Serologic survey of wild boars for mosquito-borne viruses in South Moravia (Czech Republic)

J. Halouzka¹, Z. Juricova¹, J. Jankova^{1,2}, Z. Hubalek^{1,2}

ABSTRACT: A serosurvey for mosquito-borne viruses was carried out in 93 wild boars (Sus scrofa), using a plaque-reduction neutralization microtest with Vero cells. The boars were sampled on 24 hunting grounds of the Breclav district (South Moravia) from 2000 to 2002. Specific antibodies to Flavivirus West Nile (WNV) were detected in six (6.5%) animals, and only in Lanzhot and Kostice, i.e., in the area of the "Soutok" game reserve where WNV was previously isolated from mosquitoes in South Moravia. However, the antibody titres were comparatively low (1:20–1:40). A substantially higher seroprevalence was revealed against Orthobunyavirus Tahyna (TAHV): 18 (19.4%) wild boars were positive, and the titres ranged from 1:20 up to 1:640. Only one animal (1.1%) seroreacted with Orthobunyavirus Batai (Calovo), at a low titre of 1:20. The sera were additionally examined by a haemagglutination-inhibition test against Alphavirus Sindbis: two boars (2.2%) revealed antibodies, the titres were 1:20 and 1:80. The serosurvey indicates that the activity of mosquito-borne viruses in South Moravia has decreased compared with the past decades, but that surveillance for these viruses is still necessary.

Keywords: antibodies; West Nile virus; Tahyna virus; Batai virus; Sindbis virus; Czechland; swine

Mosquito-borne viruses circulate in natural foci between endotherm (homeotherm) vertebrates and mosquitoes. Six vertebrate pathogenic mosquitoborne viruses have been reported in Central Europe (Malkova et al., 1986; Hubalek and Halouzka, 1996; Weissenbock et al., 2002): Sindbis (SINV), West Nile (WNV), Usutu (USUV), Tahyna (TAHV), Batai (BATV; synonym = Calovo), and Lednice (LEDV), and at least two are regarded as being of veterinary importance (WNV, USUV).

The aim of this study was to evaluate the present activity of WNV and other mosquito-borne viruses in South Moravia (Czech Republic), using an indirect method of serological survey of wild mammals (the wild boar, *Sus scrofa*) potentially exposed to these viruses in the field. West Nile virus was already isolated in this region in 1997 and 1999 (Hubalek et al., 1998, 2000) as well as in neighbouring Slovakia (Labuda et al., 1974), and five human cases of WNV

fever were described after floods in South Moravia in 1997 (Hubalek et al., 2000). An extensive natural focus of TAHV infections ("Valtice fever") has been well documented in South Moravia since the 1960s, and the virus has been isolated repeatedly from vector mosquitoes (e.g., Danielova et al., 1976; Rosicky and Malkova, 1980). One isolation of BATV was reported from *Anopheles maculipennis* complex mosquitoes in Rakvice (Smetana et al., 1967), while SINV has not yet been isolated in South Moravia, in contrast to neighbouring western Slovakia (Ernek et al., 1973).

MATERIAL AND METHODS

Study sites

All 24 visited hunting grounds belonged to localities in the district of Breclav (number of animals ex-

Supported by the Grant Agency of the Academy of Sciences of the Czech Republic (Grant No. IAA 600930611) and by EU Grant GOCE-2003-010284 EDEN.

¹Institute of Vertebrate Biology of the Academy of Sciences of the Czech Republic, Brno, Czech Republic

²Institute of Experimental Biology, Faculty of Science, Masaryk University, Brno, Czech Republic

amined): Boleradice (3), Charvatska Nova Ves (6), Divaky (9), Hlohovec (4), Hrusky (1), Hustopece (1), Kostice (4), Lanzhot (17), Lednice (4), Moravska Nova Ves (3), Morkuvky (3), Nikolcice (1), Novy Dvur (2), Podivin (2), Postorna (5), Rakvice (1), Tisnov (BV, 8), Tvrdonice (2), Tynec (1), Valtice (8), Velke Bilovice (2), Velke Hosteradky (1), Vranovice (1), and Zajeci (3). Most sampling sites were situated in habitats with abundant mosquito populations.

