# Immunohistochemical determination of calbindin-D28k in the kidney of postnatal rats

S. Timurkaan, B. G. Tarakci

Department of Histology-Embryology, Faculty of Veterinary Science, University of Firat, Elazig, Turkey

**ABSTRACT**: The localization of calbindin-D28k was examined in rat kidney at different stages of development. Calbindin was found specifically in the distal convoluted tubules (DCT). In the 10 day old rat kidney many cells apparently belonging to the collecting tubules were positive. In the neonatal nephrons there was also localization of calbindin-D28k to the limb of Henle, suggesting a difference in the regulation of intracellular calcium during maturation. Proximal tubule, renal corpuscle and macula densa gave no specific localization of calbindin-D28k.

Keywords: immunohistochemistry; calbindin-D28k; kidney; rat

Calbindin-D28k, a calcium binding protein with a molecular weight of approximately 28 000 (Christakos et al., 1989) was originally isolated from chick intestinal mucosa (Wasserman and Taylor, 1966). Calbindin-D28k belongs to a family of calcium binding proteins that includes calmodulin, parvalbumin, troponin C and S 100 (Christakos et al., 1989). This group of proteins contain the common calcium ion binding motif known as EF-hand domain (Kretsinger et al., 1982). Calbindin-D28k has been reported in various species (Wasserman and Taylor, 1971; Fulmer and Wasserman, 1975; Rhoten et al., 1984, 1986) and in many tissues, including kidney (Hermsdorf and Bronner, 1975; Rhoten and Christakos, 1981; Rhoten et al., 1985), bone (Christakos and Norman, 1978), pancreas (Pochet et al., 1987), and the brain (Baimbridge et al., 1982; Feldman and Christakos, 1983).

In the kidney, calbindin-D28k is localized in the distal convoluted tubule, in the connecting tubule cells and in the cortical collecting duct cells. This distribution of calbindin-D28k has been described in the chicken, rat, rabbit and human kidney with only very little variation (Roth et al., 1982; Taylor et al., 1982; Christakos et al., 1997).

The functional role of calbindin-D28k has not yet been established. It has been described as a carrier protein, which facilitates the transcellular Ca<sup>+2</sup> transport (Bronner and Stein, 1988; Christakos et al., 1997), as a buffer protein which prevents intracellular Ca<sup>+2</sup> concentrations from reaching toxic levels during Ca<sup>+2</sup> transport (Roth et al., 1981; Bronner and Stein, 1988; Johnson and Kumar, 1994), as a modulator of insulin secretion (Sooy et al., 1999; Parkash et al., 2002) and as an inhibitor of apoptosis (Christakos et al., 2003).

Although in adult rat kidney, calbindin-D28k, has been studied in some detail, very little is known early in development. The aim of present study was to examine the distribution of calbindin-D28k in the kidney of rat different developmental stages, using immunohistochemical technique. Thereby the aim was to improve the knowledge of the localization and to foster better understanding of the functional role of calbindin-D28k in the kidney.

#### MATERIAL AND METHODS

Forty Wistar rats of both sexes were examined at different developmental stages: newborn and 10,

20, 30, 40, 50, 60 day old and adult rats. The animals were anesthetized and killed using ether. The kidney was removed immediately and placed in 10% formalin in phosphate-buffered saline (PBS), pH 7.4, for 18 h before paraffin embedding. Tissues were routinely processed through a graded series of alcohols, cleared in xylol and embedded in paraffin. 5  $\mu$ m thick sections were obtained and processed for immunohistochemical staining.

**Immunohistochemistry**. Immunohistochemical staining was carried out by the peroxidase linked avidin-biotin complex (ABC) method. Blocking of endogenous peroxidase activity was done with 0.08% hydrogen peroxide ( $H_2O_2$ ) in methanol for 5 minutes (Sternberger, 1986). In order to block unspecific binding an incubation with (1:10) normal goat serum in 0.1 M PBS, pH 7.2 was performed.

