the serum OC concentration of the stalled horses returned to baseline. The authors attribute these variations to a transient slowdown in the rate of osteoblastic activity (Hoekstra et al., 1999). A recent study also revealed that three months deconditioning did not affect serum OC concentrations (Porr et al., 1998).

3.6. Influence of feeding

The effect on serum OC concentration of a forced exercise (longeing) combined or not with oral glucosamine supplementation has been studied (Fenton et al., 1999). Exercised horses had lower concentrations of serum OC compared to not exercised horses. Not exercised, supplemented horses tended to have higher serum OC levels, when compared to exercised non-supplemented and exercised supplemented horses (Fenton et al., 1999). In another study, foals fed a low-copper diet had higher serum OC levels as their withers height increased, compared to a control group (Hurtig et al., 1991). Finally it has been shown that highly exercised Arabian horses taken out of training and fed either a low calcium diet or a high calcium diet during a three months deconditioning period had no influence on their serum OC concentration (Porr et al., 1998).

3.7. Influence of bone diseases

Serum OC concentration increased slightly in horses with developmental orthopedic diseases (Hurtig et al., 1991). Copper-restricted foals may have had continuing stimulation of osteoblasts after their period of fastest growth due to increased modeling of defective bone (Hurtig et al., 1991).

A comparison between variations of serum OC and AP levels in a four months old foal before resection of a wide exostosis on the third metacarpus showed higher serum OC concentrations before surgery (107 ng/ml) and 18 days post-operatively (215 ng/ml; Lepage and Marcoux, 1991). But total AP concentrations were in the normal range of values during the whole time. The results for serum OC were compared with the mean values of eight same aged healthy foals (59 ± 13 ng/ml). A 16 months old Paint horse yearling presented with generalized dyschondroplasia and vertebral epiphysiolysis, showed serum OC levels (17.9 ng/ml) of about half the mean serum OC level of age-matched individuals (Girard et al., 1997). Serum OC levels slightly below normal (31.88 ng/ml) were observed in a seven months old foal which developed a juvenile mandibular ossifying fibroma (Lepage and Davis, 1992). This decrease was strongly pronounced during the first days after bilateral mandibulectomy (15.44 ng/ml). We interpreted the decrease by a stress

response due to surgery. Indeed, we showed in another study that corticosteroids diminished serum OC in horses (Lepage et al., 1993). Horses with chronic obstructive pulmonary disease showed normal serum OC concentrations.

3.8. Future for bone markers in horses

In horses, an evaluation of fracture risk, a response to medical treatment of the skeleton and training-monitoring will be possible, if veterinarians are able to interpret bone mineral density changes or other skeletal parameters reflecting the bone metabolism (Ducy et al., 1996). A simple blood sample will then allow the veterinarian to have more accuracy in predicting and evaluating bone pathologies and will also monitor the response of equine bone to physical training or to a medical treatment.

Osteocalcin is reflecting bone formation and since each biochemical marker reflects a different physiological process in bone, it would enhance the diagnostic power, if we could select a group of markers. This was already observed in a previous study with the concomitant use of serum ICTP and serum OC to evaluate bone metabolism in draught and warmblood horses (Lepage et al., 1998a).

But it should always be remembered that biochemical bone markers do not provide information about skeletal mass, elasticity or architecture, which are better evaluated by densitometric techniques like quantitative ultrasonography (Lepage et al., 1998b), scintigraphy (Tucker et al., 1998) or photon absorptiometry (Jeffcott et al., 1988).

