# Death of a loggerhead sea turtle (*Caretta caretta*) from ingestion of an eel (*Myrichthys ocellatus*)

Radan Elvis Matias de Oliveira<sup>1,2,3,4</sup>\*, Juliana Maia de Lorena Pires<sup>2,4</sup>, Jael Soares Batista<sup>1</sup>, Fernanda Loffler Niemeyer Attademo<sup>2,4</sup>, Daniel Solon Dias de Farias<sup>2,3,4</sup>, Augusto Carlos da Boaviagem Freire <sup>2,4</sup>, Aline da Costa Bomfim<sup>2,3,4</sup>, Laize Regina Palhares de Lima<sup>2,3,4</sup>, Rysonely Maclay de Oliveira<sup>2,4,5</sup>, Simone Almeida Gavilan<sup>2,3,4</sup>, Flavio Jose de Lima Silva<sup>2,3,4</sup>, Moacir Franco de Oliveira<sup>1</sup>

**Citation:** Oliveira REM, Pires JML, Batista JS, Attademo FLN, Farias DSD, Freire ACB, Bomfim AC, Lima LRP, Oliveira RM, Gavilan SA, Silva FJL, Oliveira MF (2020): Death of a loggerhead sea turtle (*Caretta caretta*) from ingestion of an eel (*Myrichthys ocellatus*). Vet Med-Czech 65, 415–420.

Abstract: Gastrointestinal perforation in sea turtles may be associated with the ingestion of solid waste; however, other factors, including the ingestion of marine organisms, may cause intestinal perforation. Herein, we report, presumably, the first case of a death of a loggerhead sea turtle (*Caretta caretta*) from ingesting a live goldspotted eel (*Myrichthys ocellatus*) and describe the necropsy findings. The adult female loggerhead sea turtle was registered alive by a team of the Cetáceos da Costa Branca Project of Rio Grande do Norte State University (PCCB-UERN). The animal died in captivity after 8 days of an attempted rehabilitation process, and the carcass was immediately sent for necropsy. After incision, an encapsulated structure in the cranial region of the left lung was identified as a specimen of the ingested eel. The histopathological examination of the lung showed fibrin and numerous leukocytes, mainly macrophages, in the alveolus and bronchioles. The necropsy revealed that the ingestion of a live goldspotted eel (*M. ocellatus*) caused a gastric perforation in the turtle; this resulted in the displacement of the eel to the celomatic cavity with its cranial portion in the left lung, leading to sepsis and the consequent death of the turtle.

Keywords: Cheloniidae; gastric perforation; septicaemia; Testudines

<sup>&</sup>lt;sup>1</sup>Postgraduate Program in Animal Science, Department of Animal Sciences, Federal Rural University of Semi-Árido – UFERSA, Mossoró-RN, Brazil

<sup>&</sup>lt;sup>2</sup>Cetáceos Costa Branca Project, State University of Rio Grande do Norte – PCCB-UERN, Mossoró-RN, Brazil

<sup>&</sup>lt;sup>3</sup>Vertebrate Morphophysiology Laboratory, Department of Morphology, Bioscience Center - CB, Federal University of Rio Grande do Norte – UFRN, Natal-RN, Brazil

<sup>&</sup>lt;sup>4</sup>Center for Environmental Studies and Monitoring – CEMAM, Areia Branca -RN, Brazil

<sup>&</sup>lt;sup>5</sup>Postgraduate Program in Natural Sciences, Faculty of Exact and Natural Sciences, State University of Rio Grande do Norte -UERN, Mossoró-RN, Brazil

 $<sup>*</sup>Corresponding\ author:\ radan\_elvis@hotmail.com$ 

Loggerhead sea turtles (*C. caretta*) are a globally distributed migratory species and are classified as "vulnerable" to extinction according to the International Union for Conservation of Nature red list of threatened species (Casale and Tucker 2017). Conservation of this species is a complicated task that requires knowledge of its essential biological aspects, such as its reproduction, migration, and feeding habits (Burke et al. 1993).

These animals are carnivorous throughout their life cycle, preferentially feeding on fish, followed by pelagic tunicates, crustaceans, molluscs, and other invertebrates (Tomas et al. 2001; Revelles et al. 2007). The dietary data of these specimens are relevant to the understanding of their predator-prey relationships.

Moreover, it is difficult to diagnose gastrointestinal diseases in sea turtles antemortem. Clinical signs found on necropsies provide relevant decisionmaking information regarding the rehabilitation of stranded animals.