ABC Technique. Sections were incubated for 16 to 20 h at 4°C in mouse anti-calbindin IgG (Sigma). The antibody was diluted to 1 : 500 in PBS containing 0.25% sodium azide and 2.5% bovine serum albumin. Sections were then incubated in biotinylated sheep anti-mouse IgG (Sigma) and followed with streptavidin horseradish peroxidase (Dako), both at a dilution of 1 : 50 in PBS, for 1 h at room temperature. Sections were washed in PBS for 30 minutes after each incubation. Sections were then immersed in glucose oxidase DAB-nickel ammonium sulphate (GDN) substrate (Shu et al., 1988) for 10 minutes, washed in distilled water and counterstained with eosine. Sections were examined with a light microscope and photomicrograps were taken.

The specificity of each immunohistochemical reaction was determined as recommended by

Sternberger (1979), including the replacement of specific antiserum pre-incubated with its corresponding antigen.

### **RESULTS**

Immunoreactivite cells was localized specifically to the distal convoluted tubules (DCT) examined of all rat kidneys (Figure 1). The calbindin positive cells of the DCT did not appear to have any particular regional localization within the cortical labyrinth. In the adult and young nephrons, no immunoreactivity was seen in the proximal convoluted tubules, Henle's loops or glomeruli.

In the young nephrons, many cells apparently belonging to the wall of collecting tubules showed positive reaction. Positively stained cells of the collecting tubules were seen to extend into the medulary rays and deep into the papillae (Figure 2).

In the neonatal nephrons, there was also specific and significant localization of calbindin in the Henle's loops (Figure 3).

## **DISCUSSION**

In this study using the ABC immunocytochemical technique, we have been able to localize calbindin-D28k in the adult, young and neonatal rat kidney specifically to the cells of the distal tubules in which the selective reabsorption of calcium is known to occur. This localization of calbindin-D28k is similiar to results obtained in kidneys of several diverse

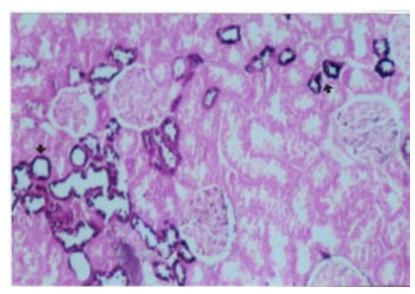


Figure 1. Calbindin-D28k immunoreactivity in adult rat kidney. Cells positive for Calbindin D-28k are seen in the distal convoluted tubules (arrows). Original magnification ×200

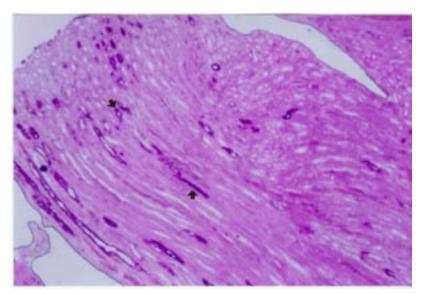


Figure 2. Calbindin-D28k immunoreactivity in the 10 day old rat kidney. Immunoreactivity was seen in the collecting tubules along the medullary rays and renal papilla (arrows). Original magnification ×50

species of vertebrates, mammalian (Rhoten and Christakos, 1981; Rhoten et al., 1985; Parmentier et al., 1987), and avian and reptilian (Rhoten et al., 1985; Parmentier et al., 1987).

In the young nephrons, not only are the collecting tubules of the cortex stained, but the collecting tubules extending along the medullary rays and deep into the papillae show a positive reaction. These results represent a more extensive distribution of calbindin-D28k than tat found in the kidneys of any other animals investigated previously (Rhoten and Christakos, 1981; Taylor et al., 1982; Schreiner et al., 1983; Rhoten et al., 1984) including the human (Roth et al., 1982). This statement is not strictly true as Rhoten et al. (1982) reported immunoreactive calbindin-D28k in the medulla, including the ascending

thick limb of the loop of Henle, as well as cortical labyrinth in neonatal rat kidney. These authors suggested a unique rol for calbindin-D28k during development which agrees with the suggestion of a different handling of calcium ion in the present study. This may also be suggestive of a different handling of Ca<sup>+2</sup> in these animals (Schreiner et al., 1983). However there is no physiological evidence to support these observations. Since the bulk of Ca<sup>+2</sup> reabsorption occurs in the proximal tubules and the ascending Henle's loops (Roth et al., 1982), this suggests that the renal calbindin-D28k does not play a role in the initial transmembrane transport of Ca<sup>+2</sup>, but that it may be involved in active Ca<sup>+2</sup> transport in the distal nephron and in those processes regulating intracellular Ca<sup>+2</sup> levels. The localization of