Blood sampling

The wild boars, which were killed by shooting, and throughout all seasons of the year, were weighed and their age determined. Blood samples were collected from the heart or from the thoracic cavity. After clotting, the blood samples were centrifuged in the laboratory and the sera were then stored at -20° C until use.

Plaque-reduction neutralization microtest (PRN μ T)

The seroneutralization method was originally proposed by Madrid and Porterfield (1969, 1974), and later adopted to a microtechnique on 96-well (flatbottomed) sterile microplates (Sarstedt) for cell culture (Hubalek et al., 1979). Vero E6 cells were serially propagated in Leibovitz L-15 medium (Sigma) supplemented with 10% of foetal calf serum (FCS, Gibco Bio-Cult) and antibiotics. Tested sera were inactivated at 56°C for 30 m, and diluted 1:10 in L-15 medium for screening; 30 µl of the diluted serum was mixed in a microplate well with 30-µl test dose of the virus (containing 20–30 PFU of individual viruses) in L-15 supplemented with 3% of inactivated FCS, and incubated at 37°C for 60 min; 60 µl of Vero cell suspension (in L-15 with 3% FCS) was then added to each test well (20 000 to 30 000 cells), and 120 µl of carboxymethylcellulose sodium salt overlay (1.5% CMC of medium viscosity – BDH in PBS mixed with the same volume of L-15 with 3% of inactivated FCS) was added after an incubation at 37°C for 4 h in each well. Sera were tested in duplicate, and controls included the virus test dose and its titration, immune WNV reference serum; control negative serum; and cells without virus. The microplates were incubated at 37°C for 5 days, and the microplate cultures were then stained with 0.1% acidic solution of naphthalene blue black (Fluka).

The virus strains used in PRNµT were: (1) West Nile virus (WNV) Egyptian topotype strain Eg-101 (Melnick et al., 1951), passaged 17 times in suckling mouse brain, homogenized in PBS with 0.4% of bovine serum albumin fraction V, Sigma (PBS-BSA), and centrifuged; (2) Tahyna virus (TAHV) strain T16 (Bardos et al., 1975), passaged six times in suckling mouse brain, homogenized in PBS-PBS, and centrifuged; (3) Batai virus (BATV) strain 184 (Bardos and Cupkova, 1962), passaged 10 times in suckling mouse brain, homogenized in PBS-BSA, and centrifuged. Unfortunately, we were unable to use Sindbis *Alphavirus* strains Eg339 or P3328 because they did not produce good plaques in Vero E6 cells.

Sera reactive with a virus, revealing 90% or greater reduction in the number of plaques at the 1:10 dilution during the screening (corresponding to the 1:20 final dilution of the serum – after mixing with the virus test dose), were titrated by twofold dilutions, and those dilutions corresponding to a 90% reduction of plaque numbers were regarded as the serum titres (PRN μT_{90}). Reciprocal titres ≥ 20 were considered positive. Boar sera reacting with WNV were also tested in PRNµT on SPEV cells with Flavivirus of tick-borne encephalitis (TBEV) strain Hypr (Pospisil et al., 1954), passaged 55 times in HeLa cells followed by 11 passages in mouse brain, in order to exclude cross reactions with this antigenically partially related virus occurring in Moravia.

Haemagglutination-inhibition test (HIT)

We encountered problems with the plaquing of Sindbis virus in microcultures of Vero E6 cells. Therefore, we used the haemagglutination-inhibition test (HIT) in microplates for this virus (Eg-339; Taylor et al., 1955). All serum samples for this assay were acetone-extracted and tested with saccharose-and acetone-processed antigen by using eight haemagglutinin units (Clarke and Casals, 1958); titres > 20 were considered positive.

Statistical methods

The statistical significance of differences between proportions was evaluated by using cross-tables 2×2 or 2×3 and the χ^2 test (SOLO 4.0 package, BMDP Statistical Software, Los Angeles).

RESULTS

The serosurvey was carried out in 93 wild boars sampled on 24 hunting grounds. Specific neutralizing antibodies to WNV were detected in six (6.5%) animals and only from the Lanzhot and Kostice hunting grounds (Table 1). However, the antibody titres were comparatively low (1:20–1:40). Some additional animals seroreacted with WNV (A35,

A36, A48, A60 – see Table 1), but at the same time they also reacted with TBEV at similar or higher titres. Those results were, therefore, regarded as flavivirus cross reactions and the corresponding animals were not taken in consideration as specific reactors.