4. REFERENCES

- Arnaud C. D. (1996): Osteoporosis: Using bone markers for diagnosis and monitoring. *Geriatrics*, 51, 24–30.
- Beresford J. N., Gallagher J. A., Poser J. W., Russell R. G. (1984): Production of osteocalcin by human bone cells *in vitro*. Effects of 1,25 (OH)₂D₃, 24,25(OH)₂D₃, parathyroid hormone and glucocorticoids. *Metab. Bone Dis. Relat. Res.*, 5, 229–234.
- Black A., Schoknecht P. A., Ralston S. L., Shapses S. A. (1999): Diurnal variation and age differences in biochemical markers of bone turnover in horses. *J. Anim. Sci.*, 77, 75–83.
- Buckingham S. H., Jeffcott L. B. (1991): Skeletal effects of a long term submaximal exercise program on standardbred yearlings. *Equine Exercise Physiol.*, 3, 411–418.
- Camarda A. J., Butler W. T., Finkelmann R. D., Nanci A. (1987): Immunocytochemical localization of gamma-carboxyglutamic acid-containing proteins (osteocalcin) in rat bone and dentin. *Calcif Tissue Int.*, 40 (6), 349–355.
- Carr S. A., Hauschka P. V., Biemann K. (1981): Gas chromatographic mass spectrometric sequence determination of osteocalcin, a gamma-carboxyglutamic acid-containing protein from chicken bone. *J. Biol. Chem.*, 256 (19), 9944–9950.