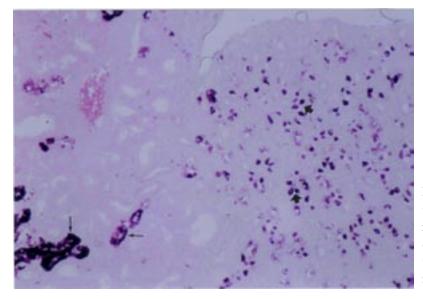


Figure 3. Calbindin-D28k immunoreactivity in the neonatal rat kidney. Immunoreactive cells were seen in the distal, convoluted tubules (thin arrows) and in the Henle's loop (thick arrows). Original magnification ×50

calbindin-D28k to a select population of cells supports the idea of an essential but highly specialized function for this protein in the kidney (Rhoten et al., 1986).

In the neonatal nephrons, there was also localization of calbindin-D28k to the limb of Henle, suggesting a difference in the regulation of intracellular calcium during maturation.

### REFERENCES

- Baimbridge K.G., Miller J.J., Parkes C.O. (1982): Calcium binding protein distribution in rat brain. Brain Res., 239, 519–525.
- Bronner F., Stein W.D. (1988): CaBP facilitates intracellular diffusion for Ca pumping in distal convoluted tubule. Am. J. Physiol., 255, F558–F562.
- Christakos S., Norman A.W. (1978): A vitamin D-dependent calcium binding protein in bone. Science, 202, 70–71.
- Christakos S., Gabrielides C., Rhoten W.B. (1989): Vitamin D-dependent calcium binding proteins: Chemistry, distribution, functional considerations, and molecular biology. Endocrin. Rev. 10, 3–26.
- Christakos S., Beck J.D., Hyllner S.J. (1997): Calbindin-D28k. In: Feldmann D. (ed.): Vitamin D. Academic Press. 209–221.
- Christakos S., Barletta F., Huening M., Dhawan P., Liu Y., Porta A., Peng X. (2003): Vitamin D Target Proteins: Function and Regulation. J. Cell. Biochem., 88, 238–244.
- Feldman S.C., Christakos S. (1983): Vitamin D-dependent calcium binding protein in rat brain: Biochemical and immunocytochemical characterization. Endocrinology, 112, 290–302.
- Fulmer C.S., Wasserman R.H. (1975): Isolation and partial characterization of intestinal calcium binding protein from cow, pig, horse, guinea pig and chick. Biochim. Biophys. Acta, 393, 134–142.
- Hermsdorf C.I., Bronner F. (1975): Vitamin D-dependent çalcium binding protein in rat kidney. Biochim. Biophys. Acta, 317, 553–561.
- Johnson J.A., Kumar R. (1994): Renal and intestinal calcium transport: Roles of vitamin D and vitamin D-dependent calcium binding proteins. Seminars in Nephrology, 14, 119–128.
- Kretsinger R.H., Mann J.E., Simmons J.G. (1982): Model of facilitated diffusion of calcium by the intestinal calcium-binding protein. In: Vitamin D, Chemical, Biochemical and Clinical Endocrinology of Calcium Metabolism. Walter de Gruyter & Co, New York. 233–248.