Herein, we report, presumably, the first case of a death of a loggerhead sea turtle (*C. caretta*) caused by ingesting a live goldspotted eel (*M. ocellatus*) and describe the necropsy findings.

## Case description

An adult female loggerhead sea turtle (*C. caretta*), previously registered by the Cetáceos Costa Branca Project of Rio Grande do Norte State University (PCCB-UERN), was rescued alive on March 27, 2018 from Requenguela beach (4°39'56.4"S, 37°25'41.7"W), Icapuí, Ceará, Brazil.

The experiment was conducted with authorisation from the Ethics Committee on Animal Use (License No. 01/2019).

The prostrate and debilitated animal was rescued and sent to the Wild Animal Rehabilitation Center of the Cetáceos Costa Branca Project (UERN) in the city of Areia Branca, Rio Grande do Norte (RN), Brazil, where it was kept in a salt water enclosure measuring 200 cm in diameter and 75 cm in depth, with a water temperature of 25–28 °C and natural light from approximately 5:30 a.m. to 5:30 p.m. The biometric data included a curved carapace length of 96.0 cm, a curved carapace width of 87.0 cm, and a body weight of 91.0 kg.

The clinical examination revealed a regular body score, anorexia, noisy breathing, severe dehydra-

tion, and the presence of algae and sessile crustaceans (barnacles) on its carapace and plastron. The turtle received supportive treatment with fluid therapy (KabiPac®, 0.9% sodium chloride, 20 ml/kg i.v.; Fresenius Kabi, São Paulo, Brazil), diluted with a vitamin (Polivin B12®, vitamin B12, 0.3 ml/kg i.v.; Bravet, Rio de Janeiro, RJ, Brazil), antibiotic therapy (CEF 50®, ceftiofur hydrochloride, 4 mg/kg i.m.; Agener União, Embu-Guaçu, São Paulo, Brazil), and an analgesia (Tramal®, tramadol hydrochloride, 5 mg/kg s.c.; Grünenthal, Aachen, Germany; Maxicam®, meloxicam, 0.1 mg/kg i.m.; Ourofino, São Paulo, Brazil; D-500®, dipyrone, 25 mg/kg i.m.; Zoetis, Campinas, São Paulo, Brazil).

The turtle died after 8 days of treatment and was immediately sent for necropsy to the Marine Biota Monitoring Laboratory of UERN in Mossoró, RN.

A complete necropsy was performed following the standard *postmortem* protocol for sea turtles (Wyneken 2004). The celomatic cavity surrounding the entire plastron was opened for a macroscopic analysis and then removed for an *in situ* analysis of the organs. Subsequently, all the organs were removed outside of the cavity for evaluation, and the most evident macroscopic findings were photographed.

Organ fragments were collected for a histopathological examination and fixed in 10% formalin. After fixation for 48 h, the samples were dehydrated using increasing concentrations of alcohol (70%, 80%, 90%, 95%, 100%, 100%, and 100%) for a period of 1 h for each concentration, followed by diaphanization consisting of two xylol baths of 30 min each and a subsequent paraffin embedding by immersion in a granular histological paraffin with a melting point of 58–62 °C (Synth®; Labsynth, São Paulo, SP, Brazil) in an oven at 58 °C.

After preparation, the histological blocks were cut into 5-µm sections using a microtome (Leica RM2125 RT) adhered to glass slides, placed in an oven at 58 °C for deparaffinisation, and stained with haematoxylin and eosin (Prophet et al. 1992).

Photomicrographs of the most representative images were acquired under a light microscope (Leica ICC50 HD) and obtained using the Leica LAS EZ software (v2.0.0.; Leica Microsystems Ltd., Heerbrugg, Switzerland).

In the initial portion of the digestive tract, the oesophagus and stomach showed no food content, but a contained mucus; the cardiac region of the gastric mucosa showed a small, healed, rounded lesion