- Chenu C., Colucci S., Grano M., Zigrino P., Barattolo R., Zamboni G., Baldini N., Vergnaud P., Delmas P. D., Zallone A. Z. (1994): Osteocalcin induces chemotaxis, secretion of matrix proteins, and calcium-mediated intracellular signaling in human osteoclast-like cells. *J. Cell. Biol.*, 127 (4), 1149-1158.
- Chiappa A., Gonzalez G., Fradinger E., Iorio G., Ferretti J. L., Zanchetta J. (1999): Influence of age and sex in serum osteocalcin levels in thoroughbred horses. *Arch. Physiol. Biochem.*, 107 (1), 50-54.
- Christenson R.H. (1999): Biochemical markers of bone metabolism: an overview. *Clin Biochem.*, 30 (8), 573-593.
- Cole D. E., Carpenter T. O., Gundberg C. M. (1985): Serum osteocalcin concentrations in children with metabolic bone disease. *J. Pediatr.*, 106 (5), 770-776.
- Conn K. M., Termine J. D. (1985): Matrix protein profiles in calf bone development. *Bone*, 6, 33-36.
- Delmas P. D. (1990): Biochemical markers of bone turnover for the clinical assessment of metabolic bone disease. *Endocrin. Metab. Clin. North. Am.*, 19 (1), 1-18.
- Desbois C., Hogue D. A., Karsenty G. (1994): The mouse osteocalcin gene cluster contains three genes with two separate spatial and temporal patterns of expression. *J. Biol. Chem.*, 269 (2), 1183-1190.
- Ducy P., Desbois C., Boyce B., Pinero G., Story B., Dunstan C., Smith E., Bonadio J., Goldstein S., Gundberg C., Bradley A., Karsenty G. (1966): Increased bone formation in osteocalcin-deficient mice. *Nature*, 382 (6590), 448-452.
- Estberg L., Stover S. M., Gardner I. A., Drake C. M., Johnson B., Ardans A. (1996): High-speed exercise history and catastrophic racing fracture in thoroughbreds. *Am. J. Vet. Res.*, 57 (11), 1549-1555.
- Fenton J. I., Orth M. W., Chlebik-Brown K. A., Nielsen B. D., Corn C. D., Waite K. S., Caron J. P. (1999): Effect of longeing and glucosamine supplementation on serum markers of bone and joint metabolism in yearling quarter horses. *Can. J. Vet. Res.*, 63, 288-291.
- Fletcher K. L., Topliff D. R., Freeman D. W., Cooper S. R., Looper M. L. (1998): Effects of sex and weaning on serum osteocalcin concentration in horses. *Anim. Sci. Res. Rep.*, Oklahoma State University, 965, 121-124.
- Geor R., Hope E., Lauper L., Piela S., Klassen J., King J., Murphy M. (1995): Effect of glucocorticoids on serum osteocalcin concentration in horses. *Am. J. Vet. Res.*, 56 (9), 1201-1205.
- Girard C., Lepage O. M., Rossier Y. (1997): Multiple vertebral osteochondrosis in a foal. *J. Vet. Diagn. Invest.*, 9 (4), 436-438.
- Goyal H. O., MacCallum F. J., Brown M. P., Delack J. B. (1981): Growth rates at the extremities of limb bones in young horses. *Can. Vet. J.*, 22 (2), 1-33.
- Grafenau P., Eicher R., Uebellhart B., Tschudi P., Lepage O. M. (1999): General anesthesia decreases osteocalcin plasma concentrations in horses. *Equine Vet. J.*, 31 (6), 533-536.
- Hausehka P. V. (1985): Osteocalcin and its functional domains. In: Butler W. T., Birmingham A.L. (eds.): *Chemistry and Biology of Mineralized Tissue*. Ebsco Media, 149-158.
- Hausehka P. V., Carr S.A. (1982): Calcium-dependent alpha-helical structure in osteocalcin. *Biochemistry*, 21 (10), 2538-2547.
- Hausehka P. V., Reid M. L. (1978): Vitamin K dependence of a calcium-binding protein containing gamma-carboxyglutamic acid in chicken bone. *J. Biol. Chem.*, 253 (24), 9063-9068.
- Hausehka P. V., Lian J. B., Gallop P.M. (1975): Direct identification of calcium-binding amino acid gamma-carboxyglutamate, in mineralized tissue. *Proc. Natl. Acad. Sci. USA*, 72 (10), 3925-3929.
- Hausehka P. V., Carr S. A., Biemann K. (1982): Primary structure of monkey osteocalcin. *Biochemistry*, 21 (4), 638-642.
- Hausehka P. V., Lian J. B., Cole D. E., Gundberg C. M. (1989): Osteocalcin and matrix Gla protein: vitamin K-dependent proteins in bone. *Physiol. Rev.*, 69 (3), 990-1047.
- Hoekstra K. E., Nielsen B. D., Orth D. S., Rosenstein D. S., Schott H. C., Shelle J. E. (1999): Comparison of bone mineral content and biochemical markers of bone metabolism in stall- vs. pasture-reared horses. *Equine Vet. J.*, 30, (Suppl.), 601-604.
- Hope E., Johnston S. D., Hegstad R. L., Geor R. J., Murphy M. J. (1993): Effects of sample collection and handling on concentration of osteocalcin in equine serum. *Am. J. Vet. Res.*, 54 (7), 1017-1020.
- Hurtig M. B., Green S. L., Dobson H., Burton J. (1991): Defective bone and cartilage in foals fed a low-copper diet. *Proc. AAEP*, 36, 637-643.
- Hylldstrup L., Pyke C., Clemmensen I. (1989): Determinations of osteocalcin in serum by enzyme-linked immunosorbent assay (ELISA). *J. Bone Min. Res.*, 4, (Suppl. 1), 980.
- Jackson B., Eastell R. E., Wilson A. M., Lanyon L. E., Goodship A. E., Price J.S. (1998): The effect of exercise on biochemical markers of bone metabolism and insulin-like growth factor-I in two year old thoroughbreds. *J. Bone Min. Res.*, 13, 521.
- Jeffcott L. B., Buckingham S. H., McCarthy R. N., Cleeland J. C., Scotti E., McCartney R.N. (1988): Non invasive measurement of bone: A review of clinical and research applications in the horse. *Equine Vet. J.*, 19, (Suppl. 6), 71-79.
- Lepage O. M. (1998): Contribution à l'étude du tissu osseux équin. [Habilitationsschrift.] Universität Bern, Switzerland.
- Lepage O. M., Davis D. M. (1992): Fibrome ossifiant mandibulaire juvénile du cheval: Diagnostic et traitement. *Prat. Vet. Eq.*, 24, 209-212.
- Lepage O. M., Marcoux M. (1991): Comparison of serum osteocalcin concentration and alkaline phosphatase activity as indicators of bone activity in a foal following surgery. *Can. Vet. J.*, 32, 239-240.
- Lepage O. M., Marcoux M., Tremblay A. (1990): Serum osteocalcin or bone Gla-protein, a biochemical marker for bone metabolism in horses: differences in serum levels with age. *Can. J. Vet. Res.*, 54 (2), 223-226.
- Lepage O. M., DesCôteaux L., Marcoux M., Tremblay A. (1991): Circadian rhythms of osteocalcin in equine serum.