A substantially higher seroprevalence was revealed against TAHV: 18 (19.4%) wild boars were seropositive, the titres ranged from 1:20 up to 1:640

Table 1. Survey of wild boars seroreacting with Tahyna or West Nile virus in PRN μ T

No.	Age (years)	Weight (kg)	Locality	Date	TAH titre	WNV titre	TBE titre
A7	< 1	20	Lanzhot	July 2002	< 20	20	< 20
A13	< 1	25	Lanzhot	July 2002	< 20	40	20
A16	< 1	12	Kostice	July 2002	< 20	40	< 20
A28	< 1	8	Lanzhot	July 2002	160	< 20	n.t.
A32	<1	10	Lednice	July 2002	640	< 20	n.t.
A41	<1	30	Lednice	July 2002	20	< 20	n.t.
B19	< 1	30	Zajeci	Nov.2000	160	≤ 20	< 20
A6	1	47	Kostice	July 2002	< 20	20	< 20
A20	1	31	Vel. Bilovice	July 2002	160	< 20	n.t.
A24	1	55	Lanzhot	July 2002	< 20	20	< 20
A29	1	44	Lanžhot	July 2002	< 20	20	< 20
A35	1	38	Charv. N. Ves	July 2002	< 20	20	20
A36	1	40	Charv. N. Ves	July 2002	< 20	20	160
A43	1	50	Lanzhot	July 2002	20	< 20	n.t.
A45	1	60	Lanzhot	July 2002	320	< 20	n.t.
A50	1	64	Postorna	July 2002	160	≤ 20	< 20
A54	1	55	Podivin	July 2002	320	< 20	n.t.
A56	1	45	Postorna	July 2002	640	≤ 20	20
A58	1	40	Charv. N. Ves	July 2002	20	< 20	n.t.
B2	1	38	Novy Dvur	July 2002	160	< 20	n.t.
B23	1–2	70	Divaky	Nov. 2000	320	< 20	n.t.
B13	> 2	70	Tisnov (BV)	Nov. 2000	80	< 20	n.t.
A42	3	80	Lanzhot	July 2002	80	< 20	n.t.
A48	3	70	Lanzhot	July 2002	< 20	20	40
A51	3	80	Tvrdonice	July 2002	320	< 20	n.t.
A60	4	80	Lanzhot	July 2002	< 20	40	80
B27	4	80	Boleradice	Jan. 2001	640	< 20	n.t.
A59	7	150	Lanzhot	July 2002	640	< 20	n.t.

n.t. = not tested; titres in bold are regarded as specific for particular virus

(Table 1). The three age groups of the examined boars were: (1) < 1 year (37 animals); (2) 1 to 2 years (46 animals); and (3) > 2 years (10 animals). There were four boars (10.8%) seropositive for TAHV in the first age group, nine (19.6%) seropositives in the second group, while five (50.0%) animals in the oldest group had specific antibodies to TAHV. The differences in the seropositivity rate between the three age groups of the wild boar are statistically significant: $\chi^2 = 7.748$ (P = 0.021; d.f. = 2). The significant pair-wise differences are those between the age groups 1 and 3 ($\chi^2 = 7.809$; P = 0.005; *d.f.* = 1), and between the age groups 2 and 3 (χ^2 = 4.058; P = 0.044), the difference between the age groups 1 and 2 ($\chi^2 = 1.190$; P = 0.275) is not significant. The oldest wild boars, therefore, showed a significantly higher proportion of seropositive animals. This statistical comparison could not be applied to the other viruses due to low numbers of seropositive individuals.

Only one animal (a 7-year old boar A59, shot on Lanzhot hunting grounds in July 2002) seroreacted with BATV (1.1%), at a low titre of 1:20.

The sera, when additionally examined by HIT against SINV, revealed two boars (2.2%) with antibodies: a young animal B4, shot on Boleradice hunting grounds in October 2000, and a > 2-year old boar B13 shot on Tisnov (BV) hunting grounds in November 2000; the titres of antibodies were 1:20 and 1:80, respectively.