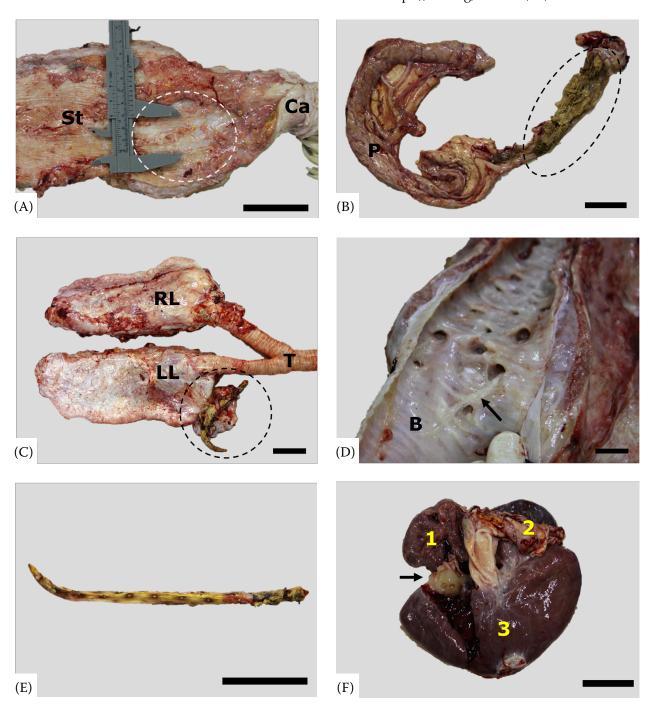


Figure 1. Macroscopic findings of a loggerhead seaturtle (*Caretta caretta*) that died from ingestion of a live goldspotted eel (*M. ocellatus*). (A) Perforation in the stomach (St), with presence of scar tissue (dashed circle) near the cardiac region (Ca) caused by the ingestion of the eel. Bar = 5.0 cm. (B) Presence of the encapsulated (dashed circle) extravasated stomach content in the pancreas (P). Bar = 5.0 cm. (C) Tracheal region (T) of the respiratory system and the right (RL) and left (LL) lungs with an *M. ocellatus* specimen in the cranial region of the left lung (LL). Bar = 5.0 cm. (D) Presence of a yellowish mucoid secretion in the bronchi (arrow). (E) Details of the *M. ocellatus* eel. Bar = 2.0 cm. (F) Dilated flaccid right atrium (1) with a thickened atrioventricular valve (arrow) and left atrium (2) and ventricle (3) without any apparent macroscopic changes. Bar = 5.0 cm

(Figure 1A). The intestines showed dilated portions containing gas and a greenish-brown liquid.

The total length of the small and large intestines was 714 cm. An encapsulated foreign body was

found adjacent to the pancreas, and an incision revealed that some unidentified food content extravasated from the stomach (Figure 1B).

The liver was enlarged, with rounded edges, capsular thickening, whitish granules distributed diffusely in the serosa, nutmeg-coloured parenchyma, and an abscess (10.0 cm diameter) with a caseous content (Figure 2A and 2B). The histopathological examination revealed multifocal areas of hepatocellular necrosis and hepatocyte cord dissociation (Figure 2C and 2D).

In the respiratory system, the trachea and bronchi showed a pale-yellow, mucoid secretion (Figure 1D) and solid parenchyma of firm and congestive consistency. An encapsulated structure in the cranial region of the left lung was identified as a speci-

men of the ingested eel after the incision. The eel was removed, and the specie was confirmed to be *M. ocellatus* (Figure 1C and 1E).

The histopathological examination of the lung showed fibrin and numerous leukocytes, mainly macrophages (Figure 3A and 3B), in the alveolus and bronchioles.

The tissue that surrounded the specimen in the left lung was predominantly vascularised fibrous tissue (Figure 3C and 3D).

In the heart, the right atrium was dilated and flaccid with a thickened atrioventricular valve. No apparent macroscopic changes were observed in the left atrium or ventricle (Figure 1F). The other organs showed no noteworthy macro- or microscopic changes.