- Correlation with alkaline phosphatase, calcium, phosphate and total protein levels. *Can. J. Vet. Res.*, 55 (1), 5–10.
- Lepage O. M., Marcoux M., Tremblay A., Dumas G. (1992): Sex does not influence serum osteocalcin levels in standardbred horses at different ages. *Can. J. Vet. Res.*, 56 (4), 379–381.
- Lepage O. M., Laverty S., Marcoux M., Dumas G. (1993): Serum osteocalcin concentration in horses treated with triamcinolone acetonide. *Am. J. Vet. Res.* 54 (8), 1209–1212
- Lepage O. M., Perron-Lepage M. F., Francois R. J. (1996): Description histologique de l'os normal chez le poney Shetland. *Prat. Vet. Eq.*, 28, 269–277.
- Lepage O. M., Eicher R., Uebelhart B., Tschudi P. (1997): Influence of type and breed of horse on serum osteocalcin concentration, and evaluation of the applicability of a bovine radioimmunoassay and a human immunoradiometric assay. *Am. J. Vet. Res.*, 58 (6), 574–578.
- Lepage O. M., Hartmann D., Eicher R., Uebelhart B., Tschudi P., Uebelhart D. (1998a): Biochemical markers of bone metabolism in draught and warmblood horses. *Vet. J.*, 156 (3), 169–175.
- Lepage O. M., Whitton C., Danieli A., Niv Y. (1998b): Multi-site quantitative ultrasound measurements in horses: Preliminary results for metacarpal bone assessment. *Bone*, 23, S236.
- Lian J. B., Rouffosse A. H., Reit B., Glimcher M. J. (1982): Concentrations of osteocalcin and phosphoprotein as a function of mineral content and age in cortical bone. *Calcif. Tissue Int.*, 34, (Suppl. 2), S82–S87.
- Linde A., Bhowm M., Butler W.T. (1980): Noncollagenous proteins of dentin. A re-examination of proteins from rat incisor dentin utilizing techniques to avoid artifacts. *J. Biol. Chem.*, 255 (12), 5931–5942.
- Lucas P. A., Caplan A. I. (1988): Chemotactic response of embryonic limb bud mesenchymal cells and muscle-derived fibroblasts to transforming growth factor-beta. *Connect. Tissue Res.*, 18 (1), 1–7.
- Lucas P. A., Price P. A., Caplan A.I. (1988): Chemotactic response of mesenchymal cells, fibroblasts and osteoblast-like cells to bone G1A protein. *Bone*, 9 (5), 319–323.
- Maenpää P. E., Pirskanen A., Koskinen E. (1988): Biochemical indicators of bone formation in foals after transfer from pasture to stables for winter months. *Am. J. Vet. Res.*, 49 (11), 1990–1992.
- Malone J. D., Teitelbaum S. L., Hauschka P. V., Kahn A.J. (1982): Presumed osteoclast precursors (monocytes) recognize two or more regions of osteocalcin. *Calcif. Tissue Int.*, 34, (Suppl.), 79A.
- McCarthy R. N., Jeffcott L. B. (1992): Effects of treadmill exercise on cortical bone in the third metacarpus of young horses. *Res. Vet. Sci.*, 52 (1), 28–37.
- Menanteau J., Neuman W. F., Neuman M. W. (1982): A study of bone proteins which can prevent hydroxyapatite formation. *Metab. Bone Dis. Relat. Res.*, 4 (2), 157–162.
- Mundy G. R., Poser J.W. (1983): Chemotactic activity of the gamma-carboxyglutamic acid-containing protein in bone. *Calcif. Tissue Int.*, 35 (2), 164–168.
- Nielsen H. K., Charles P., Mosekilde L. (1988): The effect of single oral doses of prednisone on the circadian rhythm of serum osteocalcin in normal subjects. *J. Clin. Endocrinol. Metab.*, 67 (57), 1025–1030.
- Nishimoto S. K., Price P. A. (1979): Proof that the gamma-carboxyglutamic acid containing bone protein is synthesized in calf bone. Comparative synthesis rate and effect of coumadin on synthesis. *J. Biol. Chem.*, 254 (2), 437–441.
- Pan L. C., Price P. A. (1984): The effect of transcriptional inhibitors on the bone gamma-carboxyglutamic acid protein response to 1,25-dihydroxyvitamin D3 in osteosarcoma cells. *J. Biol. Chem.*, 259 (9), 5844–5847.
- Patterson-Allen P., Brautigam C. E., Grindeland R. E., Asling C. W., Callaham P. X. (1982): A specific radioimmunoassay for osteocalcin with advantageous species cross-reactivity. *Anal. Biochem.*, 120 (1), 1–7.
- Patterson-Buckendahl P. E., Grindeland R. E., Shakes D. C., Morey-Holton E. R., Cann C. E. (1988): Circulating osteocalcin in rats is inversely responsive to changes in corticosterone. *Am. J. Physiol.*, 254, R828–R833.
- Porr C. A., Kronfeld D. S., Lawrence L. A., Pleasant R. S., Harris P.A. (1998): Deconditioning reduces mineral content of the third metacarpal bone in horses. *J. Anim. Sci.*, 76 (7), 1875–1879.
- Poser J. W., Price P.A. (1979): A method for decarboxylation of gamma-carboxyglutamic acid in proteins. Properties of the decarboxylated gamma-carboxyglutamic acid protein from calf bone. *J. Biol. Chem.*, 254 (2), 431–436.
- Poser J. W., Esch F. S., Ling N. C., Price P. A. (1980): Isolation and sequence of the vitamin K-dependent protein from human bone. Undercarboxylation of the first glutamic acid residue. *J. Biol. Chem.*, 255 (18), 8685–8691.
- Price P. A. (1985): Vitamin K-dependent formation of bone G1A (osteocalcin) and its function. *Vitam. Horm.*, 42, 65–108.
- Price P. A., Baukol S. A. (1980): 1,25 Dihydroxyvitamin D3 increases synthesis of vitamin K-dependent bone protein by osteosarcoma cells. *J. Biol. Chem.*, 255 (24), 11660–11663.
- Price P. A., Otsuka A. A., Poser J. W., Kristaponis J., Raman N. (1976): Characterization of gamma-carboxyglutamic acid-containing protein from bone. *Proc. Natl. Acad. Sci. USA*, 73 (5), 1447–1451.
- Price J. S., Jackson B., Eastell R., Goodship A. E., Blumsohn A., Wright I., Stoneham S., Lanyon L. E., Russell R.G. (1995): Age related changes in biochemical markers of bone metabolism in horses. *Equine Vet. J.*, 27 (3), 201–207.
- Price J. S., Jackson B., Gray J., Wright I. M., Harris P., Russell R. G. G., Eastell R. E., McIlwraith C. W., Ricketts S. W., Lanyon L. E. (1997): Serum levels of molecular markers in growing horses: the effects of age, season, and orthopedic disease. *Transactions Orthopaed. Res. Soc.*, 22, 587.
- Prockop D. J., Kivirikko K. I., Truderman L., Guzman N. A. (1979): The biosynthesis of collagen and its disorders. *N. Engl. J. Med.*, 301 (1), 13–23.
- Rahman S., Oberdorf A., Montecino M., Tanhauser S. M., Lian J. B., Stein G. S., Lapis G. S., Stein L. J. (1993):