DISCUSSION

In this study, we used PRNµT for all viruses except for SINV. The neutralization test is regarded as the 'gold standard' in arbovirus serology and is used for the verification of other serological tests (ELISA, heamagglutination-inhibition test - HIT) because it is generally more specific and discriminatory. However, it is well known that, e.g., flaviviruses are responsible for a high degree of serological cross-reactivity, sometimes even in the neutralization test (Theiler and Downs, 1973; Madrid and Porterfield, 1974; Calisher et al., 1989; Weingartl et al., 2003; Weissenboock et al., 2003; Niedrig et al., 2007). Several antigenically related flaviviruses might co-occur in one area, e.g., TBEV and WNV in Central Europe. It is sometimes very difficult to decide which particular antigen is responsible for the antibody production – controversial results may thus be published. It is always necessary to interpret results of flavivirus (WNV) serology with great care, especially those obtained from serosurveys in wild vertebrates (e.g., shot-killed game animals) where non-specific inhibitors of viruses may occasionally occur (e.g., Holden et al., 1965; Theiler and Downs, 1973). In the PRN μ T, we estimated the results conservatively, as a 90% reduction in the number of plaques (not a 50% reduction which is sometimes used), and 1:20 dilution (instead of the usual 1:10) as a titre cut-off point.

In general, the data indicate a limited WNV activity in South Moravia during 2000–2002, restricted to the area of the "Soutok" game reserve at Lanzhot. This is in concordance with previous attempts at isolation, when WNV was detected in *Culex pipiens* mosquitoes in this area previously (Hubalek et al., 1998, 2000). It is possible that antibodies detected in wild boar were formed to the local (enzootic) genomic lineage 3 ("Rabensburg") of WNV, antigenically indistinguishable from the lineages 1 and 2 of WNV (Bakonyi et al., 2005). In nearby western Slovakia (Zahorska lowland), where WNV was also isolated from mosquitoes (Labuda et al., 1974), Kozuch et al. (1976) detected neutralizing antibodies to WNV in 2.0% of examined wild boars.

TAHV antibodies, on the other hand, were detected in wild boar at a much higher frequency (19.4%) and over a wider range, involving the hunting grounds of Lanzhot, Tvrdonice, Postorna, Lednice, Valtice, Podivin, Zajeci, and Boleradice, all characterized by the presence of floodplain forest or wetland/fishpond ecosystems and by abundant mosquito populations. We found that the older the boar, the higher the probability of the presence of TAHV antibodies. Such a pattern is typical for animals living in an enzootic, long-term natural focus of infection. However, in the past, the seroprevalence was even higher. For instance Juricova (1992) and Juricova and Hubalek (1999) found 41.7% and 46.7% of wild boar with TAHV haemagglutination-inhibiting antibodies in the years 1990 and 1993-1997, respectively. In addition, as much as 73.3% (Danielova and Marhoul, 1968; neutralization test) and 54.8% (Kolman, 1973; HIT) of South-Moravian domestic pigs were found seropositive against TAHV in 1963–1964. Kozuch et al. (1976) detected antibodies neutralizing TAHV in 28.9% of examined wild boars in western Slovakia, and Aspock and Kunz (1971) found antibodies against TAHV in one of four tested wild boars in eastern Austria.

BATV antibodies were detected at a very low level in the present study. In the past, Juricova

and Hubalek (1999) found as much as 18.7% of wild boars in South Moravia with BATV haemagglutination-inhibiting antibodies. Kozuch et al. (1976) detected antibodies neutralizing BATV in 5.3% of wild boars in western Slovakia. In addition, Kolman (1973) detected BATV antibodies in 17.2% of South-Moravian domestic pigs in 1964.

SINV antibodies were also detected at a low frequency in this study. Previously, Juricova (1992) did not detect SINV antibodies in wild boar sampled in 1990, while 10.0% of boars were seropositive during 1993–1997 in South Moravia (Juricova and Hubalek, 1999). Kozuch et al. (1976) detected no antibodies neutralizing SINV in wild boars in western Slovakia.

We did not test the wild boars for the presence of antibodies against the two remaining mosquito-borne viruses known to occur sporadically in Central Europe, i.e. USUV and LEDV. These viruses are ornithophilic, and antibodies against them will be mainly found in free-living birds, not in mammals. Bird groups susceptible to LEDV are wetland anseriforms (Malkova et al., 1986) and for USUV largely thrushes (family Turdidae) and birds of prey (Weissenbock et al., 2002).