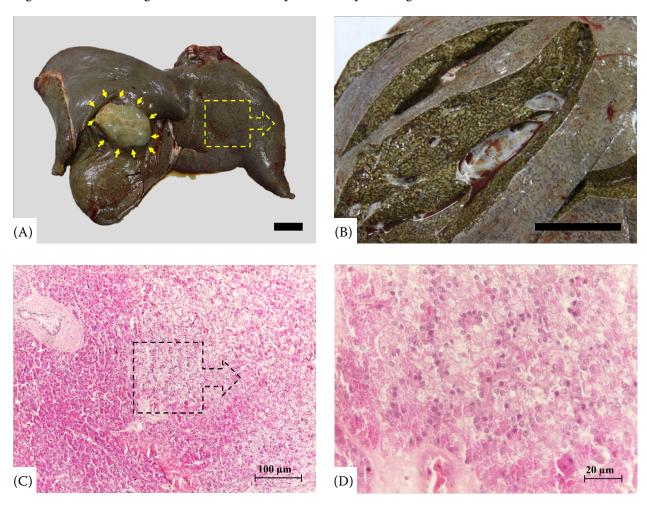


Figure 2. Histopathological findings of the liver of the loggerhead seaturtle (*Caretta caretta*) that died from ingesting a live eel (*M. ocellatus*). (A) Enlarged liver with rounded, thickened edges and a 10.0-cm diameter abscess with caseous content (arrows). Bar = 5.0 cm. (B) Nutmeg-coloured liver parenchyma. Bar = 5.0 cm. (C) Multifocal areas of hepatocellular necrosis. H&E; bar = 100  $\mu$ m. (D) Zoomed Figure 2C showing the derangement of the cellular architecture resulting from hepatic necrosis. H&E; bar = 20  $\mu$ m H&E = haematoxylin and eosin

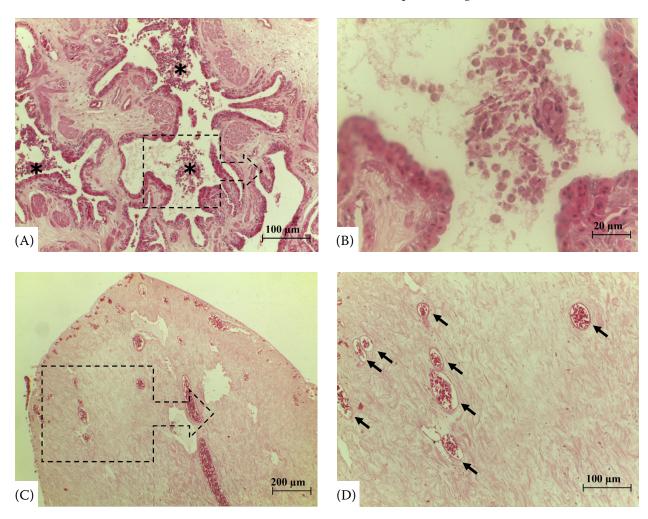


Figure 3. Histopathological findings of the lung of a loggerhead sea turtle lung (*C. caretta*) that died from ingesting a live eel (*M. ocellatus*). (A) Presence of an intra-bronchiolar fibrin-leukocyte exudate (\*). H&E; bar = 100  $\mu$ m. (B) Zoomed Figure 3A showing numerous macrophages and dendritic cells and abundant fibrin accumulation. H&E; bar = 20  $\mu$ m. (C) Fibrovascular tissue that was involved in the eel specimen in the left lung. H&E; bar = 200  $\mu$ m. (D) Zoomed Figure 3C showing a fibrous tissue with blood vessels (arrows). H&E; bar = 100  $\mu$ m H&E = haematoxylin and eosin

## **DISCUSSION AND CONCLUSIONS**

Some studies on the feeding ecology of loggerhead sea turtles reported that the diet of these animals mainly consisted of fish, followed by pelagic tunicates, crustaceans, molluscs, and other invertebrates. Loggerhead sea turtles are often opportunistic feeders, mainly eating discarded fish; the prey quantity and diversity increases proportionally with the turtle size (Tomas et al. 2001; Revelles et al. 2007).

Goldspotted eels (*M. ocellatus*) are distributed throughout the Atlantic Ocean, from Bermuda to southern Brazil (McCosker et al. 2015). They are commonly found in north-eastern Brazil at depths

of up to 30 m (Carvalho-Filho 1999). Despite being a prevalent species in the feeding areas of loggerhead sea turtles, any literature regarding eels being the feeding preference of these turtles or causing death is lacking.

In documented cases, gastrointestinal perforation in loggerhead turtles was associated with solid waste ingestion or the presence of hooks due to the incidental capture during fishing activities (Horau et al. 2014).

A study by Caracappa et al. (2018) on the incidental capture of *C. caretta* along the coast of Sicily reported that 129 of these loggerhead sea turtles had fishing hooks indifferent parts of the gastrointestinal tract, which were mostly found in the oesopha-

gus (47.3%), intestine (24.8%), stomach (14.7%), and oral cavity (13.2%).