- Multiple copies of bone specific osteocalcin gene in mouse and rat. *Endocrinology*, 133 (6), 3050–3053.
- Reid I. R., Chapman G. E., Fraser T. R., Davies A. D., Surus A. S., Meyer J., Huq N. L., Ibbertson H. K. (1986): Low serum osteocalcin levels in glucocorticoid-treated asthmatics. *J. Clin. Endocrinol. Metab.*, 62 (2), 379–383.
- Robertson S. A. (1987): Some metabolic and hormonal changes associated with general anesthesia and surgery in the horse. *Equine Vet. J.*, 19 (4), 288–294.
- Robertson S. A., Steele C. J., Chen C. L. (1990): Metabolic and hormonal changes associated with arthroscopic surgery in the horse. *Equine Vet. J.*, 22 (5), 313–316.
- Skjodt H., Gallagher J. A., Beresford J. N., Couch M., Poser J. W., Russel R. G. (1985): Vitamine D metabolites regulate osteocalcin synthesis and proliferation of human bone cells *in vitro*. *J. Endocrinol.*, 105, 391–396.
- Thorèn-Tolling K. (1988): Serum alkaline phosphatase isoenzymes in the horse-variation with age, training and different pathological conditions. *J. Vet. Med.*, 35 (1), 13–23.
- Tucker R. L., Schneider R. K., Sondhof A. H., Ragel C. A., Tyler J. W. (1998): Bone scintigraphy in the diagnosis of sacroiliac injury in twelve horses. *Equine Vet. J.*, 30 (5), 390–395.