In conclusion, the serosurvey in wild boars indicates that the activity of mosquito-borne viruses has decreased compared with the past decades in South Moravia. However, long-term precipitations or floods could reverse the situation, and surveillance for these viruses still remains necessary.

Acknowledgements

The wild boar blood samples were collected by a number of hunters, and their contribution is gratefully acknowledged. This study was funded by the Grant Agency of the Academy of Sciences of the Czech Republic (IAA 600930611), and by EU grant GOCE-2003-010284 EDEN; it is catalogued by the EDEN Steering Committee as EDEN0104 (http://www.eden-fp6project.net).

REFERENCES

Aspock H., Kunz C. (1971). Antikorper gegen Tahyna-Virus und Calovo-Virus in wildlebenden und domestizierten Saugetieren im ostlichen Neusiedlersee-Gebiet (Ost-Osterreich). Zentralblatt fur Bakteriologie I Originale, 216, 435–440.

- Bakonyi T., Hubalek Z., Rudolf I., Nowotny N. (2005): Novel flavivirus or new lineage of West Nile virus, Central Europe. Emerging Infectious Diseases, 11, 225–231.
- Bardos V., Cupkova E. (1962): The Calovo virus the second virus isolated from mosquitoes in Czechoslovakia. Journal of Hygiene Epidemiology Microbiology and Immunlogy, 6, 186–192.
- Bardos V., Medek M., Kania V., Hubalek Z. (1975): Isolation of Tahyna virus from the blood of sick children. Acta Virologica, 19, 447.
- Calisher C.H., Karabatsos N., Dalrymple J.M., Shope R. E., Porterfield J.S., Westaway E.G., Brandt W.E. (1989): Antigenic relationships between flaviviruses as determined by cross-neutralization tests with polyclonal antisera. Journal of General Virology, 70, 37–43.
- Clarke D.H., Casals J. (1958): Techniques for hemagglutination and hemagglutination-inhibition test with arthropod-borne viruses. American Journal of Tropical Medicine and Hygiene, 7, 561–573.
- Danielova V., Marhoul Z. (1968): Incidence of antibodies against some arboviruses in humans, domestic and wild animals living in the natural focus of Tahyna virus in South Moravia (in Czech). Ceskoslovenska Epidemiologie Mikrobiologie a Imunologie, 17, 155–161.
- Danielova V., Malkova D., Minar J., Ryba J. (1976): Dynamics of the natural focus of Tahyna virus in southern Moravia and species succession of its vectors, the mosquitoes of the genus *Aedes*. Folia Parasitologica, 23, 243–249.
- Ernek E., Kozuch O., Gresikova M., Nosek J., Sekeyova M. (1973): Isolation of Sindbis virus from the reed warbler (*Acrocephalus scirpaceus*) in Slovakia. Acta Virologica, 17, 359–361.
- Holden P., LaMotte L.C., Shriner R.B. (1965): Arbovirushemagglutinin inhibitor in acetone-extracted serums from normal chickens. Science, 147, 169–170.
- Hubalek Z., Chanas A.C., Johnson B.K., Simpson D.I.H. (1979): Cross-neutralization study of seven California group (Bunyaviridae) strains in homoiothermous (PS) and poikilothermous (XTC-2) vertebrate cells. Journal of General Virology, 42, 357–362.
- Hubalek Z., Halouzka J. (1996): Arthropod-borne viruses of vertebrates in Europe. Acta Scientiarum Naturalium Brno, 10, No. 4–5, 1–95.
- Hubalek Z., Halouzka J., Juricova. Z., Sebesta O. (1998): First isolation of mosquito-borne West Nile virus in the Czech Republic. Acta Virologica, 42, 119–120.
- Hubalek Z., Savage H.M., Halouzka J., Juricova Z., Sanogo Y.O., Lusk S. (2000): West Nile virus investigations in South Moravia, Czechland. Viral Immunology, 13, 427–433.