Mucosal lesions in the gastroenteric region lead to ischemia, necrosis, and perforation (Jerdy et al. 2017), possibly resulting in secondary bacterial infections and death, as in a case study reporting the bacterium *Enterococcus faecalis* in the urinary bladder, brain, intestine, kidney, liver, lung, and muscle of a loggerhead seaturtle with oesophageal perforation caused by a fishing hook and the intestinal intussusception caused by a nylon thread. In the present case, lesions in the oesophagus, intestine, or both were the probable routes of entry for this bacterium, and their presence in the organs and viscera suggested septicaemia as the cause of death (Fichi et al. 2016). The same mechanism could be underlying in the present case: The route of entry was the gastric perforation worsened by the presence of the eel in the lung. The necropsy findings in the lungs, such as an intra-bronchiolar fibrin-leukocyte exudate and cellular response, were consistent with a bacterial infection; other findings characteristic of septicaemia, such as hepatocellular necrosis and a thickened right atrioventricular valve (endocarditis) were also observed.

These findings revealed that the loggerhead sea turtle had ingested a live eel (*M. ocellatus*) that caused the gastric perforation, possibly displacing the eel into the celomatic cavity with its cranial portion in the left lung, and leading to sepsis and the consequent death of the turtle.

## Acknowledgement

We thank Petrobras (Petróleo Brasileiro S.A.) for the financial support, the Coordination for the Improvement of Higher Education Personnel (CAPES, Financial Code 001) for the PhD scholarship granted to the Postgraduate Program in Animal Science (PPGA-UFERSA), and the entire team of veterinarians and biologists of the Cetáceos da Costa Branca Project (PCCB-UERN/CEMAM) working in the Potiguar Bay Monitoring Program, who helped rescue the animal and perform its necropsy.

#### REFERENCES

Burke VJ, Standora EA, Morreale SJ. Diet of juvenile Kemp's ridley and loggerhead sea turtle from Long Island, New York. Copeia. 1993 Dec 28;1993(4):1176-80.

Caracappa S, Persichetti MF, Piazza A, Caracappa G, Gentile A, Marineo S, Crucitti D, Arculeo M. Incidental catch of loggerhead sea turtles (Caretta caretta) along the Sicilian coasts by longline fishery. PeerJ. 2018 Aug 7;6:e5392. Carvalho-Filho A. Fishes: Brazilian coast. São Paulo, SP: Melro; 1999. 320 p.

Casale P, Tucker AD. Caretta caretta (amended version of 2015 assessment). The IUCN Red List of Threatened Species [Internet]. 2017 [cited Mar 20, 2020). Available from www.iucnredlist.org/species/3897/119333622.

Fichi G, Cardeti G, Cersini A, Mancusi C, Guarducci M, Di Guardo G, Terracciano G. Bacterial and viral pathogens detected in sea turtles stranded along the coast of Tuscany, Italy. Vet Microbiol. 2016 Mar 15;185:56-61.

Hoarau L, Ainley L, Jean C, Ciccione S. Ingestion and defecation of marine debris by loggerhead sea turtles, Caretta caretta, from by-catches in the South-West Indian Ocean. Mar Pollut Bull. 2014 Jul 15;84(1-2):90-6.

Jerdy H, Werneck MR, da Silva MA, Ribeiro RB, Bianchi M, Shimoda E, de Carvalho EC. Pathologies of the digestive system caused by marine debris in Chelonia mydas. Mar Pollut Bull. 2017 Mar 15;116(1-2):192-5.

McCosker J. Myrichthys ocellatus. The IUCN Red List of Threatened Species [Internet]. 2015 [cited Mar 20, 2020]. Available from www.iucnredlist.org/species/190440/1952002.

Prophet EB, Mills B, Arrington JB, Sobin LH. Laboratory methods in histotechnology. Washington, DC: Armed Forces Institute of Pathology; 1992. 279 p.

Revelles M, Cardona L, Aguilar A, Fernandez G. The diet of pelagic loggerhead sea turtles (Caretta caretta) off the Balearic archipelago (western Mediterranean): Relevance of long-line baits. J Mar Biol Assoc. 2007 May 16;87(3): 805-13.

Tomas J, Aznar FJ, Raga JA. Feeding ecology of the loggerhead turtle Caretta caretta in the western Mediterranean. J Zool. 2001 Nov 26;255(4):525-32.

Wyneken J. The anatomy of a sea turtle. Miami, FL: NOAA Technical Memorandum NMFS-SEFSC-470; 2004.

Received: April 9, 2020 Accepted: July 16, 2020

#### **Conflict of interest**

The authors declare no conflict of interest.