Received: 99–11–23

Accepted after corrections: 00–04–27

Contact Address:

Olivier M. Lepage, DMV, PD, MSc, DES, Dipl. ECVS, École Nationale Vétérinaire de Lyon, Département Hippique, B.P. 83, F-69280 Marcy L'Étoile, France

Tel. + 33 478 87 26 77, Fax + 33 478 87 26 75, e-mail: o.lepage@vet-lyon.fr

POKYNY PRO AUTORY

Časopisy uveřejňují původní vědecké práce, krátká sdělení a výběrově i přehledné referáty, tzn. práce, jejichž podkladem je studium literatury a které shrnují nejnovější poznatky v dané oblasti. Práce jsou uveřejňovány v češtině, slovenštině nebo angličtině. Rukopisy musí být doplněny krátkým a rozšířeným souhrnem. Časopis zveřejňuje i názory, postřehy a připomínky čtenářů ve formě kurzívy, glosy, dopisu redakci, diskusního příspěvku, kritiky zásadního článku apod., ale i zkušenosti z cest do zahraničí, z porad a konferencí.

Autoři jsou plně odpovědní za původnost práce a za její věcnou i formální správnost. K práci musí být přiloženo prohlášení o tom, že práce nebyla publikována jinde.

O uveřejnění práce rozhoduje redakční rada časopisu, a to se zřetelem k lektorským posudkům, vědeckému významu a přínosu a kvalitě práce. Redakce přijímá práce imprinované vedoucím pracoviště nebo práce s prohlášením všech autorů, že se zveřejněním souhlasí.

Rozsah původních prací nemá přesáhnout 10 stran psaných na stroji včetně tabulek, obrázků a grafů. V práci je nutné používat jednotky odpovídající soustavě měrových jednotek SI.

Rukopis má být napsán na papíře formátu A4 (30 řádek na stránku, 60 úhozů na řádku, mezi řádky dvojitě mezery). K rukopisu je vhodné přiložit disketu s textem práce, popř. s grafickou dokumentací pořízenou na PC s uvedením použitelného programu. Tabulky, grafy a fotografie se dodávají zvlášť, nepodlepují se. Na všechny přílohy musí být odkazy v textu.

Pokud autor používá v práci zkratky jakéhokoliv druhu, je nutné, aby byly alespoň jednou vysvětleny (vypsány), aby se předešlo omylům. V názvu práce a v souhrnu je vhodné zkratky nepoužívat.

Název práce (titul) nemá přesáhnout 85 úhozů a musí dát přesnou představu o obsahu práce. Jsou vyloučeny podtitulky článků.

Krátký souhrn (Abstrakt) musí vyjádřit všechno podstatné, co je obsaženo v práci, a má obsahovat základní číselné údaje včetně statistických hodnot. Nemá překročit rozsah 170 slov. Je třeba, aby byl napsán celými větami, nikoliv heslovitě.