- Parkash J., Chaudhry M.A., Amer A.S., Christakos S., Rhoten W.B. (2002): Intracellular calcium ion response to glucose in  $\beta$ -cells of calbindin-D<sub>28k</sub> nullmutant mice and in  $\beta$ HC13 cells over-expressing calbindin-D<sub>28k</sub>. Endocrine, 18, 221–220.
- Parmentier M., Lawson D.E.M., Vassart G. (1987): Human 27kDa calbindin complementary DNA sequences. Eur. J. Biochem., 170, 207–212.
- Pochet R., Pipeleers D.G., Malaisse W.J. (1987): Calbindin D-27kDa: Preferential localization in non- B islet cells of the rat pancreas. Biol. Cell., 61, 155–161.
- Rhoten W.B., Christakos S. (1981): Immunocytochemical localization of vitamin D-dependent calcium binding protein in mammalian nephron. Endocrinology, 109, 981–983.
- Rhoten W.B., Van Eldik L.J., Marshak D., Christakos S. (1982): Calmodulin and D-dependent calcium-binding protein: functional, immunological, and immunocytochemical comparison. In: Norman A.W., Schaefer K., Herrath D.V., Grigoleit H.G. (eds.): Vitamin D: Chemical, Biochemical and Clinical Endocrinology of Calcium Metabolism. Walter de Gruyter, New York, 267–271.
- Rhoten W.B., Lubit B., Christakos S. (1984): Avian and mammalian calcium- binding protein in reptilian nephron. Gen. Comp. Endocrinol., 55, 96–103.
- Rhoten W.B., Bruns M.E., Christakos S. (1985): Presence and localization of two vitamin D dependent calcium binding proteins in kidneys of higher vertebrates. Endocrinology, 117, 674–683.
- Rhoten W.B., Gona O., Christakos S. (1986): Calciumbinding protein (28 000M Calbindin-D28k) in kidneys of the bullfrog Rana catesbeiana during metamorphosis. Anat. Rec., 216, 127–132.
- Roth J., Thorens B., Hunziker W., Norman A.W. (1981): Vitamin D-dependent calcium binding protein: Immunocytochemical localization in chick kidney. Science, 214, 197–200.
- Roth J., Brown D., Norman A.W., Orci L. (1982): Localization of the vitamin D-Dependent calcium binding protein in mammalian kidney. Am. J. Physiol., 243, F243–F252.
- Schreiner D.S., Jande S.S., Parkes C.O., Lawson D.E.M., Thomasset M. (1983): Immunocytochemical demonstration of two vitamin D-dependent calcium binding proteins in mammalian kidney. Acta. Anat., 117, 1–14.
- Shu S., Ju G., Fan L. (1988): The Glucose Oxidase-DAB-Nickel Method in Peroxidase Histochemistry of the Nevous System. Neurosci. Lett., 85, 169–171.
- Sooy K., Schermerhorn T., Noda M., Surana M., Rhoten W.B., Meyer M., Fleischer N., Sharp G.W.G., Christakos S. (1999): Calbindin-D<sub>28k</sub> controls [Ca<sup>2+</sup>]<sub>i</sub> and insulin

- release: evidence obtained from calbindin-D $_{28k}$  knockout mice and  $\beta$  cell lines. J. Biol. Chem., 274, 34343–34349.
- Sternberger L.A. (1979): The Unlabelled Antibody Peroxidase-Antiperoxidase (PAP) Method. In: Stenberg L.A. (ed.): Immunocytochemistry. John Wiley and Sons. New York. 104–169.
- Sternberger L.A. (1986): Immunocytochemistry. 3rd ed. Wiley, New York.
- Taylor A.N., McIntosh J.E., Bordeau J.E. (1982): Immunocytochemical localization of vitamin D-dependent cal-

- cium-binding protein in renal tubules of rabbit, rat and chick. Kidney Int., 21, 765–773.
- Wasserman R.H., Taylor A.N. (1966): Vitamin D<sub>3</sub> induced calcium-binding protein in chick intestinal mucosa. Science, 152, 791–793.
- Wasserman R.H., Taylor A.N. (1971): Evidence of a vitamin D3 induced calcium binding protein in new world primates. Proc. Soc. Exp. Biol. Med., 136, 25–28.

Received: 04-03-12

Accepted after corrections: 04-07-21

# Corresponding Author

Sema Timurkaan, Department of Histology-Embryology, Faculty of Veterinary Science, University of Firat, 23119 Elazig, Turkey

Tel. +90 424 2370000/4051, fax+90 4242388173, e-mail: stimurkaan@hotmail.com