- Juricova Z. (1992): Antibodies to arboviruses in game animals in South Moravia (in Czech). Veterinarni Medicina, 37, 633–636.
- Juricova Z., Hubalek Z. (1999): Serological surveys for arboviruses in the game animals of southern Moravia (Czech Republic). Folia Zoologica, 48: 185–189.
- Kolman J.M. (1973): Serologic examination of some domestic animals from South Moravia on the presence of antibodies to selected arboviruses of the A, B, California and Bunyamwera groups. Folia Parasitologica, 20, 353–360.
- Kozuch O., Nosek J., Gresikova M., Ernek E. (1976): Surveillance of mosquito-borne natural focus in Zahorska Lowland, 115-118. In: Sixl W. (ed.): 2nd International Arbeitskolloquium uber die Naturherde von Infektionskrankheiten in Zentraleuropa. Hygiene Institut der Universitat, Graz.
- Labuda M., Kozuch O., Gresikova M. (1974): Isolation of West Nile virus from *Aedes cantans* mosquitoes in West Slovakia. Acta Virologica, 18, 429–433.
- Madrid A.T., Porterfield J.S. (1969): A simple micro-culture method for the study of group B arboviruses. Bulletin WHO, 40, 113–121.
- Madrid A.T., Porterfield J.S. (1974): The flaviviruses (group B arboviruses): a cross-neutralization study. Journal of General Virology, 23, 91–96.
- Malkova D., Danielova V., Holubova J., Marhoul Z. (1986): Less known arboviruses in Central Europe (in Czech). Rozpravy CSAV, rada matem.-prirodnich ved, 96, No. 5, 1–75.
- Melnick J.L., Paul J.R., Riordan J.F., Barnett V.H.H., Goldblum N., Zabin E. (1951): Isolation from human sera in Egypt of a virus apparently identical to West Nile virus. Proceedings of the Society for Experimental Biology and Medicine, 77, 661–665.
- Niedrig M., Sonnenberg K., Steinhagen K., Paweska J.T. (2007): Comparison of ELISA and immunoassays for measurement of IgG and IgM antibody to West Nile

- virus in human sera against virus neutralisation. Journal of Virological Methods, 139, 103–105.
- Pospisil L., Jandasek L., Pesek J. (1954): Isolation of new strains of meningoencephalitis virus in the Brno area during summer of 1953 (in Czech). Lekarske Listy (Brno), 9, 3–5.
- Rosicky B., Malkova D., eds. (1980): Tahyna virus natural focus in southern Moravia (in Czech). Rozpravy CSAV, rada matem.-prirodnich ved, 90, 1–107.
- Smetana K., Danielova V., Kolman J.M., Malkova D., Minar J. (1967): The isolation of the Calovo virus from the mosquitoes of the group *Anopheles maculipennis* in southern Moravia. Journal of Hygiene Epidemiology Microbiology and Immunology, 11, 55–59.
- Taylor R.M., Hurlbut H.S., Work T.H., Kingston J.R., Frotingham T.E. (1955): Sindbis virus: a newly recognized arthropod-transmitted virus. American Journal of Tropical Medicine and Hygiene, 4, 844–862.
- Theiler M., Downs W.G. (1973): The Arthropod-Borne Viruses of Vertebrates. Yale University Press, New Haven and London. 578 pp.
- Weingartl H.M., Drebot M.A., Hubalek Z., Halouzka J., Andonova M., Dibernardo A., Cottam-Birt C., Larence J., Marszal P. (2003): Comparison of assays for the detection of West Nile virus antibodies in chicken serum. Canadian Journal of Veterinary Research, 67, 128–132.
- Weissenboock H., Kolodziejek J., Url A., Lussy H., Rebel-Bauder B., Nowotny N. (2002): Emergence of Usutu virus, an African mosquito-borne flavivirus of the Japanese encephalitis virus group, Central Europe. Emerging Infectious Diseases, 8, 652–656.
- Weissenboock H., Hubalek Z., Halouzka J., Pichlmair A., Maderner A., Fragner K., Kolodziejek J., Loupal G., Kolbl S., Nowotny N. (2003): Screening for West Nile virus infections in susceptible animal species in Austria. Epidemiology and Infection, 131, 1023– 1027.

Received: 2008-01-28

Corresponding Author:

Prof. RNDr. Zdenek Hubalek D.Sc., Medical Zoology Laboratory, Institute of Vertebrate Biology of the ASCR, Klasterni 2, CZ-69142 Valtice, Czech Republic Tel. +420 519 352 961; e-mail: zhubalek@ivb.cz