Rozšířený souhrn prací v češtině nebo slovenštině je uveřejňován v angličtině, měly by v něm být v rozsahu cca 1–2 strojopisných stran komentovány výsledky práce a uvedeny odkazy na tabulky a obrázky, popř. na nejdůležitější literární citace. Je vhodné jazyk (včetně názvu práce a klíčových slov) dodat v angličtině, popř. v češtině či slovenštině jako podklad pro překlad do angličtiny.

Literární přehled má být krátký, je třeba uvádět pouze citace mající úzký vztah k problému. Tato úvodní část přináší také informaci, proč byla práce provedena.

Metoda se popisuje pouze tehdy, je-li původní, jinak postačuje citovat autora metody a uvádět jen případné odchylky. Ve stejné kapitole se popisuje také pokusný materiál a způsob hodnocení výsledků.

Výsledky tvoří hlavní část práce a při jejich popisu se k vyjádření kvantitativních hodnot dává přednost grafům před tabulkami. V tabulkách je třeba shrnout statistické hodnocení naměřených hodnot. Tato část by neměla obsahovat teoretické závěry ani dedukce, ale pouze faktické nálezy.

Diskuse obsahuje zhodnocení práce, diskutuje se o možných nedostatecích a výsledky se konfrontují s údaji publikovanými (požaduje se citovat jen ty autory, jejichž práci mají k publikované práci bližší vztah). Je přípustné spojení v jednu kapitolu spolu s výsledky.

Literatura citovaná v textu práce se uvádí jménem autora a rokem vydání. Do seznamu se zařadí jen publikace citované v textu. Citace se řadí abecedně podle jména prvních autorů.

Klíčová slova mají umožnit vyhledání práce podle sledovaných druhů zvířat, charakteristik jejich zdravotního stavu, podmínek jejich chovu, látek použitých k jejich ovlivnění apod. Jako klíčová slova není vhodné používat termíny uvedené v nadpisu práce.

Na zvláštním listě uvádí autor plné jméno (i spoluautorů), akademické, vědecké a pedagogické tituly a podrobnou adresu pracoviště s PŠČ, číslo telefonu a faxu, popř. e-mail.

Úplné znění pokynů pro autory s dodatky najdete na URL adrese <http://www.vri.cz> nebo <http://www.uzpi.cz>

For full text of instructions for authors see <http://www.vri.cz> or <http://www.uzpi.cz>

INSTRUCTIONS FOR AUTHORS

Original scientific papers, short communications, and selectively reviews, that means papers based on the study of technical literature and reviewing recent knowledge in the given field, are published in this journal. Published papers are in Czech, Slovak or English. Each manuscript must contain a short or a longer summary. The journal also publishes readers' views, remarks and comments in form of a text in italics, gloss, letter to the editor, short contribution, review of a major article, etc., and also experience of stays in foreign countries, meetings and conferences.

The authors are fully responsible for the originality of their papers, for its subject and formal correctness. The authors shall make a written declaration that their papers have not been published in any other information source.

The board of editors of this journal will decide on paper publication, with respect to expert opinions, scientific importance, contribution and quality of the paper. The editors accept papers approved to print by the head of the workplace or papers with all the authors' statement they approve it to print.

The extent of original papers shall not exceed ten typescript pages, including tables, figures and graphs.

Manuscript should be typed on standard paper (quarto, 30 lines per page, 60 strokes per line, double-spaced typescript). A PC diskette with the paper text or graphical documentation should be provided with the paper manuscript, indicating the used editor program. Tables, figures and photos shall be enclosed separately. The text must contain references to all these annexes.

The **title** of the paper shall not exceed 85 strokes and it should provide a clear-cut idea of the paper subject. Subtitles of the papers are not allowed either.

Abstract. It must present information selection of the contents and conclusions of the paper, it is not a mere description of the paper. It must present all substantial information contained in the paper. It shall not exceed 170 words. It shall be written in full sentences, not in form of keywords and comprise base numerical data including statistical data.

Introduction has to present the main reasons why the study was conducted, and the circumstances of the studied problems should be described in a very brief form. This introductory section also provides information why the study has been undertaken.

Review of literature should be a short section, containing only literary citations with close relation to the treated problem.

Only original method shall be described, in other cases it is sufficient enough to cite the author of the used method and to mention modifications of this method. This section shall also contain a description of experimental material and the method of result evaluation.

In the section **Results**, which is the core of the paper, figures and graphs should be used rather than tables for presentation of quantitative values. A statistical analysis of recorded values should be summarized in tables. This section should not contain either theoretical conclusions or deductions, but only factual data should be presented here.

Discussion contains an evaluation of the study, potential shortcomings are discussed, and the results of the study are confronted with previously published results (only those authors whose studies are in closer relation with the published paper should be cited). The sections Results and Discussion may be presented as one section only.

References in the manuscript are given in form of citations of the author's name and year of publication. A list of references should contain publications cited in the manuscript only. References are listed alphabetically by the first author's name.

Key words should make it possible to retrieve the paper on the basis of the animal species investigated, characteristics of their health, husbandry conditions, applied substances, etc. The terms used in the paper title should not be used as keywords.

If any abbreviation is used in the paper, it is necessary to mention its full form at least once to avoid misunderstanding. The abbreviations should not be used in the title of the paper nor in the summary.

The author shall give his full name (and the names of other collaborators), academic, scientific and pedagogic titles, full address of his workplace and postal code, telephone and fax number, or e-mail.

VETERINARY MEDICINE - CZECH

Volume 45, No. 7, July 2000

CONTENTS

| | |
|---|-----|
| Konrádová V., Vajner L., Uhlík J., Zocová J.: The effect of Placebo-inhaler on the tracheal epithelium in rabbits | 189 |
| Machala M., Hilscherová K., Kubínová R., Ulrich R., Vykusová B., Kolářová J., Máchová J., Svobodová Z.: Biochemical markers of dioxin-like toxicity and oxidative stress in hepatic microsomes of bream (<i>Abramis brama</i>) and perch (<i>Perca fluviatilis</i>) in the Elbe river | 195 |
| Schlegelová J., Ryšánek D.: Effects of heat-processing of milk on validation characteristics of the Charm AIM-96 and Delvotest SP screening tests | 201 |
| REVIEW ARTICLE | |
| Grafenau P., Carstanjen B., Lepage O. M.: Osteocalcin: a biochemical marker of bone formation in equine medicine | 209 |

VETERINÁRNÍ MEDICÍNA

Ročník 45, č. 7, Červenec 2000

OBSAH

| | |
|--|-----|
| Konrádová V., Vajner L., Uhlík J., Zocová J.: Účinek preparátu Placebo-inhaler na epitel trachey králíků .. | 189 |
| Machala M., Hilscherová K., Kubínová R., Ulrich R., Vykusová B., Kolářová J., Máchová J., Svobodová Z.: Biochemické markery toxicity dioxinového typu a porovnání parametrů oxidativního stresu v cejnu velkém (<i>Abramis brama</i>) a okounu říčním (<i>Perca fluviatilis</i>) z řeky Labe | 195 |
| Schlegelová J., Ryšánek D.: Vliv tepelného ošetření mléka na charakteristiky validace screeningových testů Charm AIM-96 a Delvotest SP | 201 |
| PŘEHLED | |
| Grafenau P., Carstanjen B., Lepage O. M.: Osteokalcin: biochemický marker tvorby kostí v medicíně koní | 209 |

Vědecký časopis VETERINÁRNÍ MEDICÍNA ● Vydává Česká akademie zemědělských věd – Ústav zemědělských a potravinářských informací ● Redakce: Slezská 7, 120 56 Praha 2, tel.: + 420 2 24 25 79 39, fax: + 420 2 22 51 40 03, e-mail: edit@uzpi.cz ● Sazba a tisk: ÚZPI Praha ● © Ústav zemědělských a potravinářských informací, Praha 2000

Rozšiřuje Ústav zemědělských a potravinářských informací, referát odbytu, Slezská 7, 120 56 Praha 2
Podávání novinových zásilek povoleno Českou poštou, s. p., Odštěpný závod Střední Čechy, č. j. NOV-6588/00-P/1 dne 9. 5. 2000